

Current Status of Clinical Trials Design and Outcomes in Retinal Gene Therapy

Boris Rosin,¹ Eyal Banin,² and Jose-Alain Sahel^{1,2,3}

¹The UPMC Vision Institute, University of Pittsburgh, Pittsburgh, Pennsylvania 15219, USA

²Division of Ophthalmology, Hadassah-Hebrew University Medical Center, Jerusalem 91120, Israel

³Institut Hospitalo-Universitaire FOrESeIGHT, Paris 75012, France

Correspondence: brosin@pitt.edu



With the rapid expansion of methods encompassed by the term gene therapy, new trials exploring the safety and efficacy of these methods are initiated more frequently. As a result, important questions arise pertaining the design of these trials and patient participation. One of the most important aspects of any clinical trial is the ability to measure the trial's outcome in a manner that will reflect the effect of the treatment and allow its quantification, whether the trial is aimed at preservation or restoration of retinal cells (photoreceptors and others), vision, or both. Here we will review the existing methods for quantification of trial outcomes, stressing the importance of assessing the participant's visual function and not just visual acuity. We will also describe the key considerations in trial design. Finally, as patient safety remains the primary concern in any trial participation, we will outline the key principles in that regard.

The FDA approval of voretigene neparvovec rzy1 (AAV2-hRPE65v2), a subretinal injection of a viral vector for the treatment of Leber's congenital amaurosis (LCA) consecutive to mutations in the *RPE65* gene (LCA2; NCT00999609), ushered in a new era of gene therapy trials and, more broadly, vision/visual function restoration trials. With well over 250 known genes causing inherited retinal dystrophies (IRDs) (Pontikos et al. 2020), the potential for novel treatments using gene therapy opened a very large perspective. Gene-replacement techniques continue to evolve and improve. Adjustments to existing technique include methods to enhance mitochondrial function (Chadderton et al. 2023), employed in mitochondrial DNA

mutations such as Leber's hereditary optic neuropathy (LHON) (Newman et al. 2021). Interestingly, such approaches need not be limited to gene therapy, as much simpler interventions (e.g., increasing the intraocular pressure as a means of induction of ocular stress) were shown to enhance the transfer of gene therapy for LHON (McGrady et al. 2023). However, gene therapy is by no means limited to the classic gene-replacement approach, exemplified by voretigene neparvovec rzy1, as the term encompasses many different techniques.

Gene-editing techniques, such as CRISPR-Cas9, have shown great initial promise in in vitro studies (Zhang et al. 2021) and are now being tested in clinical trials in human patients affect-

Editors: Eyal Banin, Jean Bennett, Jacque L. Duncan, Botond Roska, and José-Alain Sahel
Additional Perspectives on Retinal Disorders: Genetic Approaches to Diagnosis and Treatment available at www.perspectivesinmedicine.org

Copyright © 2024 Cold Spring Harbor Laboratory Press; all rights reserved; doi: 10.1101/cshperspect.a041301
Cite this article as *Cold Spring Harb Perspect Med* 2024;14:a041301

B. Rosin et al.

ed by IRDs (NCT03872479). In a subtype of IRDs characterized by a gain-of-function mutation, gene replacement needs to be paired with gene-silencing techniques, some of which act at the RNA level and are designed to control/interfere with the expression of an aberrant protein causing the disorder (Orlans et al. 2021). Gene augmentation, the change of function of existing cells and genes to achieve a therapeutic goal, is another novel approach to gene therapy (Xi et al. 2022). While the approaches described above require surviving target cells, either photoreceptors or ganglion cells, for success, optogenetics, a novel gene therapy approach to the treatment of IRDs, can be used even in cases of widespread photoreceptor loss. The introduction of opsins into the surviving inner retinal cells by means of gene therapy in essence converts them into photoreceptors, with initial safety studies showing great promise (Sahel et al. 2021a) and clinical trials continuing (NCT03326336). Finally, employing gene therapy for introduction of trophic factors known to increase the survival of photoreceptors (e.g., rod-derived cone viability factor [RdCVF]) was shown to promote protection of photoreceptors in murine models of cone-rod dystrophy (Byrne et al. 2015), leading to ongoing phase Ib-IIa trials in patients (NCT05748873).

Unfortunately, far from all gene therapy trials culminate in success, as was the case with the first X-linked *RPGM* gene therapy trial (NCT03116113), despite showing promise in safety trials and some post hoc analyses (von Krusenstiern et al. 2023). Similarly, an RNA-based approach for the treatment of LCA10, while initially showing promise in safety trials (Russell et al. 2022), failed to reach the predefined end points in the subsequent phase 3 trial (NCT03913143). However, such “failures” may reflect flaws in the design of trials. Thus, the need for improving trial design as well as the methods for estimation of outcomes, such as vision/visual function and retinal structure, is self-evident.

The above is not meant to represent a comprehensive review of gene therapy approaches and trials, but rather to impress upon the reader the current scope of the field. This includes in particular the marked differences between the approaches encompassed by the term “gene

therapy,” as well as the variability of outcomes, even in treatments initially showing great promise. We have not even mentioned nongene therapy approaches, such as cell therapy and retinal prostheses, for instance.

METHODS FOR OUTCOME ESTIMATION

Visual Acuity and Contrast Sensitivity

While visual acuity is the main method for estimation of visual system function in the clinical setting, and it remains a main component of every attempt to estimate the outcomes of a therapeutic intervention, its usefulness in clinical trials for the treatment of IRDs is somewhat limited. In many of the patients considered for IRD treatment trials, visual acuity is rudimentary. Such is the case in LCA, where poor vision from birth is one of the defining features of the disorder described by Leber in 1869 (Leber 1869). In many cases, ascertaining poor visual acuity is an important parameter for patient inclusion in the study, as patients with preserved visual acuity could potentially lose some of that acuity in an unsuccessful trial. With gene-replacement attempts aimed toward preservation of remaining cells and not their restoration, one does not expect visual acuity to change significantly in a case of a successful intervention. Nevertheless, in trials targeting macular dystrophies and optic nerve disorders, visual acuity may provide compelling data.

Estimation of visual acuity could be performed by the classic ETDRS (Ferris et al. 1982) or the Bailey–Lovie letter charts (Bailey and Lovie 1976). Contrast sensitivity, classically measured by the Pelli–Robson charts (Pelli et al. 1988), could also be estimated by more novel technological approaches (Pelli and Bex 2013). Interestingly, studies have shown that contrast sensitivity (along with visual field) affects functional vision, as estimated by mobility performance, more significantly than visual acuity (Marron and Bailey 1982). In children and non-verbal adults, both visual acuity and contrast sensitivity could be estimated by objective measures employing vision-dependent reflexes, such as the optokinetic nystagmus (OKN) (Hyon et al. 2010), albeit the relationship between these

Clinical Trials Design and Outcomes

parameters is complex (Çetinkaya et al. 2008). Of note, OKN responses are the main method for estimation of visual acuity in laboratory animals, and many commercial systems employing the OKN exist (Prusky et al. 2004).

In patients with low vision, the measurement of vision off-chart is less accurate and various attempts to provide reliable quantitative measurements have been put forth (see collections.lib.utah.edu/ark:/87278/s6768n5w) (Schulze-Bonsel et al. 2006; Karanjia et al. 2016).

An alternative approach used in estimation of functional vision in low-vision patients is the use of self-reporting questionnaires, which incorporate the patient's self-assessment of their performance of tasks of daily living. These include the Veterans Affairs Low-Vision Visual Functioning Questionnaire (Stelmack et al. 2004) and the National Eye Institute Visual Function Questionnaire (Mangione et al. 2001), among others. Patient-reported outcomes are becoming increasingly important in this respect (Lacy et al. 2021).

Advanced Psychophysics

Electrophysiology

Since its original description by Ragnar Granit in 1933 (Granit 1933), the full-field electroretinogram (FFERG) has been the mainstay of electrophysiological testing of retinal function in generalized retinal dystrophies. In a similar manner, since the almost simultaneous description of Adrian and Matthews in 1934 (Adrian and Matthews 1934), the visual-evoked potentials (VEPs) test has been used largely for the estimation of function of the visual pathways and during the pre-magnetic resonance imaging (MRI) era was in fact considered to be an important testing modality for the diagnosis of multiple sclerosis presenting with optic neuritis (Halliday et al. 1972). Notably, the VEP response is predominated by macular function and can thus serve as a measure of interventions aimed at the macula (Holder 2004). Multifocal ERG (MFERG) is another useful tool to estimate macular function and offer localization of retinal defects (Hoffmann et al. 2021). In pattern VEP, stimuli are used to corre-

late optic pathways function to visual acuity and checkered pattern VEP has been used to that aim for many years (Harter and White 1968; Regan 1973). Pattern ERG is another method (Riggs et al. 1964) employed for estimation of both retina and optic pathways function (Berninger et al. 1988). The International Society for Clinical Electrophysiology of Vision (ISCEV) defines standards for the reporting of electrophysiological recordings and their analysis (Hamilton et al. 2021; Robson et al. 2022).

Color Vision

Farnsworth hue testing (Kinnear and Sahraie 2002) and the Ishihara plates (Ishihara 1972) have been the mainstay of color vision testing and provide a good estimation of macular function and by extension of the optic nerve (Ménage et al. 1993), where the macular fibers have a much more prominent representation than the fibers originating in the peripheral retina (Holder 2004). Of these, the Farnsworth–Munsell D-15 is perhaps the most useful in the setting of IRDs, where it has been successfully used to estimate cone function (Okajima et al. 1982; Wissinger et al. 2008). This is especially true since the Ishihara and other color vision tests require significantly preserved levels of visual acuity, as opposed to the hue recognition-based tests (Ng and Shih 2017).

Dark Adaptation

Dark adaptation testing is another useful modality for the estimation of retinal function. In particular, the longer latency rod dark adaptation curve is useful in quantifying rod photoreceptor function (Roman et al. 2005), whereas the absence of the rod-cone break is a useful tool in the estimation of cone dysfunction in achromatopsia (Aboshiha et al. 2014), which has recently emerged as the target for gene therapy in several trials (NCT02935517, NCT02599922).

Visual Fields

Visual fields are progressively constricted in many of the IRDs, causing significant dysfunc-

B. Rosin et al.

tion and impacting the patients' quality of life (Sugawara et al. 2009). The use of the static Humphrey visual fields, the mainstay of clinical practice in the management of glaucoma, is limited in the evaluation and management of IRDs. However, the 10-2 field has been used in more advanced disease with only a central island of viable photoreceptors remaining (Sayo et al. 2017). Nevertheless, kinetic perimetry has traditionally been the modality of choice for the evaluation of visual fields in IRD patients (Berson et al. 1985). In the past two decades, the classic Goldmann kinetic visual field test has been gradually replaced by the Octopus semiautomatic kinetic visual field, and studies have found the two testing modalities to be comparable (Barnes et al. 2019). Recently, the static GATE (German adaptive threshold estimation) protocol has been used more frequently in the evaluation of visual fields of patients participating in clinical trials (Schiefer et al. 2009; Buckley et al. 2022). In addition, microperimetry, somewhat better termed fundus-controlled perimetry, is slowly gaining a very central role in the management of patients of IRDs. In this testing modality, the stimuli are projected directly onto the retina with retinal image registration, allowing to control for patients' eye movements, and the test provides a localized measure of retinal sensitivity mapping (Bagdonaitė-Bejarano et al. 2019). As such, this approach allows to map out the preferential retinal locus (PRL) of each individual patient (Schönbach et al. 2022), which is of utmost importance for clinical trials aimed at preservation of the remaining photoreceptors (Yang and Dunbar 2021) as well as the planning of interventions better served by avoiding the PRL, such as in retinal prostheses implantation (Palanker et al. 2022).

Imaging in Relation to Psychophysics

Fundus autofluorescence (FAF) has been demonstrated to be a useful adjunct in the estimation of retinal function, as an abnormal fluorescence area in FAF negatively correlates with the extent of the patients' visual field (Oishi et al. 2013). With the considerable advancements of ocular coherence tomography

(OCT) technologies, the OCT analysis of IRD patients has taken the leap beyond the mere volumetric changes, which still provide significant amounts of useful information (Oh et al. 2020). OCT angiography is used to demonstrate angiographic changes in IRD patients (Cabral et al. 2020) and even monitor progression (Jauregui et al. 2018). Furthermore, due to the high resolution of imaging achieved by OCT, techniques to image the substructures of the photoreceptors have proven to be useful in ascertaining their viability. Thus, in an ongoing study, a specialized algorithm looking at OCTs of patients with generalized IRDs demonstrated a substantial subpopulation of such patients with a measurable volume of alive but dormant cone photoreceptors, making them perhaps amenable to gene therapy and other preservation therapy approaches (Janeschitz-Kriegl et al. 2022). In addition, capitalizing on the ability of the newer OCT devices to demonstrate changes on the micrometer scale, such changes were demonstrated to occur in the initial phase of phototransduction and were found to be repeatable and quantifiable. This, in essence, introduced a new discipline bridging imaging and psychophysics, termed optoretinography (Pandiyan et al. 2020). Potential implications are profound, as one can envision incorporating functional testing into the standard OCT testing performed for many of the patients seen in a retinal clinic. Adaptive optics, an imaging technique allowing in essence resolution on the level of a single cell, in itself an important imaging modality in assessing photoreceptor health (Salmon et al. 2017), further adds to the capability of optoretinography (Cooper et al. 2020).

Visual Function versus Functional Vision

Introduction

The difference between visual acuity, defined as the ability to discern optotypes at a given distance, and visual function, defined as the ability to perform a vision-dependent task, is well established. For instance, it has long been known that visual acuity in patients with advanced age-relat-

Clinical Trials Design and Outcomes

ed macular degeneration does not predict the ability to recognize certain objects and faces, given the same contrast sensitivity levels (Alexander et al. 1988). In the recent preliminary results of IRD treatment by means of optogenetics, an algae-derived opsin, ChrimsonR, was introduced into the surviving cells of the inner retina of a subject with advanced retinitis pigmentosa (RP) and light perception (LP) vision. Following treatment, he was able to detect objects when using the specialized goggles required for the activation of ChrimsonR, while his visual acuity remained LP, in a clear separation between visual acuity and function (Sahel et al. 2021a). On the other hand, patients with RP and visual field constriction, reported a significant functional impairment using the vision-related quality of life questionnaire, despite having relatively preserved visual acuity (Lange et al. 2021). Thus, the importance of developing methods to estimate visual function independently of visual acuity is paramount. To that means, the FDA has put forth guidelines, the Investigational Device Exemption (IDE) Guidance for Retinal Prostheses, describing the essential requirements for any test aimed toward quantifying visual function in trials evaluating retinal prosthetic devices (www.fda.gov/regulatory-information/search-fda-guidance-documents/investigational-device-exemption-ide-guidance-retinal-prostheses).

Object Recognition

As described above, object recognition has recently been shown as a useful method of visual function estimation (Sahel et al. 2021a). Similar approaches include picture discrimination tests (Gulati et al. 2011) and tools designed to assess instrumental activities of daily living in low vision patients (Finger et al. 2014). While these may seem as rudimentary levels of visual function, especially compared to these of patients with measurable visual acuity, the impact on patient's lives can be dramatic. Furthermore, while IRDs are relatively rare, these disorders represent a frequent cause of blindness in both children and young adults, whereas blindness in these age groups is considered to be one of the major socioeconomic burdens on medical-relat-

ed expenditures in the developed world (Frick et al. 2007, 2010).

Navigation Tasks

Introduction

When examining the components of disability of patients with retinal dystrophies, one of the most frequently reported issues is the inability to navigate in unfamiliar environments (Prem Senthil et al. 2017). Thus, navigation tasks have quickly become the preferred method for estimation of visual function in IRD patients.

The MLMT

The multiluminance mobility test (MLMT) (see Chung et al. 2023) was the functional test used during the trials leading to the FDA approval of voretigene neparvovec rzy1 (Chung et al. 2018). In this test, the subject performs a navigational task upon a relatively small (7 × 12 feet) canvas divided into squares. The desired path is denoted by arrows of sufficient size to allow visualization by an individual with a visual acuity of 20/200. Obstacles are introduced in squares both within and outside of the desired path, either conceptually (e.g., black squares representing holes) or physically (e.g., elevated squares within the path). A randomized path is created before each trial. The speed of completion as well as accuracy are scored and combined into a single score.

The importance of the MLMT is undoubtedly in the introduction of a standardized method to assess patient mobility. However, its design had some significant drawbacks. First and foremost, the limited navigation space required patients to adjust their navigation speed to the constricted environment. Furthermore, the composition of the score, reliant only on speed and accuracy of completion and offering strict discretization (lowest score of -1 to highest score of 6), did not adequately reflect the intrinsic complexities of the mobility challenges experienced by patients with IRDs. Finally, the use of conceptual rather than real obstacles created an unrealistic environment poorly representative of day-to-day navigational tasks.

B. Rosin et al.

The Streetlab

The Streetlab is an indoor simulation environment designed to mimic the complexities of a real urban street. Its large size (30×23 feet) is further enhanced by images projected on the walls of the setup (Fig. 1). Real-time obstacles (e.g., plants, garbage bins, stepladder) are introduced. A sound system is used to introduce real street sounds into the environment.

Multiple parameters are recorded and analyzed. In addition to the accuracy and time to completion, as measured by the MLMT, time to motion initiation after the “go” signal, time to walk 12 feet in a straight line without obstacles, and other similar parameters are recorded. Furthermore, the subjects’ motion is recorded by a closed-loop video system and their gait is analyzed to include, in addition to their trajectory, the number of turns and collisions. The tasks are performed under different illumination conditions (Sahel et al. 2021b).

In addition to providing the means to create a controlled environment for visual function testing, the Streetlab offers several advantages

over the MLMT. First, it creates a realistic environment of a real-life situation, such as navigating an unfamiliar street, a situation which IRD patients consistently report as one of the most challenging aspects of their visual disability (Sugawara et al. 2009). Furthermore, it offers a multitude of parameters for the quantification of the changes in visual function, adequately representing the intricacies of all the aspects of a successful navigational task. Finally, the use of highly flexible and adaptable environment, allows the utilization of the individual patient’s “coping mechanisms” (i.e., the strategies they adapt to improve their visual function) (Authié et al. 2017; Sahel et al. 2021b).

Virtual Reality Constructs

With the use of augmented and virtual reality gaining a more central place in our lives, the use of virtual reality constructs is an intuitive approach to creation of custom navigational tasks for visual function estimation. To date, no uniform approach to the design of such tasks exists

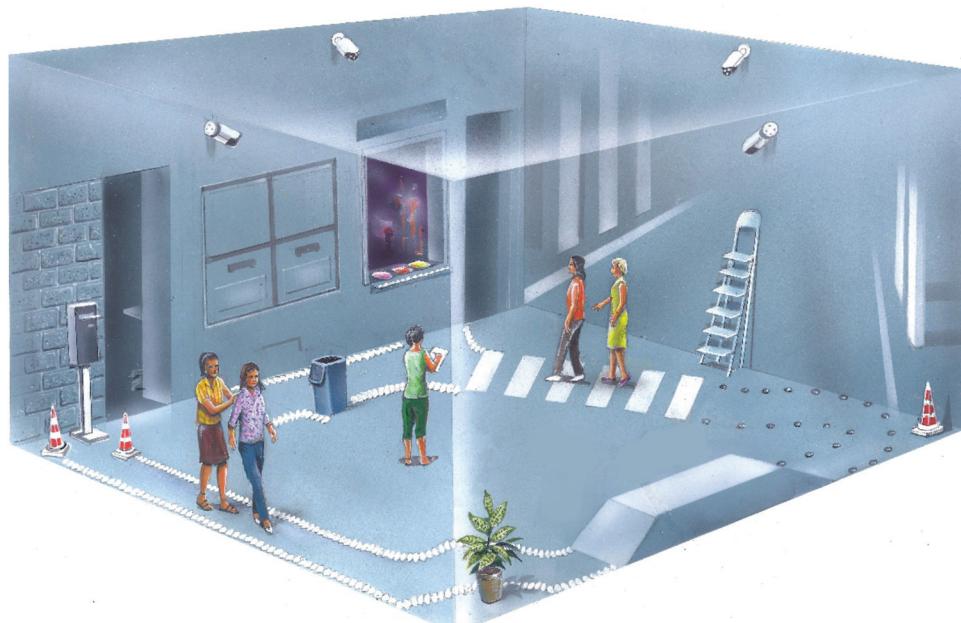


Figure 1. A schematic representation of the Streetlab. Each patient is accompanied by a mobility specialist. A central coordinator is monitoring their performance and taking notes. Video recording is stored for offline analysis of gait and movement initiation. (Illustration provided by Tuvia Kurtz.)

Clinical Trials Design and Outcomes

and standardization is required to incorporate such tests as acceptable outcome measures in both observational and interventional trials (Authié et al. 2023; Pur et al. 2023).

Considerations in the Design of Clinical Trials

Introduction

With IRDs presenting a significant cause of visual disability worldwide (Cross et al. 2022), standardization of performing clinical trials is of the utmost importance. The Second Monaciano Symposium for the Advancement of Clinical Trials for IRDs set forth recommendations for the performance of such trials (Thompson et al. 2020). While a comprehensive discussion regarding such strategies is beyond the scope of this review, we will mention here what we believe to be the most pivotal points when designing a study aimed at furthering the understanding and developing treatments of IRDs.

Observational Studies

These should ideally be performed prior to the interventional study, with the aim to adequately describe the natural history of a genetic condition. Naturally, for any study aimed at furthering the understanding of or developing a new treatment modality for IRDs, a precise genetic diagnosis of the condition is crucial. This will not only exclude nongenetic cases, but could also provide information about the significance of different mutations in disease phenotype. Notably, phenotypes tend to express variation even for the same mutation in different patients.

Ideally, an observational study should collect as much clinical information as possible regarding the IRD in question. Practically, and as shown in this review, the large amount of different testing modalities makes it impossible to perform all for each study. Out of the modalities listed above, visual acuity, contrast sensitivity, color vision, visual fields, basic imaging (fundus photos, FAF, and OCT) and a baseline FFERG are essential. The latter is also useful as an end point test, provided the baseline ERG is recordable. Additional testing modalities should

be adapted to fit the disease in question (e.g., MFERG for maculopathies vs. dark adaptation for achromatopsia). Visual function tests are very useful to quantify the resultant disability.

Interventional Studies

Perhaps the most important aspect of an interventional study is the selection of an appropriate outcome measure. Many times, no single appropriate testing modality will exist, and multiple modalities or even custom-developed testing modalities will have to be used as the outcome measures. Custom modalities will need to be standardized and validated. As shown above, functional tests are of the utmost importance for any interventional study, as they cannot only ensure benefit but also guard against possible harm.

Among the key questions to address while designing a trial are the following

- Is the expected outcome vision restoration or solely preservation? What are the patient's expectations?
- Are solid, reliable natural history data available to refine outcome measures?
- What are ways to cope with the natural variability of structural and functional parameters?
- Is efficacy better demonstrated using structural versus functional changes, or both, and over what time period?

Ethical considerations are of utmost importance for any interventional trial. We need to ensure that the patient receives all the information about the intended treatment in accessible language and is allowed enough time to reach an informed decision about participation. *Primum non nocere* should continue to be the main guiding principle for all interventions. We should strive to ensure that patients of any socioeconomic status would be able to benefit from a novel treatment in an equal manner. Finally, with a multitude of information available online, we should be prepared to provide guidance to patients seeking our advice regarding a planned trial offering them participation.

B. Rosin et al.

SUMMARY

As we enter a new and exciting era of a multitude of therapies and approaches being either developed or, in some cases, already available for IRDs, we should strive to continue to improve all aspects of clinical trials. Where possible, standardization of outcome measures should be employed. Testing modalities should be tailored to the individual patient, aiming to not only to allow better quantification of interventional outcomes, but also to ease the patient's clinical burden associated with trial participation.

REFERENCES

- *Reference is also in this subject collection.
- Aboshiha J, Luong V, Cowing J, Dubis AM, Bainbridge JW, Ali RR, Webster AR, Moore AT, Fitzke FW, Michaelides M. 2014. Dark-adaptation functions in molecularly confirmed achromatopsia and the implications for assessment in retinal therapy trials. *Invest Ophthalmol Vis Sci* **55**: 6340. doi:10.1167/iovs.14-14910
- Adrian ED, Matthews BHC. 1934. The Berger rhythm: potential changes from the occipital lobes in man. *Brain* **57**: 355–385. doi:10.1093/brain/57.4.355
- Alexander MF, Maguire MG, Lietman TM, Snyder JR, Elman MJ, Fine SL. 1988. Assessment of visual function in patients with age-related macular degeneration and low visual acuity. *Arch Ophthalmol* **106**: 1543–1547. doi:10.1001/archoph.1988.01060140711040
- Authié CN, Berthoz A, Sahel JA, Safran A. 2017. Adaptive gaze strategies for locomotion with constricted visual field. *Front Hum Neurosci* **11**: 387. doi:10.3389/fnhum.2017.00387
- Authié CN, Poujade M, Talebi A, Defer A, Zenouda A, Coen C, Mohand-Said S, Chaumet-Riffaud P, Audo I, Sahel JA. 2023. Development and validation of a novel mobility test for rod-cone dystrophies, from reality to virtual reality. *Am J Ophthalmol* doi:10.1016/j.ajo.2023.06.028
- Bagdonaitė-Bejarano L, Hansen RM, Fulton AB. 2019. Microperimetry in three inherited retinal disorders. *Semin Ophthalmol* **34**: 334–339. doi:10.1080/08820538.2019.1622025
- Bailey IL, Lovie JE. 1976. New design principles for visual acuity letter charts. *Am J Optom Physiol Opt* **53**: 740–745. doi:10.1097/00006324-197611000-00006
- Barnes CS, Schuchard RA, Birch DG, Dagnelie G, Wood L, Koenekoop RK, Bittner AK. 2019. Reliability of semiautomated kinetic perimetry (SKP) and Goldmann kinetic perimetry in children and adults with retinal dystrophies. *Transl Vis Sci Technol* **8**: 36–36. doi:10.1167/tvst.8.3.36
- Berninger TA, Arden GB, Arden GB. 1988. The pattern electroretinogram. *Eye* **2**: S257–S283. doi:10.1038/eye.1988.149
- Berson EL, Sandberg MA, Rosner B, Birch DG, Hanson AH. 1985. Natural course of retinitis pigmentosa over a three-year interval. *Am J Ophthalmol* **99**: 240–251. doi:10.1016/0002-9394(85)90351-4
- Buckley TMW, Josan AS, Taylor LJ, Jolly JK, Cehajic-Kapetanovic J, MacLaren RE. 2022. Characterizing visual fields in RPGR related retinitis pigmentosa using octopus static-automated perimetry. *Transl Vis Sci Technol* **11**: 15. doi:10.1167/tvst.11.5.15
- Byrne LC, Dalkara D, Luna G, Fisher SK, Clérin E, Sahel JA, Léveillard T, Flannery JG. 2015. Viral-mediated RdCVF and RdCVFL expression protects cone and rod photoreceptors in retinal degeneration. *J Clin Invest* **125**: 105–116. doi:10.1172/JCI65654
- Cabral D, Cascas F, Pereira T, Français C, Geraldes C, Laininhias R, Rodrigues C, Kashi AK, Nogueira V, Falcão M, et al. 2020. Quantitative optical coherence tomography angiography biomarkers in a treat-and-extend dosing regimen in neovascular age-related macular degeneration. *Transl Vis Sci Technol* **9**: 18. doi:10.1167/tvst.9.3.18
- Çetinkaya A, Oto S, Akman A, Akova YA. 2008. Relationship between optokinetic nystagmus response and recognition visual acuity. *Eye (Lond)* **22**: 77–81. doi:10.1038/sj.eye.6702529
- Chadderton N, Palfi A, Maloney DM, Carrigan M, Finnegan LK, Hanlon KS, Shortall C, O'Reilly M, Humphries P, Cassidy L, et al. 2023. Optimisation of AAV-ND1 significantly enhances its therapeutic value for correcting retinal mitochondrial dysfunction. *Pharmaceutics* **15**: 322. doi:10.3390/pharmaceutics15020322
- Chung DC, McCague S, Yu ZF, Thill S, DiStefano-Pappas J, Bennett J, Cross D, Marshall K, Wellman J, High KA. 2018. Novel mobility test to assess functional vision in patients with inherited retinal dystrophies. *Clin Exp Ophthalmol* **46**: 247–259. doi:10.1111/ceo.13022
- * Chung D, Authié C, Blouin L. 2023. Mobility testing and other performance-based assessments of functional vision in patients with inherited retinal disease. *Cold Spring Harb Perspect Med* doi: 10.1101/cshperspect.a041299
- Cooper RF, Brainard DH, Morgan JIW. 2020. Optoretinography of individual human cone photoreceptors. *Opt Express* **28**: 39326. doi:10.1364/OE.409193
- Cross N, van Steen C, Zegaoui Y, Satherley A, Angelillo L. 2022. Retinitis pigmentosa: burden of disease and current unmet needs. *Clin Ophthalmol* **16**: 1993–2010. doi:10.2147/OPTH.S365486
- Ferris FL, Kassoff A, Bresnick GH, Bailey I. 1982. New visual acuity charts for clinical research. *Am J Ophthalmol* **94**: 91–96. doi:10.1016/0002-9394(82)90197-0
- Finger RP, McSweeney SC, Deverell L, O'Hare F, Bentley SA, Luu CD, Guymer RH, Ayton LN. 2014. Developing an instrumental activities of daily living tool as part of the low vision assessment of daily activities protocol. *Invest Ophthalmol Vis Sci* **55**: 8458–8466. doi:10.1167/iovs.14-14732
- Frick KD, Gower EW, Kempen JH, Wolff JL. 2007. Economic impact of visual impairment and blindness in the United States. *Arch Ophthalmol* **125**: 544. doi:10.1001/archoph.125.4.544
- Frick KD, Kymes SM, Lee PP, Matchar DB, Pezzullo ML, Rein DB, Taylor HR; Vancouver Economic Burden of Vision Loss Group. 2010. The cost of visual impairment: purposes, perspectives, and guidance. *Invest Ophthalmol Vis Sci* **51**: 1801–1805. doi:10.1167/iovs.09-4469

Clinical Trials Design and Outcomes

- Granit R. 1933. The components of the retinal action potential in mammals and their relation to the discharge in the optic nerve. *J Physiol* **77**: 207–239. doi:10.1113/jphysiol.1933.sp002964
- Gulati R, Roche H, Thayaparan K, Hornig R, Rubin GS. 2011. The development of a picture discrimination test for people with very poor vision. *Invest Ophthalmol Vis Sci* **52**: 1197–1197
- Halliday AM, McDonald WI, Mushin J. 1972. Delayed visual evoked response in optic neuritis. *Lancet* **1**: 982–985. doi:10.1016/S0140-6736(72)91155-5
- Hamilton R, Bach M, Heinrich SP, Hoffmann MB, Odom JV, McCulloch DL, Thompson DA. 2021. ISCEV extended protocol for VEP methods of estimation of visual acuity. *Doc Ophthalmol* **142**: 17–24. doi:10.1007/s10633-020-09780-1
- Harter MR, White CT. 1968. Effects of contour sharpness and check-size on visually evoked cortical potentials. *Vision Res* **8**: 701–711. doi:10.1016/0042-6989(68)90044-8
- Hoffmann MB, Bach M, Kondo M, Li S, Walker S, Holopigian K, Viswanathan S, Robson AG. 2021. ISCEV standard for clinical multifocal electroretinography (mfERG) (2021 update). *Doc Ophthalmol* **142**: 5–16. doi:10.1007/s10633-020-09812-w
- Holder GE. 2004. Electrophysiological assessment of optic nerve disease. *Eye* **18**: 1133–1143. doi:10.1038/sj.eye.6701573
- Hyon JY, Yeo HE, Seo JM, Lee IB, Lee JH, Hwang JM. 2010. Objective measurement of distance visual acuity determined by computerized optokinetic nystagmus test. *Investig Ophthalmol Vis Sci* **51**: 752–757. doi:10.1167/iovs.09-4362
- Ishihara S. 1972. *Tests for colour-blindness*. Kanehara Shuppan, Tokyo.
- Janeschitz-Kriegel L, Calzetti G, Michaelides M, Sahel JA, Nagy Z, Stringl K, Zi-Bing J, Duncan JL, Banin E, Lam BL, et al. 2022. Worldwide multicenter ocular imaging study (EyeConic) to identify patients eligible for cone-based optogenetics therapy. *Invest Ophthalmol Vis Sci* **63**: 455–455
- Jauregui R, Park KS, Duong JK, Mahajan VB, Tsang SH. 2018. Quantitative progression of retinitis pigmentosa by optical coherence tomography angiography. *Sci Rep* **8**: 1–7. doi:10.1038/s41598-018-31488-1
- Karanja R, Hwang TJ, Chen AF, Pouw A, Tian JJ, Chu ER, Wang MY, Tran JS, Sadun AA. 2016. Correcting finger counting to Snellen acuity. *Neuroophthalmology* **40**: 219–221. doi:10.1080/01658107.2016.1209221
- Kinnear PR, Sahraie A. 2002. New Farnsworth-Munsell 100 hue test norms of normal observers for each year of age 5–22 and for age decades 30–70. *Br J Ophthalmol* **86**: 1408–1411. doi:10.1136/bjo.86.12.1408
- Lacy GD, Abalem MF, Andrews CA, Popova LT, Santos EP, Yu G, Rakine HY, Baig N, Ehrlich JR, Fahim AT, et al. 2021. The Michigan Retinal Degeneration Questionnaire: a patient-reported outcome instrument for inherited retinal degenerations. *Am J Ophthalmol* **222**: 60–68. doi:10.1016/j.ajo.2020.08.032
- Lange R, Kumagai A, Weiss S, Zaffke KB, Day S, Wicker D, Howson A, Jayasundera KT, Smolinski L, Hedlich C, et al. 2021. Vision-related quality of life in adults with severe peripheral vision loss: a qualitative interview study. *J Patient Rep Outcomes* **5**: 7. doi:10.1186/s41687-020-00281-y
- Leber T. 1869. Ueber retinitis pigmentosa und angeborene amaurose. *Arch für Ophthalmologie* **15**: 1–25. doi:10.1007/BF02721213
- Mangione CM, Lee PP, Gutierrez PR, Spritzer K, Berry S, Hays RD; National Eye Institute Visual Function Questionnaire Field Test Investigators. 2001. Development of the 25-item National Eye Institute Visual Function Questionnaire. *Arch Ophthalmol* **119**: 1050–1058. doi:10.1001/archophth.119.7.1050
- Marron JA, Bailey IL. 1982. Visual factors and orientation-mobility performance. *Optom Vis Sci* **59**: 413–426. doi:10.1097/00006324-198205000-00009
- McGrady NR, Boal AM, Risner ML, Tael M, Sahel JA, Calkins DJ. 2023. Ocular stress enhances contralateral transfer of lenadogene olparvovec gene therapy through astrocyte networks. *Mol Ther* **31**: 2005–2013. doi:10.1016/j.mthe.2023.03.035
- Ménage MJ, Papakostopoulos D, Dean Hart JC, Papakostopoulos S, Gogolitsyn Y. 1993. The Farnsworth-Munsell 100 hue test in the first episode of demyelinating optic neuritis. *Br J Ophthalmol* **77**: 68–74. doi:10.1136/bjo.77.2.68
- Newman NJ, Yu-Wai-Man P, Carelli V, Moster ML, Biousse V, Vignal-Clermont C, Sergott RC, Klopstock T, Sadun AA, Barboni P, et al. 2021. Efficacy and safety of intravitreal gene therapy for Leber hereditary optic neuropathy treated within 6 months of disease onset. *Ophthalmology* **128**: 649–660. doi:10.1016/j.ophtha.2020.12.012
- Ng JS, Shih B. 2017. Level of visual acuity necessary to avoid false-positives on the HRR and Ishihara color vision tests. *Eur J Ophthalmol* **27**: 363–366. doi:10.5301/ejo.5000855
- Oh JK, Nuzbrokh Y, Lima de Carvalho JR, Ryu J, Tsang SH. 2020. Optical coherence tomography in the evaluation of retinitis pigmentosa. *Ophthalmic Genet* **41**: 413–419. doi:10.1080/13816810.2020.1780619
- Oishi A, Ogino K, Makiyama Y, Nakagawa S, Kurimoto M, Yoshimura N. 2013. Wide-field fundus autofluorescence imaging of retinitis pigmentosa. *Ophthalmology* **120**: 1827–1834. doi:10.1016/j.ophtha.2013.01.050
- Okajima O, Tanino T, Okamoto M. 1982. Color vision defects in pigmentary retinal dystrophy. *Jpn J Ophthalmol* **26**: 292–301.
- Orlans HO, McClements ME, Barnard AR, Martinez-Fernandez de la Camara C, McLaren RE. 2021. Mirtron-mediated RNA knockdown/replacement therapy for the treatment of dominant retinitis pigmentosa. *Nat Commun* **12**: 4934. doi:10.1038/s41467-021-25204-3
- Palanker D, Le Mer Y, Mohand-Said S, Sahel JA. 2022. Simultaneous perception of prosthetic and natural vision in AMD patients. *Nat Commun* **13**: 1–6. doi:10.1038/s41467-022-28125-x
- Pandian VP, Maloney-Bertelli A, Kuchenbecker JA, Boyle KC, Ling T, Chen ZC, Park BH, Roorda A, Palanker D, Sabesan R. 2020. The optoretinogram reveals the primary steps of phototransduction in the living human eye. *Sci Adv* **6**: eabc1124. doi:10.1126/sciadv.abc1124
- Pelli DG, Bex P. 2013. Measuring contrast sensitivity. *Vision Res* **90**: 10–14. doi:10.1016/j.visres.2013.04.015

B. Rosin et al.

- Pelli DG, Robson JG, Wilkins AJ. 1988. The design of a new letter chart for measuring contrast sensitivity. *Clin Vis Sci* **2**: 187–199.
- Pontikos N, Arno G, Jurkute N, Schiff E, Ba-Abbad R, Malka S, Gimenez A, Georgiou M, Wright G, Armengol M, et al. 2020. Genetic basis of inherited retinal disease in a molecularly characterized cohort of more than 3000 families from the United Kingdom. *Ophthalmology* **127**: 1384–1394. doi:10.1016/j.ophtha.2020.04.008
- Prem Senthil M, Khadka J, Pesudovs K. 2017. Seeing through their eyes: lived experiences of people with retinitis pigmentosa. *Eye* **31**: 741–748. doi:10.1038/eye.2016.315
- Prusky GT, Alam NM, Beekman S, Douglas RM. 2004. Rapid quantification of adult and developing mouse spatial vision using a virtual optomotor system. *Invest Ophthalmol Vis Sci* **45**: 4611–4616. doi:10.1167/iovs.04-0541
- Pur DR, Lee-Wing N, Bona MD. 2023. The use of augmented reality and virtual reality for visual field expansion and visual acuity improvement in low vision rehabilitation: a systematic review. *Graefes Arch Clin Exp Ophthalmol* **261**: 1743–1755. doi:10.1007/s00417-022-05972-4
- Regan D. 1973. Rapid objective refraction using evoked brain potentials. *Invest Ophthalmol* **12**: 669–679.
- Riggs LA, Parker Johnson E, Schick AML. 1964. Electrical responses of the human eye to moving stimulus patterns. *Science* **144**: 567–567. doi:10.1126/science.144.3618.567
- Robson AG, Frishman LJ, Grigg J, Hamilton R, Jeffrey BG, Kondo M, Li S, McCulloch DL. 2022. ISCEV standard for full-field clinical electroretinography (2022 update). *Doc Ophthalmol* **144**: 165–177. doi:10.1007/s10633-022-09872-0
- Roman AJ, Schwartz SB, Aleman TS, Cideciyan AV, Chico JD, Windsor EA, Gardner LM, Ying GS, Smilko EE, Maguire MG, et al. 2005. Quantifying rod photoreceptor-mediated vision in retinal degenerations: dark-adapted thresholds as outcome measures. *Exp Eye Res* **80**: 259–272. doi:10.1016/j.exer.2004.09.008
- Russell SR, Drack AV, Cideciyan AV, Jacobson SG, Leroy BP, Van Cauwenbergh C, Ho AC, Dumitrescu AV, Han IC, Martin M, et al. 2022. Intravitreal antisense oligonucleotide sepfarsen in Leber congenital amaurosis type 10: a phase 1b/2 trial. *Nat Med* **28**: 1014–1021. doi:10.1038/s41591-022-01755-w
- Sahel JA, Boulanger-Scemama E, Pagot C, Arleo A, Galluppi F, Martel JN, Esposti SD, Delaux A, de Saint Aubert JB, de Montleau C, et al. 2021a. Partial recovery of visual function in a blind patient after optogenetic therapy. *Nat Med* **27**: 1223–1229. doi:10.1038/s41591-021-01351-4
- Sahel JA, Grieve K, Pagot C, Authié C, Mohand-Said S, Paques M, Audit I, Becker K, Chaumet-Riffaud AE, Azoulay L, et al. 2021b. Assessing photoreceptor status in retinal dystrophies: from high-resolution imaging to functional vision. *Am J Ophthalmol* **230**: 12–47. doi:10.1016/j.ajo.2021.04.013
- Salmon AE, Cooper RF, Langlo CS, Baghaie A, Dubra A, Carroll J. 2017. An automated reference frame selection (ARFS) algorithm for cone imaging with adaptive optics scanning light ophthalmoscopy. *Transl Vis Sci Technol* **6**: 9. doi:10.1167/tvst.6.2.9
- Sayo A, Ueno S, Kominami T, Nishida K, Inooka D, Nakaniishi A, Yasuda S, Okado S, Takahashi K, Matsui S, et al. 2017. Longitudinal study of visual field changes determined by Humphrey field analyzer 10-2 in patients with retinitis pigmentosa. *Sci Rep* **7**: 16383. doi:10.1038/s41598-017-16640-7
- Schiefer U, Pascual JP, Edmunds B, Feudner E, Hoffmann EM, Johnson CA, Lagrèze WA, Pfeiffer N, Sample PA, Staubach F, et al. 2009. Comparison of the new perimetric GATE strategy with conventional full-threshold and SITA standard strategies. *Invest Ophthalmol Vis Sci* **50**: 488. doi:10.1167/iovs.08-2229
- Schönbach EM, Strauss RW, Cattaneo MEGV, Fujinami K, Birch DG, Cideciyan AV, Sunness JS, Zrenner E, Sadda SR, Scholl HPN, et al. 2022. Longitudinal changes of fixation stability and location within 24 months in Stargardt disease: ProgStar Report No. 16. *Am J Ophthalmol* **233**: 78–89. doi:10.1016/j.ajo.2021.07.013
- Schulze-Bonsel K, Feltgen N, Burau H, Hansen L, Bach M. 2006. Visual acuities “hand motion” and “counting fingers” can be quantified with the Freiburg visual acuity test. *Invest Ophthalmol Vis Sci* **47**: 1236–1240. doi:10.1167/iovs.05-0981
- Stelmack JA, Szlyk JP, Stelmack TR, Demers-Turco P, Williams RT, Moran D, Massof RW. 2004. Psychometric properties of the Veterans Affairs Low-Vision Visual Functioning Questionnaire. *Invest Ophthalmol Vis Sci* **45**: 3919–3928. doi:10.1167/iovs.04-0208
- Sugawara T, Hagiwara A, Hiramatsu A, Ogata K, Mitamura Y, Yamamoto S. 2009. Relationship between peripheral visual field loss and vision-related quality of life in patients with retinitis pigmentosa. *Eye* **24**: 535–539. doi:10.1038/eye.2009.176
- Thompson DA, Iannaccone A, Ali RR, Arshavsky VY, Audi I, Bainbridge JW, Besirli CG, Birch DG, Branham KE, Cideciyan AV, et al. 2020. Advancing clinical trials for inherited retinal diseases: recommendations from the Second Monaciano Symposium. *Transl Vis Sci Technol* **9**: 2. doi:10.1167/tvst.9.7.2
- von Krusenstiern L, Liu J, Liao E, Gow JA, Chen G, Ong T, Lotery AJ, Jalil A, Lam BL, MacLaren RE, et al. 2023. Changes in retinal sensitivity associated with cotoretigene toliparvovec in X-linked retinitis pigmentosa with RPGR gene variations. *JAMA Ophthalmol* **141**: 275–283. doi:10.1001/jamaophthalmol.2022.6254
- Wissinger B, Dangel S, Jägle H, Hansen L, Baumann B, Rudolph G, Wolf C, Bonin M, Koepken K, Ladewig T, et al. 2008. Cone dystrophy with supernormal rod response is strictly associated with mutations in KCNV2. *Invest Ophthalmol Vis Sci* **49**: 751–757. doi:10.1167/iovs.07-0471
- Xi Z, Vats A, Sahel JA, Chen Y, Byrne LC. 2022. Gene augmentation prevents retinal degeneration in a CRISPR/Cas9-based mouse model of PRPF31 retinitis pigmentosa. *Nat Commun* **13**. doi:10.1038/s41467-022-35361-8
- Yang Y, Dunbar H. 2021. Clinical perspectives and trends: microperimetry as a trial endpoint in retinal disease. *Ophthalmol Int J Ophthalmol* **244**: 418–450. doi:10.1159/000515148
- Zhang X, Zhang D, Thompson JA, Chen SC, Huang Z, Jennings L, McLaren TL, Lamey TM, De Roach JN, Chen FK, et al. 2021. Gene correction of the CLN3 c.175G > A variant in patient-derived induced pluripotent stem cells prevents pathological changes in retinal organoids. *Mol Genet Genomic Med* **9**: 1601. doi:10.1002/mgg3.1601



Current Status of Clinical Trials Design and Outcomes in Retinal Gene Therapy

Boris Rosin, Eyal Banin and Jose-Alain Sahel

Cold Spring Harb Perspect Med 2024; doi: 10.1101/cshperspect.a041301 originally published online September 11, 2023

Subject Collection [Retinal Disorders: Genetic Approaches to Diagnosis and Treatment](#)

Optogenetic Vision Restoration

Volker Busskamp, Botond Roska and Jose-Alain Sahel

The Extraordinary Phenotypic and Genetic Variability of Retinal and Macular Degenerations: The Relevance to Therapeutic Developments

Isabelle Audo, Marco Nassisi, Christina Zeitz, et al.

Retinal Disorders

José-Alain Sahel, Eyal Banin, Jean Bennett, et al.

Considerations for Developing an Autologous Induced Pluripotent Stem Cell (iPSC)-Derived Retinal Pigment Epithelium (RPE) Replacement Therapy

Devika Bose, Davide Ortolan, Mitra Farnoodian, et al.

Canine and Feline Models of Inherited Retinal Diseases

Simon M. Petersen-Jones and András M. Komáromy

Immunology of Retinitis Pigmentosa and Gene Therapy–Associated Uveitis

Paul Yang, Debarshi Mustafi and Kathryn L. Pepple

Choroideremia: Toward Regulatory Approval of Retinal Gene Therapy

Imran H. Yusuf and Robert E. MacLaren

RPGR-Related Retinopathy: Clinical Features, Molecular Genetics, and Gene Replacement Therapy

Shaima Awadh Hashem, Michalis Georgiou, Robin R. Ali, et al.

Current Status of Clinical Trials Design and Outcomes in Retinal Gene Therapy

Boris Rosin, Eyal Banin and Jose-Alain Sahel

Gene Therapies for Retinitis Pigmentosa that Target Glucose Metabolism

Yunlu Xue and Constance L. Cepko

Pig Models in Retinal Research and Retinal Disease

Maureen A. McCall

Toward Retinal Organoids in High-Throughput

Stefan Erich Spirig and Magdalena Renner

Comparison of Worldwide Disease Prevalence and Genetic Prevalence of Inherited Retinal Diseases and Variant Interpretation Considerations

Mor Hanany, Sapir Shalom, Tamar Ben-Yosef, et al.

Developing New Vectors for Retinal Gene Therapy

Emilia A. Zin, Bilge E. Ozturk, Deniz Dalkara, et al.

Cell-Based Therapies: Strategies for Regeneration

Marina Pavlou and Thomas A. Reh

Neurotrophic Factors in the Treatment of Inherited Retinal Diseases

Laure Blouin, José-Alain Sahel and Daniel C. Chung

For additional articles in this collection, see <http://perspectivesinmedicine.cshlp.org/cgi/collection/>

For additional articles in this collection, see <http://perspectivesinmedicine.cshlp.org/cgi/collection/>

Copyright © 2024 Cold Spring Harbor Laboratory Press; all rights reserved