# nature portfolio

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### **Reporting Summary**

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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| For         | all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.  |
|-------------|--|
| n/a         | Confirmed  |
|             | The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement  |
|             | A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly  |
|             | The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.   |
| $\boxtimes$ | A description of all covariates tested   |
|             | A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons  |
|             | A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
|             | For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>                        |
| $\boxtimes$ | For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings   |
| $\boxtimes$ | For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes   |
|             | Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated   |
|             | Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.  |

### Software and code

Policy information about availability of computer code

Data collection

No custom software was used for data collection

Data analysis

No custom algorithms or software packages were used.

Publicly available software packages used in the analysis (software name, source, and version number) are specified in the manuscript's

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

#### Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

All genomic and transcriptomic data used in this study are available on NCBI through BioProject accession number PRJNA1129834. –Supporting statistical analysis and supplementary data files (S1: genome assembly metrics, S2: pangenome annotations, S3: virulence associated genes in A. fischeri, S4: select BGC information,

| , ,  |  | lts, S6: chemical profiles, S7: in vitro (macrophage) experimental results, S8: RNA quality and RNAseq differential expression y Fungal Metabolites) are available on FigShare (https://doi.org/10.6084/m9.figshare.25316452). |  |  |  |
|--|--|--|--|--|--|
| Research inv   | olving hui   | man participants, their data, or biological material   |  |  |  |
| Policy information a and sexual orientat                                     |  | vith human participants or human data. See also policy information about sex, gender (identity/presentation), thnicity and racism.   |  |  |  |
| Reporting on sex   | sex and gender Our current studies do not involve human research   |  |  |  |  |
| Reporting on race, ethnicity, or other socially relevant groupings           |  | Our current studies do not involve human research  |  |  |  |
| Population characteristics Our current studies do not involve human research |  | Our current studies do not involve human research  |  |  |  |
| Recruitment  |  | Our current studies do not involve human research  |  |  |  |
| Ethics oversight   |  | Our current studies do not involve human research  |  |  |  |
| Note that full informa   | tion on the appro  | oval of the study protocol must also be provided in the manuscript.  |  |  |  |
| Field-spe  |  |  |  |  |  |
|  | ne below that is<br>—  | the best fit for your research. If you are not sure, read the appropriate sections before making your selection.   |  |  |  |
| Life sciences  | ☐ Be   | ehavioural & social sciences   |  |  |  |
| For a reference copy of t  | he document with a   | all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>  |  |  |  |
| Life scier   | nces stu   | udy design   |  |  |  |
| All studies must dis   | close on these   | points even when the disclosure is negative.   |  |  |  |
| Sample size  | The sample sizes used in this study were determined based upon the cost, time, or convenience of collecting the data, and the need for it to offer sufficient statistical power. The rationale for determining the sample size is based on routine experiments where at least three samples are used for microbial or growth or metabolic activity while sample size for cell lines and animal experiments was between 6 and 10 samples. |  |  |  |  |
| Data exclusions  | No data were excluded from the analysis  |  |  |  |  |
| Replication  | All the experiments in this manuscript were repeated at least twice. When feasible, phenotyping assays were repeated in triplicate. All of the transcriptomic and metabolomic assays were repeated in triplicate.  |  |  |  |  |
| Randomization  | '  | ents were randomized either by the positions of plates/flasks in incubators (i.e. for microbial growth, metabolic activity, and profiling) or in the distribution of cages in the cage incubator                               |  |  |  |

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Although all the experiments were randomized, blinding was not relevant for interpreting the results of our study. We are not dealing with

| Materials & experimental systems |                               | Methods     |                        |  |
|----------------------------------|-------------------------------|-------------|------------------------|--|
| n/a                              | Involved in the study         | n/a         | Involved in the study  |  |
| $\boxtimes$                      | Antibodies                    | $\boxtimes$ | ChIP-seq               |  |
|                                  | Eukaryotic cell lines         | $\boxtimes$ | Flow cytometry         |  |
|                                  | Palaeontology and archaeology | $\boxtimes$ | MRI-based neuroimaging |  |
|                                  | Animals and other organisms   |             |                        |  |
| $\boxtimes$                      | Clinical data                 |             |                        |  |
| $\boxtimes$                      | Dual use research of concern  |             |                        |  |
| $\times$                         | Plants                        |             |                        |  |

clinical trials or experiments that have to be blinded.

Blinding

### Eukaryotic cell lines

Policy information about cell lines and Sex and Gender in Research

Cell line source(s)

A549 (Banco de Células do Rio de Janeiro, https://bcrj.org.br/)

Authentication The cell line used was acquired and authenticated by the cell bank in Rio de Janeiro, Brazil (https://bcri.org.br/)

The cell line was tested for mycoplasm contamination by using the methodology described by Young L, Sung J, Stacey G, Masters JR. Detection of Mycoplasma in cell cultures. Nat Protoc. 2010;5:929–34. The absence of mycoplasm contamination was verified by a PCR test.

Commonly misidentified lines (See ICLAC register)

Mycoplasma contamination

There are no misidentified cell lines in our study. Our the cell line tested negative for mycoplasma contamination.

### Palaeontology and Archaeology

Specimen provenance

Provide provenance information for specimens and describe permits that were obtained for the work (including the name of the issuing authority, the date of issue, and any identifying information). Permits should encompass collection and, where applicable, export.

Specimen deposition

Indicate where the specimens have been deposited to permit free access by other researchers.

Dating methods

If new dates are provided, describe how they were obtained (e.g. collection, storage, sample pretreatment and measurement), where they were obtained (i.e. lab name), the calibration program and the protocol for quality assurance OR state that no new dates are provided.

Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

Ethics oversight

Identify the organization(s) that approved or provided guidance on the study protocol, OR state that no ethical approval or guidance was required and explain why not.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

### Animals and other research organisms

Policy information about <u>studies involving animals</u>; <u>ARRIVE guidelines</u> recommended for reporting animal research, and <u>Sex and Gender in Research</u>

Laboratory animals

Wild-type BALB/c mice, eight to nine weeks old females. Cages are well ventilated, softly lit and subjected to a 12:12 light-dark cycle. The relative humidity was kept at 40 to 60%. Mouse rooms and cages were kept at a temperature range of 22oC.

Wild animals

This study did not involve wild animals

Reporting on sex

Sex was visually assigned.

Field-collected samples

The study did not involve samples collected from the field

Ethics oversight

The principles that guide our studies are based on the Declaration of Animal Rights ratified by UNESCO on January 27, 1978 in its 8th and 14th articles. All protocols adopted in this study were approved by the local ethics committee for animal experiments from the University of São Paulo, Campus of Ribeirão Preto (Permit Number: 08.1.1277.53.6; Studies on the interaction of Aspergillus fumigatus with animals). Groups of five animals were housed in individually ventilated cages and were cared for in strict accordance with the principles outlined by the Brazilian College of Animal Experimentation (COBEA) and Guiding Principles for Research Involving Animals and Human Beings, American Physiological Society. All efforts were made to minimize suffering. Animals were clinically monitored at least twice daily and humanely sacrificed if moribund (defined by lethargy, dyspnea, hypothermia and weight loss). All stressed animals were sacrificed by cervical dislocation.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

### **Plants**

Seed stocks

Report on the source of all seed stocks or other plant material used. If applicable, state the seed stock centre and catalogue number. If plant specimens were collected from the field, describe the collection location, date and sampling procedures.

Novel plant genotypes

Describe the methods by which all novel plant genotypes were produced. This includes those generated by transgenic approaches, gene editing, chemical/radiation-based mutagenesis and hybridization. For transgenic lines, describe the transformation method, the number of independent lines analyzed and the generation upon which experiments were performed. For gene-edited lines, describe the editor used, the endogenous sequence targeted for editing, the targeting guide RNA sequence (if applicable) and how the editor was applied.

Describe any authentication procedures for each seed stock used or novel genotype generated. Describe any experiments used to

Authentication

Describe any authentication procedures for each seed stock used or novel genotype generated. Describe any experiments used to assess the effect of a mutation and, where applicable, how potential secondary effects (e.g. second site T-DNA insertions, mosiacism, off-target gene editing) were examined.