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Multi-functional coating of cellulose nanocrystals for flexible packaging applications

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Abstract:	<p>In this paper, we systematically address the performance of cellulose nanocrystals (CNs)- coated flexible food packaging films. Firstly, the morphology of CNs from cotton linters and homogeneity of its coating on different substrates were characterized by transmission electronic microscopy and atomic force microscopy. Then, the 1.5 μm thick CNs coating on polyethylene terephthalate (PET), oriented polypropylene (OPP), orientated polyamide (OPA), and cellophane films were characterized for their mechanical, optical, anti-fog, and barrier properties. CNs coating reduces the coefficient of friction while maintaining high transparency ($\sim 90\%$) and low haze (3-4%) values, and shows remarkable oxygen barrier (Oxygen coefficient permeability of CNs coating, KPO_2, $0.003 \text{ cm}^3 \text{ m}^{-2} \text{ 24h}^{-1} \text{ kPa}^{-1}$). In addition, the Gelbo flex test combined with PO_2 measurements and optical microscopy are firstly reported for evaluating the durability of coatings, revealing that the CNs-coated PET and OPA provide the best performance among the investigated coated films. CNs are therefore considered to be a promising multi-functional coating for flexible food packaging.</p>
Suggested Reviewers:	<p>Tsuguyuki Saito, Ph. D. Assistant Professor, The University of Tokyo asaitot@mail.ecc.u-tokyo.ac.jp He is an expert of TEMPO cellulose nanofibres and has plenty of publications on nano-cellulose applications including coating.</p> <p>Nathalie GONTARD, Ph. D. Professor, Université Montpellier II guillard@univ-montp2.fr She is a food packaging expert who is researching on new packaging materials with nanotechnology.</p> <p>José M. Kenny, Ph. D. Professor, University of Pergia</p>

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Cover letter

Milan, 21st Mar., 2013

Dear Editor,

I am pleased to enclose here an electronic copy of the original research manuscript entitled “Multi-functional coating of cellulose nanocrystals for flexible packaging applications” for publication on *Cellulose*.

The present manuscript describes the morphology and multiple functions of cellulose nanocrystals (CNs) which is a safe and sustainable nano-material produced from cotton linters by acid-hydrolysis method. Such thin CNs coating significantly improves the mechanical, anti-fog, and barrier properties, while maintaining excellent optical properties.

Firstly, CNs dispersion is coated on four different conventional flexible packaging materials, including polyethylene terephthalate (PET), oriented polypropylene (OPP), oriented polyamide (OPA), and cellophane (CELL). The morphology and homogeneity of CNs coating on different substrates were characterized by atomic force microscopy (AFM). Secondly, the uniform CNs coating (a) reduces the coefficient of friction (COF) resulting in less opportunity of bio-film formation; (b) maintains high transparency and low haze values so that the customers can clearly see the products inside; (c) exhibits excellent anti-fog property determined by calculated surface energy and its components from static and dynamic contact angles values; (d) performs very good oxygen barrier even after Gelbo flex test and improves the water vapour barrier. In addition, we find that the long entangled cellulose fibres are not the only crucial point for obtaining high gas barrier and that different COF and oxygen permeability are attributed to the interaction between CNs and various substrates. We conclude that a multi-functional CNs coating is promising for flexible packaging applications. Therefore, we are expecting that publishing on *Cellulose* might strongly contribute and accelerate technology transfer in this specific field.

The Authors understand the objectives of *Cellulose* and I have formatted the manuscript to fit the style and the needs of the Journal. We also understand the procedure that will be followed in the review process. I declare that the manuscript has been prepared for and sent only to *Cellulose* for publication consideration and it has not been submitted to any other Journal at this time. I attest to the fact that all Authors listed on the title page have directly participated in the planning, execution, and in the discussion of the results of this study. They also have read the manuscript, attested to the validity and legitimacy of the data and their interpretation, and agree to its submission to *Cellulose*. Also, we prefer “Free online colour”.

I hope that you consider this manuscript. If there is anything else that you would like to know, please don't hesitate to get in touch with me.

Looking forward to hearing from you soon.

Best regards,

Fei LI

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1 **Multi-functional coating of cellulose**
2 **nanocrystals for flexible packaging**
3 **applications**

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19 **Abstract**

20 In this paper, we systematically address the performance of cellulose nanocrystals (CNs)- coated
21 flexible food packaging films. Firstly, the morphology of CNs from cotton linters and
22 homogeneity of its coating on different substrates were characterized by transmission electronic
23 microscopy and atomic force microscopy. Then, the 1.5 μm thick CNs coating on polyethylene
24 terephthalate (PET), oriented polypropylene (OPP), orientated polyamide (OPA), and cellophane
25 films were characterized for their mechanical, optical, anti-fog, and barrier properties. CNs coating
26 reduces the coefficient of friction while maintaining high transparency (~90%) and low haze (3-
27 4%) values, and shows remarkable oxygen barrier (Oxygen coefficient permeability of CNs
28 coating, KPO₂, 0.003 $\text{cm}^3 \text{m}^{-2} 24\text{h}^{-1} \text{kPa}^{-1}$). In addition, the Gelbo flex test combined with PO₂
29 measurements and optical microscopy are firstly reported for evaluating the durability of coatings,
30 revealing that the CNs-coated PET and OPA provide the best performance among the investigated
31 coated films. CNs are therefore considered to be a promising multi-functional coating for flexible
32 food packaging.

33

34 *Keywords: Cellulose nanocrystals (CNs); food packaging; oxygen barrier; anti-*
35 *fog properties; bio-coating*

36 **Introduction**

37 Nowadays, the vast majority of food packaging materials is constituted of petrol-
38 based plastics, increasing the dependency of the global economy on fossil
39 resources. Therefore, considering also environmental problems, the interest in bio-
40 based materials, such as poly lactic acid (PLA) (Vert et al., 1995, Drumright et al.,
41 2000, Auras et al., 2004, Lim et al., 2008), starch (Tharanathan, 2003, Avella et
42 al., 2005) or other bio-polymers (Cha and Chinnan, 2004, No et al., 2007, Rhim
43 and Ng, 2007, Hansen and Plackett, 2008, Muzzarelli et al., 2012) has recently
44 been hugely rising. However, such materials are not yet widely applicable because
45 of their inferior properties (Ray et al., 2002, Krikorian and Pochan, 2003, Ray et
46 al., 2003) and high cost, compared with conventional ones, and still many
47 challenges exist before substituting bio-based materials for conventional plastics.

48

49 The use of plastic materials for flexible food packaging also poses a challenge in
50 finding appropriate strategies to improve their barrier properties. In current
51 research, inorganic coating, such as aluminum (Chatham, 1996, Lange and Wyser,
52 2003) and SiO₂ (Erlat et al., 1999, Haas et al., 1999, Creatore et al., 2002) or
53 nano-clays fillers (Sánchez-Valdes et al., 2006, Priolo et al., 2010, Ghasemi et al.,
54 2012, Svagan et al., 2012), are used as oxygen or water vapor barriers with
55 inevitable disadvantages that include a tendency to crack (Priolo et al., 2010) and
56 potential health risks (Lordan et al., 2011). Therefore, a sound strategy consists in
57 partially replacing conventional petrol-based plastics with bio-based materials, for
58 instance, utilizing bio-coatings with the two-fold aim of improving the original
59 plastic properties and reducing the plastic use. Bio-coatings can, therefore, be
60 considered as one of the suitable solutions for food packaging applications.

61 Nevertheless, the number of directly related bio-coating publications is still
62 limited. Gelatin (Farris et al., 2009) or pullulan (Farris et al., 2012) have been
63 recently reported as oxygen barriers on PET or OPP plastic films. Isogai and his
64 group (Kato et al., 2005) compared the oxygen barrier properties of 12 μm -thick
65 PET films coated by TEMPO-oxidized microcrystalline cellulose, chitosan and
66 starch, whilst TEMPO-oxidized nano-fiber coatings have been demonstrated as an
67 oxygen barrier on PLA and PET film (Fukuzumi et al., 2009, Fujisawa et al.,
68 2011, Rodionova et al., 2012). Besides cast coating, some research groups
69 successfully improved barrier properties of food packaging materials through
70 layer-by-layer (LbL) assembly (Jang et al., 2008, de Mesquita et al., 2010, Priolo
71 et al., 2010, Zhang and Sun, 2010, Yang et al., 2011, Svagan et al., 2012, Li et al.,
72 2013). However, at present TEMPO-oxidized and LbL coating processes are still
73 difficult to apply at the industrial scale due to their cost and process complexity.

74

75 In this work, we investigate the physical, mechanical, and optical properties of a
76 bio-coating made of cellulose nanocrystals (CNs), which can be obtained from the
77 most abundant natural polymer on Earth. Besides the promising results that are
78 discussed throughout the paper, such material brings advantages also in terms of
79 low weight, low cost, and biodegradability. Over the last few years, CNs have
80 been extracted from different original sources by chemical, physical, enzymatic
81 processes, or a combination of them (Siró and Plackett, 2010). However, physical
82 and enzymatic processes imply high cost and high energy consumption,
83 respectively, hence we chose a chemical-hydrolysis method for CNs production
84 from cotton linters and we deposited them on different conventional flexible food
85 packaging materials to produce a multi-functional coating. In particular, we used a
86 dispersion of CNs as the coating material deposited on PET, OPP, OPA and

87 cellophane films. The morphology, coefficient of friction, anti-fog, optical,
88 oxygen barrier and water vapor barrier properties of coated films were measured
89 and systematically interpreted.

90 **Materials and methods**

91 **Materials.**

92 Cotton linters were provided by S.S.C.C.P. (Milan, Italy) as the raw material to
93 produce CNs.

94 Four different plastic substrates were coated and used for experiments: (1)
95 poly(ethylene terephthalate) (PET, 12 ± 0.5 μm thickness), (2) oriented
96 polypropylene (OPP, 20 ± 0.5 μm thickness), (3) oriented polyamide (OPA, 12 ± 0.5
97 μm thickness), and cellophane (CELL, 12 ± 0.5 μm thickness). All plastic films
98 have been provided by Radici Film, San Giorgio di Nogaro, Italy.

99

100 **Methods.**

101 *CNs extraction.*

102 1 wt% Cellulose nanocrystals (CNs) dispersion was produced from cotton linter
103 by a procedure described elsewhere (Li et al., 2013). Briefly, milled cotton linters
104 were hydrolyzed by 64 wt% sulfuric acid with vigorous stirring at 45 °C for 45
105 minutes. The reaction mixture was diluted with deionized water and then rinsed
106 and centrifuged at 5000 rpm repeatedly until the supernatant became turbid.
107 Further purification was then done by dialysis against deionized water (Molecular
108 Weight Cut Off 12 000 and higher). Sequentially, the suspension was sonicated
109 (UP400S 400 W, Hielscher Co., Germany) to create cellulose crystals of colloidal
110 dimensions. Finally, the suspension was filtered under vacuum with MukteLL
111 (grade GF/C, 1.2 μm pore diameter) and Whatman glass microfiber filter (grade
112 GF/F, 0.7 μm pore diameter) to remove contamination and big aggregations. The
113 CNs content of the resulting aqueous suspension was determined by drying

114 several samples (1 ml each) at 105 °C for 15 min intervals (to avoid
115 decomposition or burning) until weight constancy, giving a cellulose
116 concentration of ~1 wt% and a yield of ~50%. To prepare a given concentration of
117 CNs solution, the resulting CNs dispersion was adjusted to pH ~7 by 1M
118 NaOH(aq), freeze-dried and stored in tightly sealed container under dry
119 conditions for later analysis and experiments.

120

121 *Particle size distribution*

122 1% CNs dispersion was scanned by a laser diffraction particle size analyzer
123 (Mastersizer 2000, Malvern Instruments), combining a blue source with 470 nm
124 wavelength and a red source with 632.8 nm wavelength.

125

126 *Preparation of Coating Dispersion.*

127 An 8 wt% CNs water dispersion was obtained by dissolving the CNs into distilled
128 water assisted with ultrasonic treatments until the dispersion became visually
129 homogenous. During the process, the sonication should be carried out every ten
130 minute in water bath to avoid overheating. After recovering to room temperature,
131 the CNs dispersion was coated on different plastic films.

132

133 *Coated Film Preparation.*

134 According to ASTM D823-07, practice C, the corona-treated sides (external sides)
135 of four different rectangular (25×20 cm²) plastic films were coated by an
136 automatic film applicator (ref 1137, Sheen Instruments, Kingston, U.K.) at a
137 constant speed of 2.5 mm s⁻¹. Water was evaporated using a constant mild air flow

138 (25±0.3 °C for 5 min) at a perpendicular distance of 40 cm from the automatic
139 applicator. The coated films were stored under controlled conditions (20±2 °C,
140 45±2.0% RH) for 24 h, and then stored in sealed anhydrous desiccators for 24 h
141 before analysis. All substrates are distinguished by external (Ex) and internal (In)
142 sides. The external side is the pre-treated part, while the internal side is without
143 corona treatments.

144

145 *Thickness measurements*

146 For the coating thickness measurement, a 10×10 cm² sample (plastic substrate
147 with coating) was cut and weighed (m_1 , g). The coating was then removed by
148 running hot water (~70 °C) and the resulting bare film was weighed (m_2 , g). The
149 coating thickness (l , μm) was obtained according to the following equation:

$$150 \quad l = \frac{m_1 - m_2}{\rho} \times 100, \quad (1)$$

151 where $\rho \sim 1.58 \text{ g cm}^{-3}$ is the density of CNs (Mazeau and Heux, 2003). Three
152 measurements were performed for each coating type.

153

154 *Microscopy*

155 Transmission Electron Microscopy (TEM)

156 Drops of aqueous dispersions of CNs (0.05 wt%) were deposited on carbon-
157 coated electron microscope grids, negatively stained with uranyl acetate and
158 allowed to dry. The samples were analyzed with a Hitachi Jeol-10084 TEM
159 operated at an accelerating voltage of 80 kV.

160

161 *Atomic force microscopy (AFM)*

162 AFM topography images have been acquired in tapping mode with a Veeco
163 Innova instrument. Super-sharp silicon probes (typical radius of curvature 2 nm)
164 have been used for high-resolution imaging of nanocrystals, while standard silicon
165 probes have been employed for large-area scans in order to evaluate the sample
166 roughness.

167 The root mean square roughness S is calculated as the standard deviation of the
168 topography

169 ($M \times N$ pixels):

170
$$S = \sqrt{\frac{1}{MN} \sum_{i=1}^M \sum_{j=1}^N |z(x_i, y_j) - \bar{z}|^2}, \quad (2)$$

171 where \bar{z} is the mean value of the topography $z(x, y)$.

172

173 *Optical microscopy*

174 Gelbo Flex treated CNs-coated samples were observed using an optical
175 microscope (Micro Nikon Eclipse ME600 Laboratory Imaging; Nikon
176 Instruments, Sesto Fiorentino, Italy) at $5\times$ and $10\times$ magnification. Pieces of film
177 ($30 \times 30 \text{ mm}^2$) were mounted on a rectangular glass sample holder and observed
178 without any pretreatment. Images were captured by NIS-Element software (Nikon
179 Instruments, Sesto Fiorentino, Italy).

180

181 *Coefficient of Friction*

182 The static (μ_s) and dynamic (μ_d) friction coefficients were measured by a
183 dynamometer (model Z005, Zwick Roell, Ulm, Germany), in accordance with the

184 standard method ASTM D 1894-87. The software TestXpert V10.11 (Zwick
185 Roell, Ulm, Germany) Master was used for data analysis.

186

187 *Optical properties*

188 Transparency measurements

189 The transmittance of the sample was measured at a wavelength of 550 nm,
190 according to the ASTM D 1746-70, by means of a spectro-photometer (model
191 L650, Perkin-Elmer, Milano, Italy).

192

193 *Haze*

194 Haze was measured in accordance with ASTM D 1003-61 by means of a
195 spectrophotometer (Perkin Elmer L650). The haze values of uncoated and coated
196 films were obtained as:

$$197 \quad \text{haze} = 100 \times \frac{I_s}{I_T}, \quad (3)$$

198 where I_s and I_T are the scattered and total transmitted light, respectively.

199

200 *Contact angle measurements*

201 Contact angles were measured to estimate the surface energies of the tested
202 substrates by OCA 15 Plus angle goniometer (Data Physics Instruments GmbH,
203 Filderstadt, Germany). The software (SCA20 and SCA21) provided by the
204 instrument manufacturer calculate the surface energy based on contact angles
205 measurements. Measurements of static and advancing contact angle were
206 performed at room temperature with two polar liquids and one apolar liquid:

207 Milli-Q water, formamide (FOM, $\geq 99.5\%$, Carlo Erba, Milano, Italy), and
 208 diiodomethane (DIM, 99%, Sigma Aldrich), respectively. Each measurement was
 209 repeated on at least five different positions for each sample. The surface energies
 210 were calculated from the contact angle data at equilibrium by the Van Oss method
 211 (van Oss, 2006), which divides the total surface free energy into two components,
 212 the dispersive and the polar components, where the polar interactions originate
 213 from the Lewis acid-base interactions:

214

$$215 \quad \gamma_i^{total} = \gamma_i^D + \gamma_i^P, \quad (4)$$

216 where

$$217 \quad \gamma_i^P = 2\sqrt{\gamma_i^+ \gamma_i^-}, \quad (5)$$

218 The subscripts i indicate the solid ($i=s$) or liquid ($i=l$) phase, and the superscripts
 219 refer to the dispersive (D) and polar (P) components of the total surface energy.
 220 γ^+ and γ^- are electron-acceptor and donor parts of the Lewis acid-base
 221 interactions.

222

223 When combined with Young's equation, the equations developed by Chaudhury,
 224 Good, and Van Oss yield the equation (van Oss, 2006)

$$225 \quad \gamma_l(1 + \cos\theta) = 2(\sqrt{\gamma_l^D \gamma_s^D} + \sqrt{\gamma_s^+ \gamma_l^-} + \sqrt{\gamma_s^- \gamma_l^+}), \quad (6)$$

226 where θ is the contact angle, γ_l is the liquid surface tension (mJ m^{-2}), and γ_s^+ ,
 227 γ_s^- and γ_l^+ , γ_l^- are electron-acceptor and donor contributions to the polar
 228 component of the solid and liquid, respectively (mJ m^{-2}). The values of the surface
 229 tension and its components for each liquid that were used in calculations were

230 determined by Van Oss (van Oss, 2003). Water, formamide, and diiodomethane,
231 with known γ_l^D , γ_l^+ , and γ_l^- values (Table 1) and measured contact angle (θ)
232 were used to determine γ_s^D , γ_s^+ , and γ_s^- combined with Equation (6). Finally,
233 γ_s^{total} and γ_s^P were calculated from Equation (4) and (5).

234

235

Table 1

236 *Oxygen and water vapor permeability (P_{O_2} and P_{H_2O}) measurements.*

237 The P_{O_2} and P_{H_2O} of CNs coated plastic films were measured by permeation
238 instruments (MOCON, OX-TRAN[®] Model 702 and PERMATRAN-W[®] Model
239 700) at 23 °C and 0% relative humidity (RH) and at 38 °C and 100% RH
240 difference, complying with ASTM D-3985, F-1927, F-1307 and ASTM F-1249,
241 respectively.

242

243 The P_{O_2} of the CNs coating in the coated film [i.e., $P_{O_2}(Coating)$] was calculated
244 using the following equation:

245
$$\frac{1}{P_{O_2}(coating)} = \frac{1}{P_{O_2}(coated\ film)} - \frac{1}{P_{O_2}(film)}, \quad (7)$$

246 where $P_{O_2}(coated\ film)$ and $P_{O_2}(film)$ are the oxygen permeabilities for the coated
247 substrate and the bare substrate, respectively.

248

249 *Gelbo Flex testing*

250 According to slightly modified ASTM F392, the CNs-coated samples were treated
251 by 20 cycles with Gelbo Flex tester (model S/N 80 03 32, VINATORU
252 ENTERPRISES INC, Graham, USA), shown in Fig. 1. Each cycle includes back

253 and forth steps with 440° on long stroke. After treatments, the samples were
254 observed by optical microscopy and the oxygen permeability was measured
255 according to the method above.

256

257 Fig. 1

258 **Results and discussion**

259 **Morphology**

260

261 Fig. 2

262

263 Fig. 3

264

265 The rod-like CNs produced by sulfuric acid hydrolysis of cotton linters were
266 characterized by TEM as shown in Fig. 2. CNs observations obtained from casted
267 diluted dispersion (approximate 0.05 wt%) show individual nanocrystals and
268 some aggregates. The appearance of aggregated elementary crystallites in TEM
269 images is expected due to the high specific area and strong hydrogen bonds
270 established between the CNs. From several TEM images, the mean values of the
271 length (L) and diameter (d) of the isolated CNs were determined to be 120 ± 30 nm
272 and 6 ± 3 nm, respectively, giving an aspect ratio (L/d) ~ 20 . Similar dimensions
273 have been reported in the literature (Angles and Dufresne, 2000, Elazzouzi-
274 Hafraoui et al., 2007). In order to better understand the CNs size distribution, the
275 probability histogram for the particle size distribution of 1% CNs dispersion is
276 shown in Fig. 3. It indicates that the range of the CNs length is from 90 to 160 nm
277 (mean value ~ 115 nm), which is highly similar with the results obtained from
278 TEM (Fig. 2) and in other references (Habibi et al., 2010). A certain degree of size
279 distribution is inevitable owing to the acid diffusion-controlled nature of the
280 hydrolysis (de Mesquita et al., 2010). However, the results demonstrate that the
281 acidic hydrolysis process is highly reliable and effective to extract relatively
282 uniform CNs from cotton linters.

283

284 Fig. 4

285

286 Fig. 4 shows high-resolution AFM images of CNs-coated films. The image shows
287 a dense packing and uniform coverage of nanofibers. We therefore conclude that a
288 new CNs layer was homogeneously established on different conventional
289 packaging materials. Dense packing of CNs has been previously described in
290 other reports (Fujisawa et al., 2011). The continuous layer of overlapping CNs
291 fibers (Siró and Plackett, 2010) points towards possible improvements of the
292 oxygen barrier properties of the different substrates, which we will demonstrate
293 and discuss later on. Moreover, we acquired large-area AFM images, as shown in
294 Fig. 5, where CNs cobble-stone pathway-like aggregations appear due to the
295 strong hydrogen bonds. All images are qualitatively very similar with each other.
296 Root-mean-square roughness values calculated from such AFM investigations all
297 lay in a low-value range (6-13 nm), which is definitely more narrow than the one
298 measured for the external side of bare films (2-21 nm, not shown). A low
299 roughness has been correlated to a less opportunity for bio film formation
300 (Shellenberger and Logan, 2001, Li and Logan, 2004, Ringus and Moraru, 2013),
301 thus bacterial fouling occurrence can be likely reduced by such a coating when
302 applied onto rough substrates.

303

304 Fig. 5

305

306

307

308

309 **Coefficient of friction (COF).**

310

311

Table 2

312

313 The values for the coefficient of friction (COF) of CNs-coated films against films
314 are presented in Table 2. By means of the statistical analysis, we again can
315 conclude that the casting deposition created a completely new CNs-coating layer
316 on different substrates with one exception. In fact, it can be clearly noted that
317 three of the investigated systems (coated-PET, OPA, and CELL) present similar
318 values in dynamic (μ_d) COFs, while only CNs-coated OPP is significantly
319 different from the other coated films and close to the bare one, probably due to
320 weak adhesion between CNs coating and OPP surface which leads to the removal
321 of CNs from substrate during dynamic measurements. The COF results and
322 oxygen permeability discussed later indicate that for thin CNs coating a
323 homogenous independent layer was established by cast coating. It is thus
324 concluded that CNs coating results in an improvement for practical applications,
325 because of the reduction of friction between the films, which might represent a
326 premium feature for high-speed packaging machineries.

327

328 **Optical properties.**

329

330

Table 3

331

332 As for optical properties, the transparency and haze values of bare films are 87-
333 92% and 2.1-3.0%, respectively, while the ones of coated films are 88-91% and

334 3.3-4.0%. Detailed results in Table 3 show that the CNs-coated films still
335 maintain high transparency, as requested to ensure easy evaluation of the product
336 quality inside the package. Although their haze values increase, the maximum
337 values (4%) are yet within an acceptable range. Overall, the thin CNs coating
338 (~1.5 μm , as determined by weighting the samples) has no significant influences
339 on the optical properties of coated films.

340

341 **Anti-fog properties.**

342 Besides the COF improvements and maintaining excellent optical properties,
343 empirical boiling water and breathe (Huff) tests both showed that the CNs-coated
344 films have excellent anti-fog properties, which are presented in Fig. 6.
345 Particularly, we deposited CNs directly on different substrates without any
346 primers or any chemicals to guarantee the safety of food contact. In Fig. 6 we
347 show the results for the coated OPP film since OPP is the most hydrophobic
348 material among the substrates under investigation, but very similar results were
349 obtained for all the substrates. We observed the border [blue dashed line in panels
350 (b) and (c)] between bare and coated films by eye inspection. In order to better
351 observe the anti-fog property, we set a black sponge in the water containers, as
352 shown in Fig. 6(b). As known, fog is formed by small discontinuous water
353 droplets that diffuse the incident light, thereby decrease the transparency and
354 increase the haze (Nuraje et al., 2010, Introzzi et al., 2012). Fig. 6 (c), obtained
355 with optical microscopy, presents two parts, an uncoated one with water droplets,
356 which forms fog, and a coated one with a homogeneous water layer without any
357 droplets, which remained transparent. In the following part, we will further

358 interpret the excellent anti-fog properties of CNs-coated films from the
359 comparison of surface energies obtained from contact angle goniometry.

360

361 Fig. 6

362

363 Table 4a

364 Table 4b

365

366 From Table 4, we could notice that the CNs-coated films show highly similar
367 static contact angles, which indicates that a completely new CNs layer was
368 established through cast coating process. The static contact angles of CNs coated
369 substrates are much lower than the ones of bare substrates, since cellulose chains
370 contain many hydroxyl (-OH) groups leading to its hydrophilicity. Meanwhile, the
371 dynamic contact angles, which might better reveal the real behavior of hydrophilic
372 surfaces, were also determined. Moreover, it was reported that the differences
373 between the advancing and receding contact angles are usually related with
374 surface's roughness and water absorption often observed on natural fibers
375 (Dankovich and Gray, 2011).

376

377 We will use contact angles combined with surface energies of CNs coating to
378 interpret the principles behind the anti-fog performance. The static and dynamic
379 contact angle values (Table 4a and b) are in good agreement with results recently
380 presented on nano-cellulose coated or casted films (Dankovich and Gray, 2011).

381 Regarding the anti-fog properties, it was reported that one possible measure of the
382 level of interaction of water with a material is the advancing contact angle and it
383 has been suggested that hydrophilic surfaces with contact angles lower than 40°

384 should exhibit anti-fog behavior (Briscoe and Galvin, 1991, Howarter and
385 Youngblood, 2008). Indeed, the measured advancing contact angle values (θ_{Water})
386 of CNs-coated films are about 26-27°.

387

388  Fig. 7

389

390 To further interpret the anti-fog behavior, we calculated the surface energies
391 values from contact angles, presented in Fig. 7, through Lifshitz-van der
392 Waals/acid-base theory. In the literature, the surface energy of cellulose
393 determined by contact angle measurements is known to be $\sim 55 \text{ mJ m}^{-2}$ (Aulin et
394 al., 2009). Also, a value for the surface energy of CNs films on different
395 substrates was reported as high as 65 mJ m^{-2} with $\pm 15\%$ experimental error
396 (Kontturi et al., 2007). These values are consistent with the values obtained in our
397 results of CNs-coated PET and OPP. Contact angles were measured by using
398 water, FOM, and DIM to provide information regarding the contributions of
399 dispersive (D) and polar (P) components to the total surface energy of the
400 substrates. The calculated values in Fig. 7(a) indicate that the dispersive part of
401 the CNs surface energy is larger than the polar contribution as well as the CNs
402 hydrophilic property, we thus further investigate the electron-acceptor and donor
403 components to the polar part of total surface energy shown in Fig. 7(b). From Fig.
404 7(b), bare PET and OPP have different asymmetric electron-acceptor and donor
405 patterns due to PET and OPP being relatively hydrophilic and hydrophobic,
406 respectively, while the CN-coated PET and OPP have similar asymmetric patterns
407 which disclose that the thin CNs coating results in an independent and
408 homogeneous layer on PET and OPP substrates. This is a common occurrence
409 since, for a given polar substrate, a strong asymmetry is usually observed between

410 the contribution of electron-acceptor and electron-donor interactions, making one
411 of the two dominant (van Oss, 2006). Furthermore, the electron-donor component
412 (γ_s^-) values of coated PET and OPP shown in Fig. 7(b) are at least 5 and 2000
413 times the bare ones, which is likely the fundamental factor for different anti-fog
414 performances between bare and coated films. Also, cellulose always presents a
415 polar and hydrophilic surface (Moon et al., 2011) and CNs are grafted with a few
416 sulfate ester groups ($-O-SO_3^-$) due to the sulfuric acid hydrolysis process.
417 These results, therefore, suggest that the anti-fog property induced on the PET and
418 OPP substrates should be attributed to the electron-donor parameter of the polar
419 component. For hydrophilic biopolymers, similar results were also reported
420 (Nuraje et al., 2010, Introzzi et al., 2012).

421

422 **Barrier properties.**

423

424 Fig. 8

425

426 In addition to excellent performances on COF, optical, and anti-fog properties, the
427 CNs barrier function is evaluated as follows. Fig. 8(a) shows that the oxygen
428 barrier properties of coated films improve significantly thanks to the thin (1.5 μm)
429 CNs coating. The coatings allow for a dramatic reduction (>99%) of oxygen
430 permeability (P_{O_2}) of all CNs-coated samples. Oxygen permeability coefficient
431 (KP_{O_2}) of CNs coating on OPA is down to $0.003 \text{ cm}^3 \mu\text{m m}^{-2} 24\text{h}^{-1} \text{ kPa}^{-1}$, which is
432 much lower than that of commercialized oxygen barrier, ethylene vinyl alcohol
433 (EVOH), under dry condition (Lee et al., 2008). Also, this KP_{O_2} value is
434 comparable with the best oxygen barrier from wood TEMPO-oxidized cellulose

435 nanofiber (TOCN-COONa) (Fukuzumi et al., 2011) and this is, according to our
436 best knowledge, the first systematic report on P_{O_2} of CNs-coated films. Such
437 excellent oxygen barrier properties achieved on CNs-coated OPA can be
438 tentatively attributed to the strong hydrogen bonds among CNs and between CNs
439 and OPA. We also calculated the KP_{O_2} of CNs coatings applied on other
440 substrates, finding always an improved barrier compared to the bare substrate.
441 Therefore, our results indicate that CNs coating provides very good oxygen
442 barrier (especially under dry conditions) but also that the effectiveness of this
443 property is related to the possible interactions between CNs and different plastic
444 films used as coating substrates. One more interesting point is the achievement of
445 high oxygen barrier using short CNs (~120 nm). In other words it seems to be
446 demonstrated that long entangled cellulose fibers (Belbekhouche et al., 2011) are
447 not the only crucial point in achieving certain functionalities.

448

449 The P_{O_2} of CNs-coated films after Gelbo Flex test is also presented in Fig. 8(a).
450 As known (Habibi et al., 2008, Isogai et al., 2011, Moon et al., 2011), nano-
451 cellulose is stiff and rigid, which should result in fragility of the coating and,
452 thereby, a possible partial loss of the gas barrier properties during usage. Based on
453 the above considerations, we compared the oxygen barrier before and after Gelbo
454 Flex tests. The results indicate that strong distortions led to some destructions of
455 CNs coating and reduce the oxygen barrier properties of all CNs-coated films.
456 Gelbo Flex treated CNs-coated PET and OPA films, however, still maintain a
457 significant low P_{O_2} . Compared with others, Gelbo Flex treated coated OPP and
458 cellophane films show relatively higher P_{O_2} , probably due to their weak adhesion,
459 which was also observed in dynamic COF tests especially for OPP. In order to
460 confirm this point, we did observations by optical microscopy and the figures are

461 presented in Fig. 9. The morphologies of PET and OPA in Fig. 9 (a), and (c)
462 reveal better adhesions between CNs and PET and OPA, since only a small
463 portion of the CNs film was peeled off from substrates even though there are
464 many chaps (crackles) especially at strongly twisted parts of CNs coatings.
465 However, in case of Gelbo Flex treated CNs-coated OPP, from Fig. 9 (b) we
466 found peeled-off/blister and cracks of CNs coating, which is likely to result in the
467 lower oxygen barrier properties observed. Peeling-off cracks also unexpectedly
468 occurred to cellophane, shown in Fig. 9 (d). This phenomenon might be explained
469 by the presence of resins coated or laminated on the commercialized cellophanes,
470 in this case, low wettable polyester resin. The images from optical microscopy are
471 therefore consistent with the lower oxygen barrier on Gelbo Flex treated coated
472 OPP and cellophane. In this work, the oxygen permeability values were measured
473 only under the dry condition. Since CNs is a hydrophilic biopolymer, the oxygen
474 barrier will be certainly reduced under high RH as other paper reported
475 (Fukuzumi et al., 2012). This limitation is common to the currently used synthetic
476 barrier polymers (polyamide (PA), polyvinyl alcohol (PVOH), ethylene vinyl
477 alcohol (EVOH)) and led to the development of multilayer structures (Li et al.,
478 2013), designed in order to protect moisture sensitive polymers with polyolefin
479 such as low-density polyethylene (LDPE), high-density polyethylene (HDPE),
480 and polypropylene (PP).

481

482 The four coated films were also evaluated for their water vapor permeability
483 (P_{H_2O}) and Fig. 8(b) reports the results obtained. The P_{H_2O} of CNs-coated PET,
484 OPP, OPA and CELL films is reduced by 22, 26, 24 and 6.5% respectively,
485 compared to bare films, while the corresponding thickness increase is 7.5% for
486 OPP and 12.5% for the other three films. Such a limited improvement of the

487 moisture barrier is due to CNs hydrophilicity (Hult et al., 2010, Sanchez-Garcia et
488 al., 2010, Moon et al., 2011) and also to the heavy conditions of the test used
489 (38 °C under a difference of 100% RH). However, the water vapor barrier
490 properties of CNs coating might be satisfactory in real-use conditions which
491 generally involve lower temperature and RH values. Also, high water vapor
492 barrier can already be achieved by conventional and convenient synthetic
493 polymers such as PP and PE.

494

495 Fig. 9

496 **Conclusions**

497 To the best of our knowledge, this is the first time that CNs are deposited on
498 different substrates as multi-functional coatings. In the literature, CNs are usually
499 considered as a filler (Noorani et al., 2007, Habibi et al., 2008, Siqueira et al.,
500 2010, Goffin et al., 2011, Dong et al., 2012) or used for small-scale spin or layer-
501 by-layer coating (Kontturi et al., 2007, Cranston and Gray, 2008, Cerclier et al.,
502 2010, Hoeger et al., 2011, Li et al., 2013), and are assumed not to function as a
503 practical coating material (Isogai et al., 2011). In this paper, we systematically
504 investigated the properties of conventional films coated with CNs, demonstrating
505 that CNs coatings mainly lead to a reduction of friction, a premium feature for
506 industrial applications, and that they do not influence significantly the optical
507 properties of coated films. Moreover, CNs coating shows excellent anti-fog
508 property, which is a strongly required performance for flexible food packaging,
509 intended to be used for fresh food. Finally, CNs coatings not only dramatically
510 improve the oxygen barrier properties of conventional flexible food packaging
511 material, but also lead to a certain reduction in the water vapor permeability.
512 While substitution of conventional plastics might still be far ahead because of
513 their low cost, large flexibility and availability, the perspective use of CNs as
514 multi-functional coatings to favor a reduction of the required thickness for plastic
515 films, towards a more environmentally-friendly and sustainable approach to
516 packaging seems promising and feasible. The best substrate to be covered with
517 CNs appeared to be PET and OPA films due to their intrinsic polarity and high
518 surface energies; however, the possible use of activation treatments and other
519 plastic substrates remain to be deeply investigated, as well as the possible

520 measures to reduce the moisture sensitivities and mechanical rigidity of the thin
521 nano-cellulose coatings.

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527 particle size distribution measurement.

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696

697 **Figure Caption**

698 Fig. 1 Gelbo Flex tester, a, b, and c indicate different phases of a back and forth
699 cycle.

700 Fig. 2 TEM image of individual CNs

701 Fig. 3 Probability histogram for the particle size distribution measured from 1%
702 CNs dispersion

703 Fig. 4 High-resolution AFM images of CNs-coated PET (a), OPP (b), OPA (c),
704 and CELL (d).

705 Fig. 5 AFM for roughness of coated substrates, coated PET (a), OPP (b), OPA (c),
706 and cellophane (d).

707 Fig. 6 Boiling water test; panel (a) the foggy bare OPP observed with an optical
708 microscope; panel (b) comparison between bare and CNs-coated OPP during the
709 test; panel (c) the border between CNs-coated and bare parts observed with an
710 optical microscope.

711 Fig. 7 Surface energies and their respective polar and dispersive components of
712 bare and CNs-coated PET and OPP, (a); the electron-acceptor and donor of polar
713 component, (b)

714 Fig. 8 The oxygen permeability (P_{O_2}) of bare, CNs-coated, and CNs-coated after
715 Gelbo Flex tests PET, OPP, OPA, and CELL films at 23 °C under dry condition
716 (a); the water vapor permeability of bare and CNs-coated PET, OPP, OPA, and
717 CELL films at 38 °C under 100% RH difference (b).

718 Fig. 9 The morphology of Gelbo Flex treated CNs-coated PET (a), OPP (b), OPA
719 (c), and CELL (d) films by optical microscopy.

720 **Tables**

721 Table 1 Surface tension components and parameters of the liquids used in direct
722 contact angle determination in mJ m^{-2} , at 20 °C (van Oss et al., 2001, vanOss,
723 2003)

724

725 Table 2 The coefficient of friction (COF) of Plastic films (Ex) against plastic film
726 (In)

727

728 Table 3 Transparency at 550 nm and haze of bare and CNs-coated films

729

730 Table 4a Static contact angles of bare and CNs-coated films

731 Table 4b Advancing and receding contact angles of bare and CNs-coated films

732

Figure1
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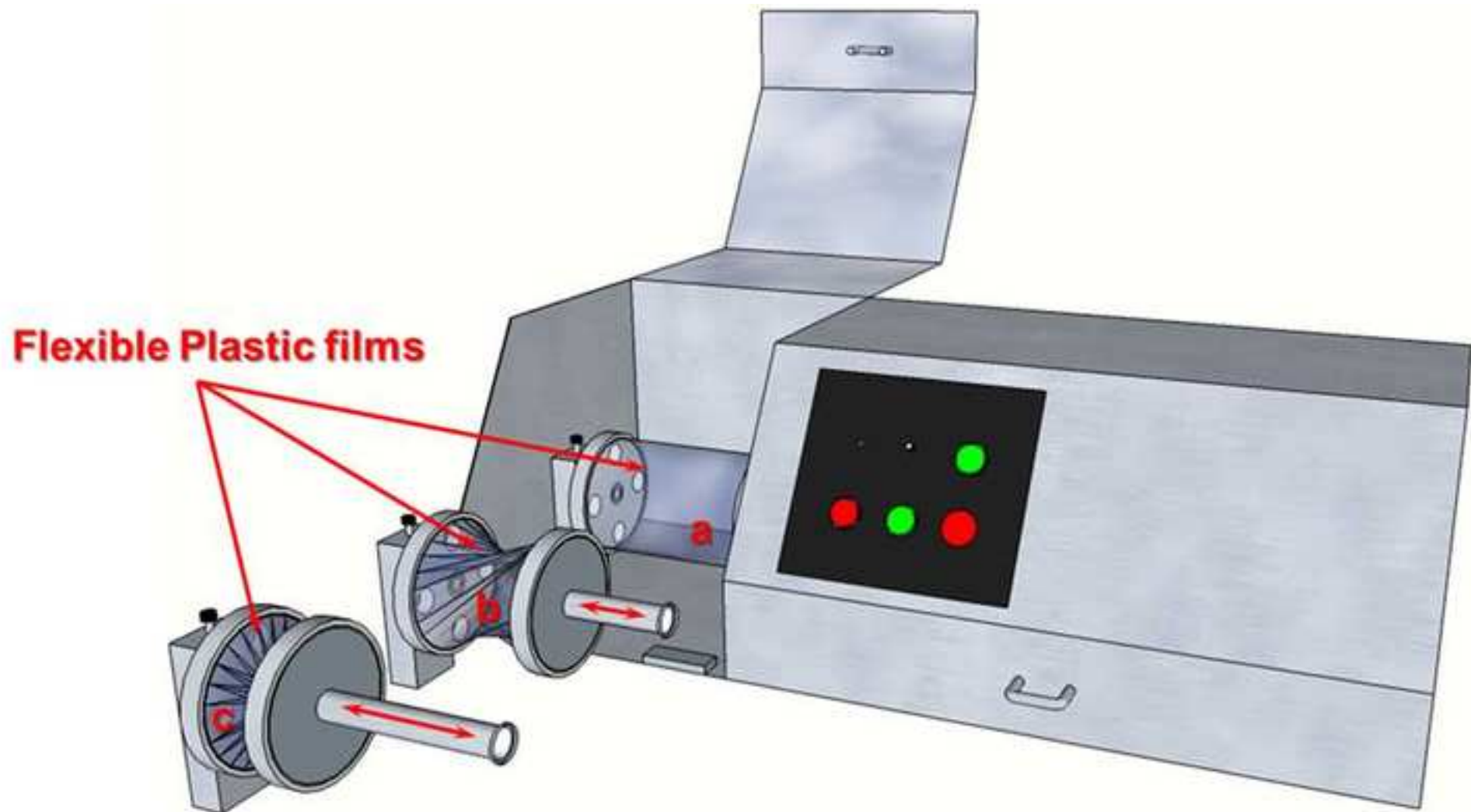


Figure2

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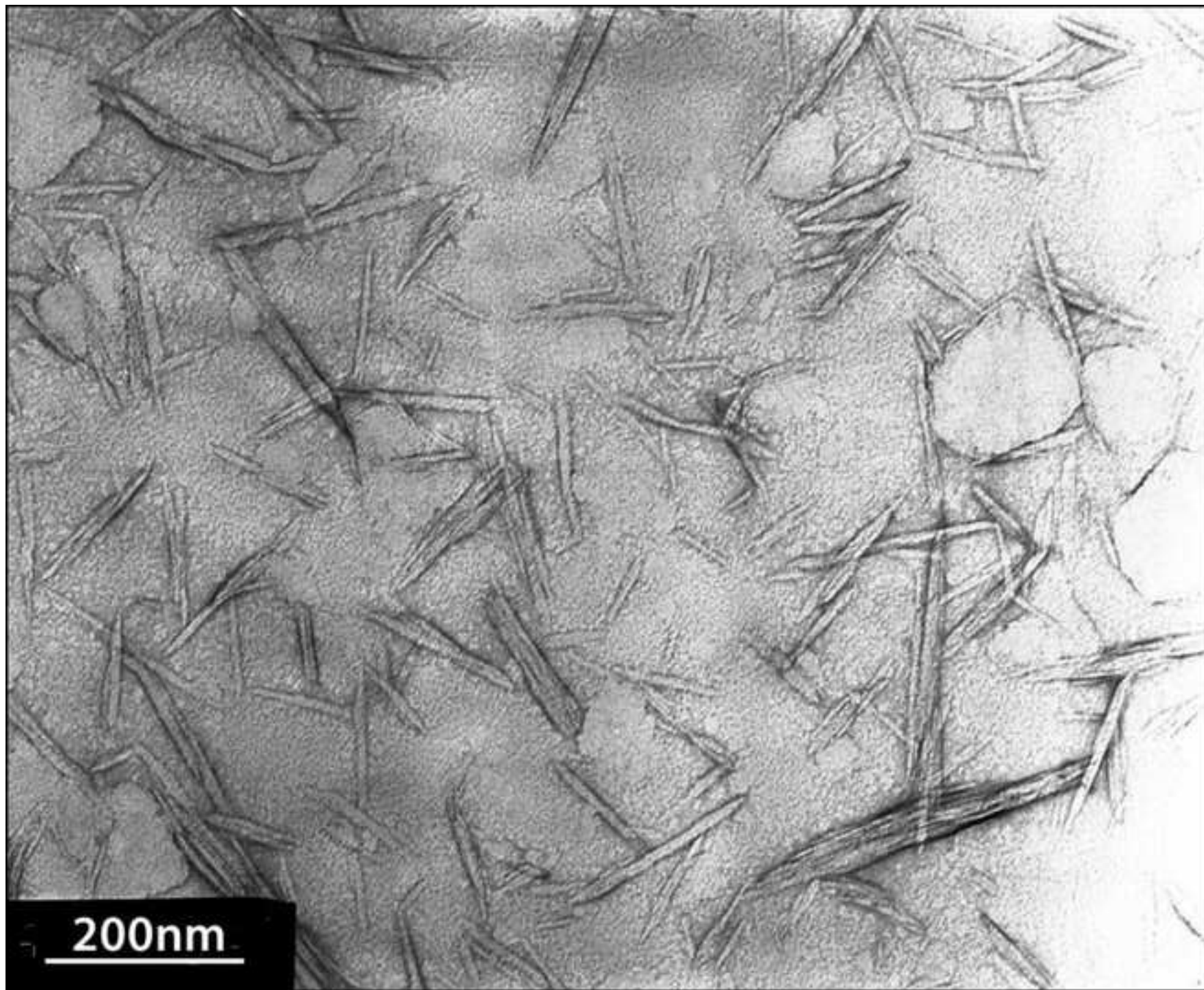


Figure3
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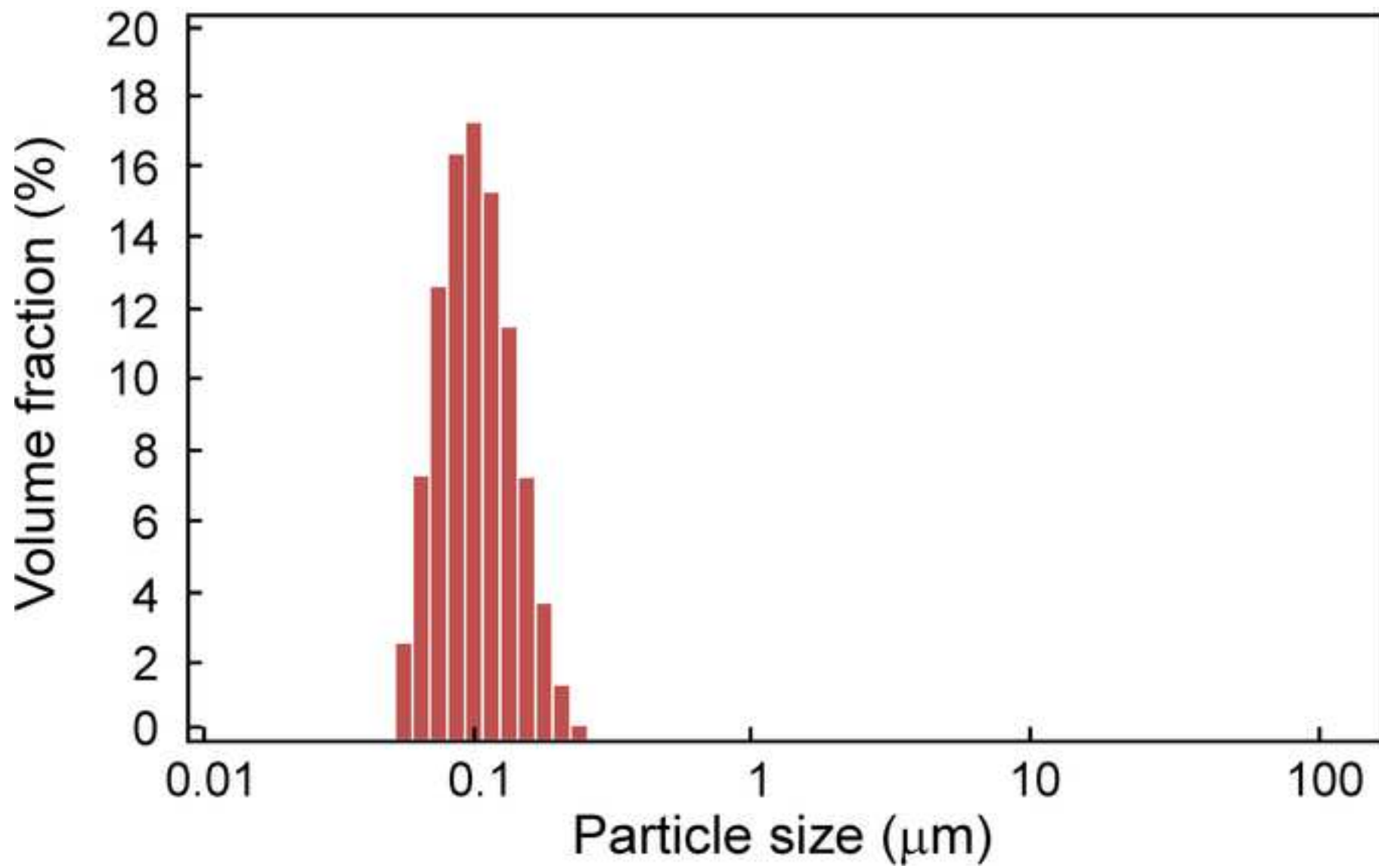


Figure4
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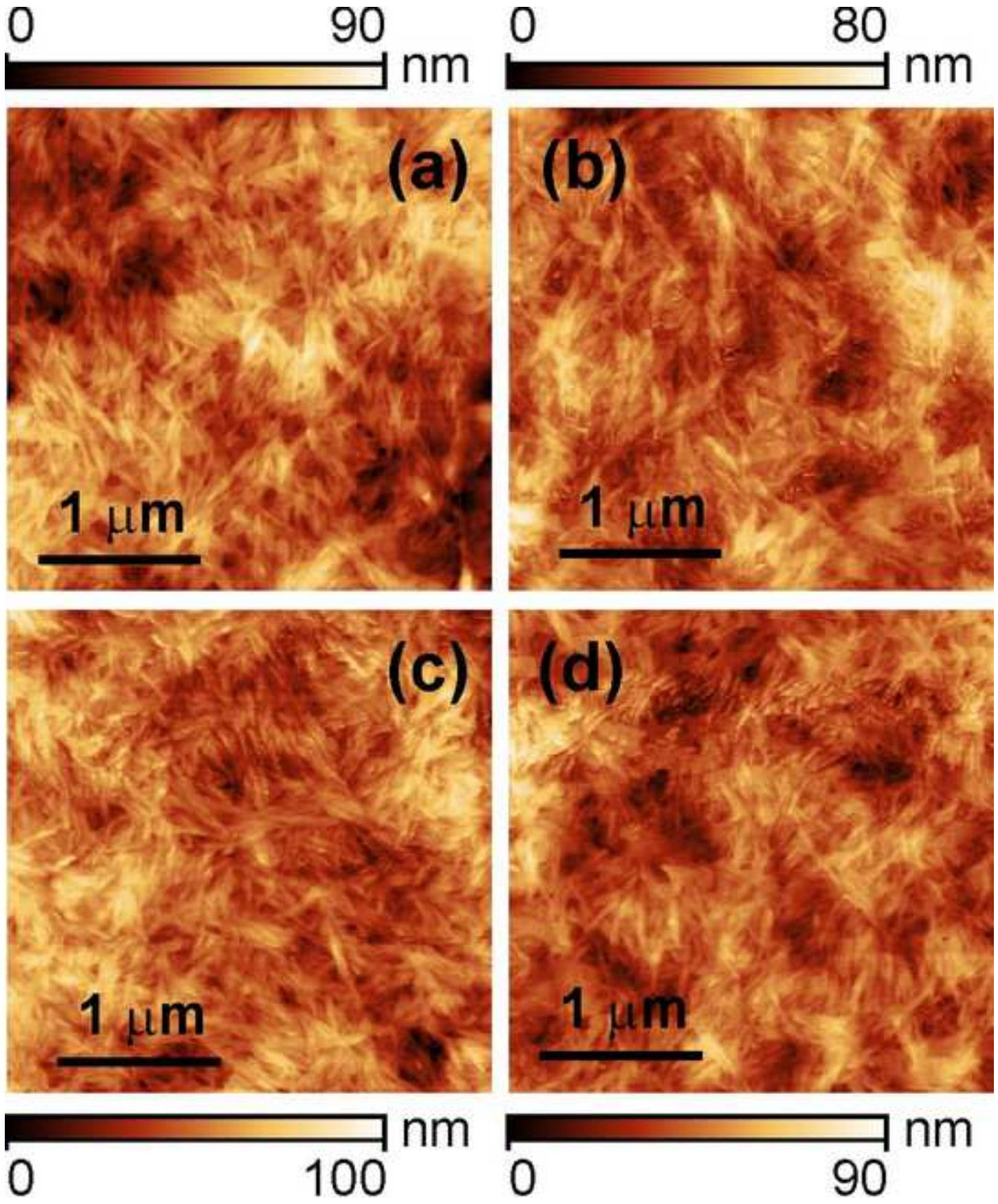


Figure5
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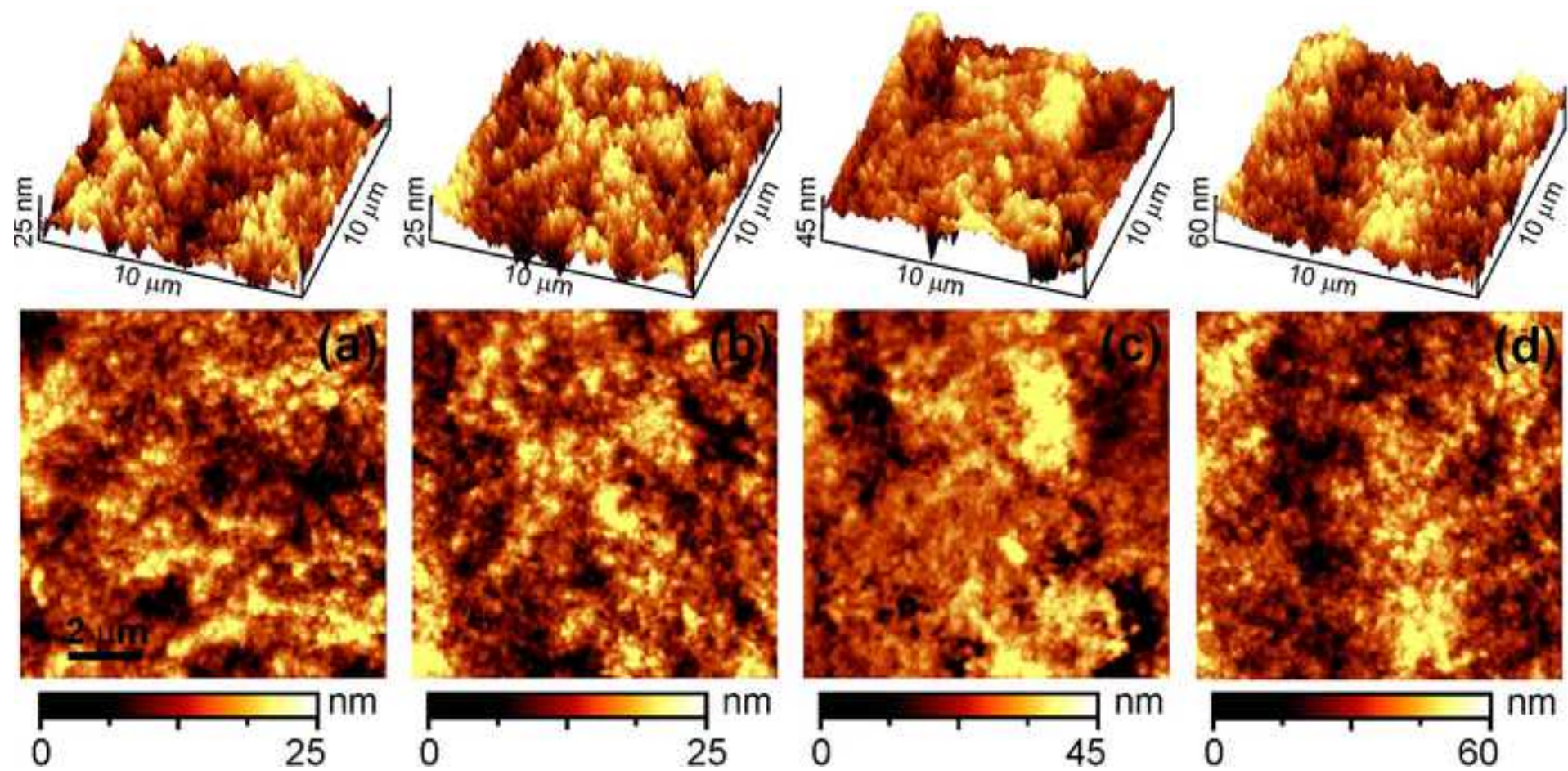
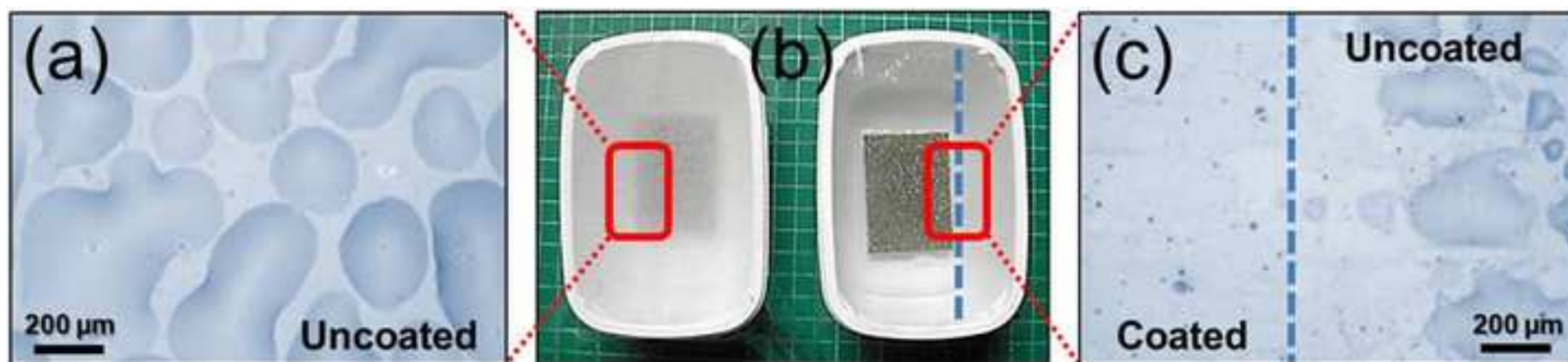
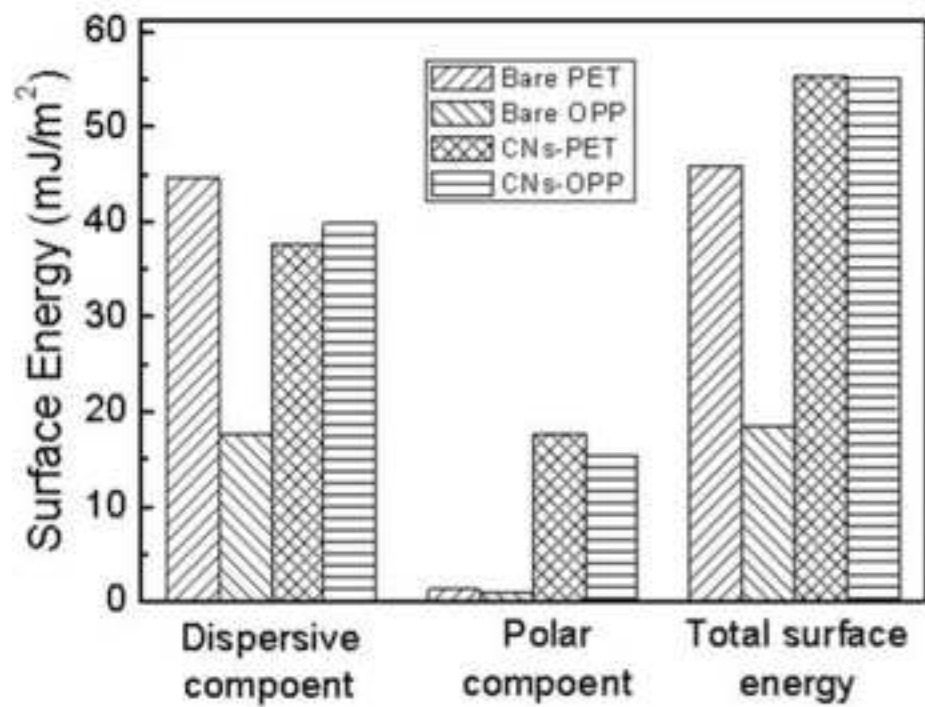
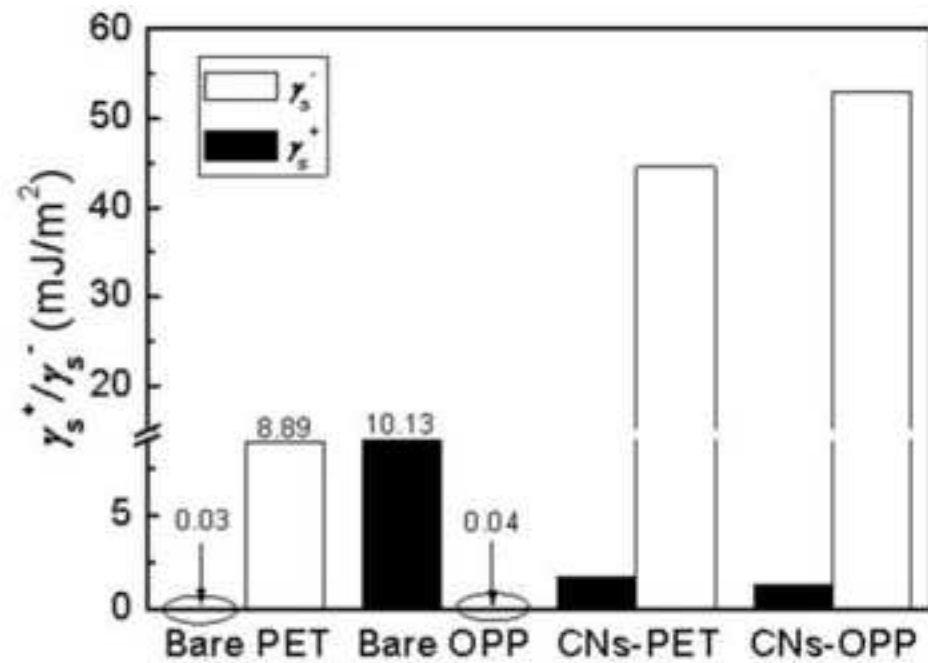


Figure6
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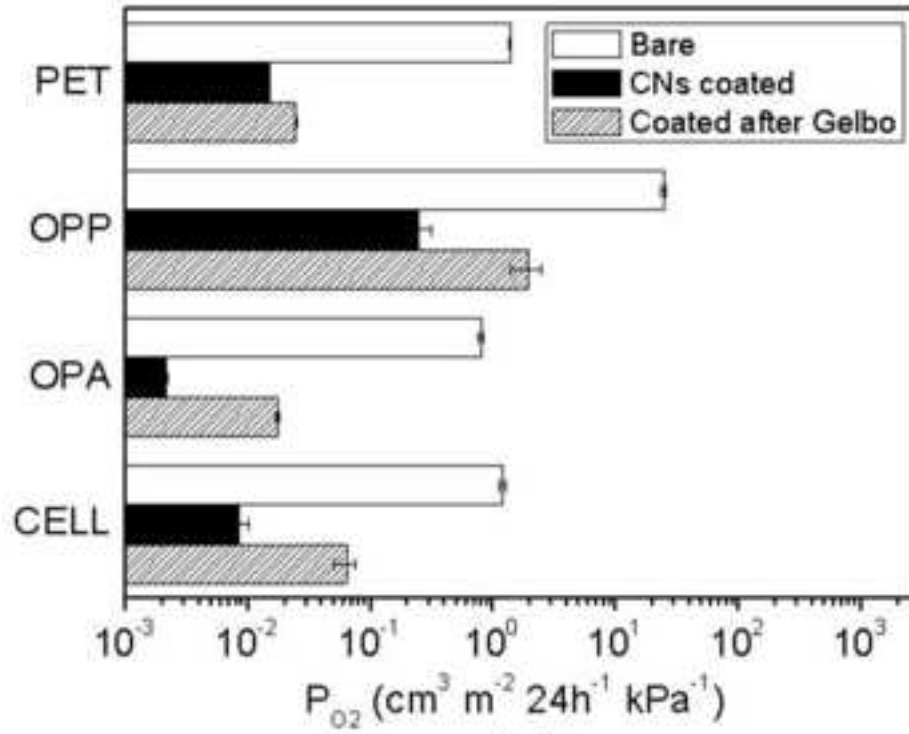
(a)



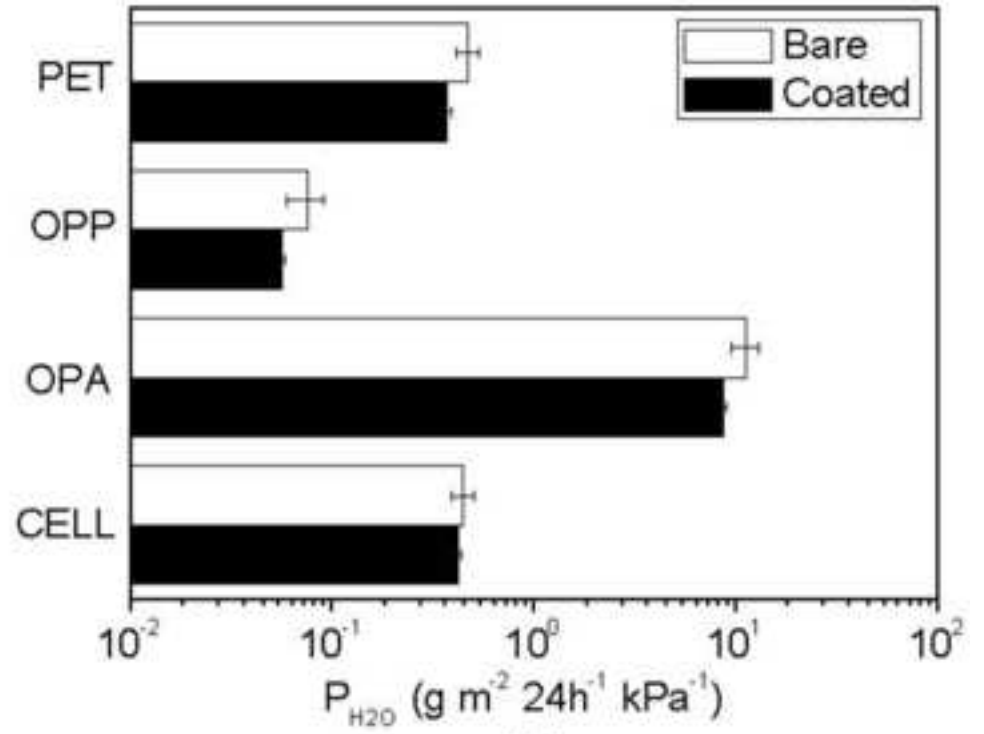
(b)

Figure8

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(a)



(b)

Figure9
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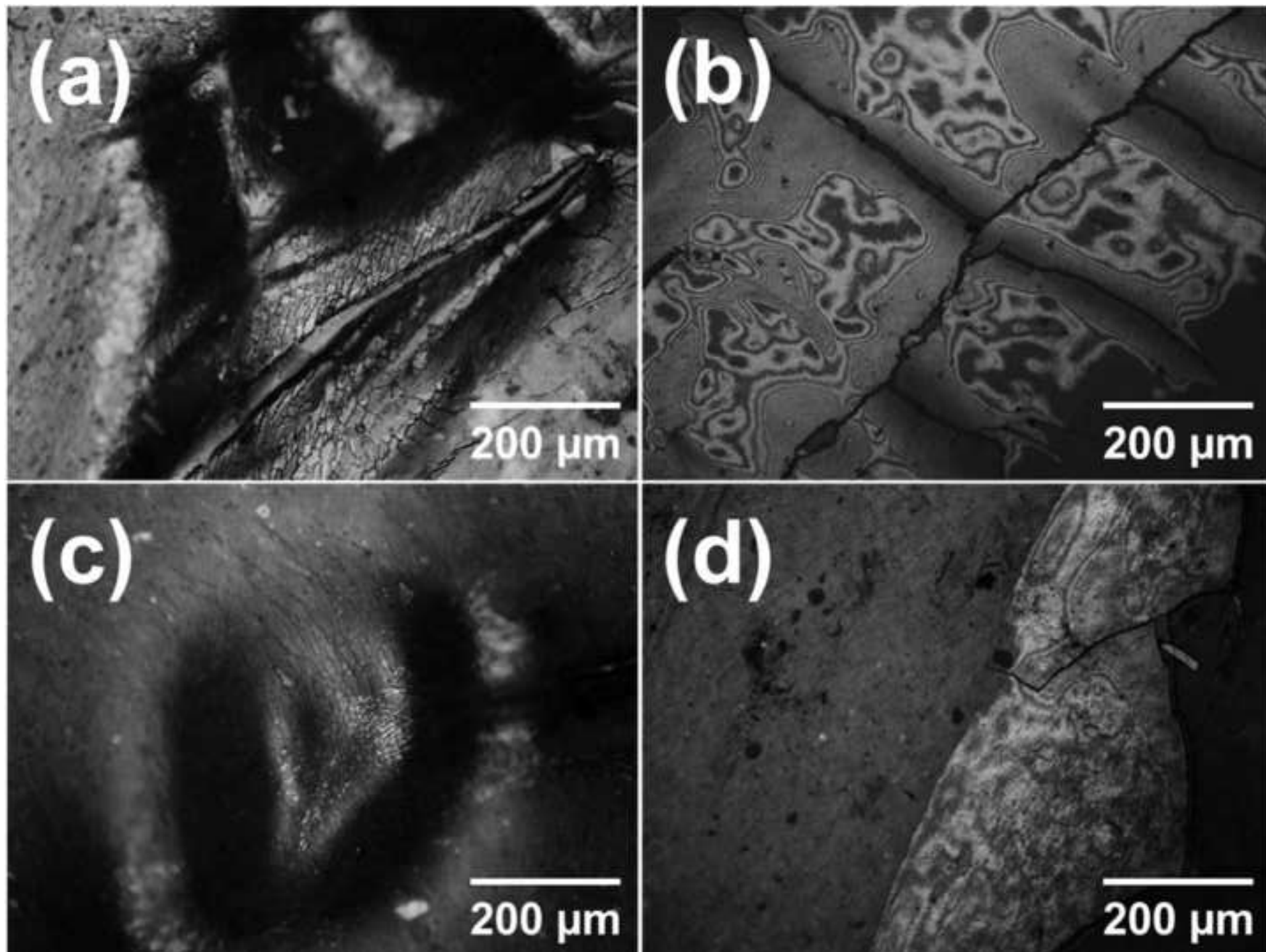


Table 1 Surface tension components and parameters of the liquids used in direct contact angle determination in mJ m^{-2} , at 20 °C (van Oss et al., 2001, van Oss, 2003)

Liquid	γ_t	γ_t^D	γ_t^P	γ_t^+	γ_t^-
APOLAR					
Diiodomethane	50.8	50.8	0	≈ 0.01	0
POLAR					
Water	72.8	21.8	51.0	25.5	25.5
Formamide	58.0	39.0	19.0	2.28	39.6

Table 2 The coefficient of friction (COF) of Plastic films (Ex) against plastic film (In)

Substrate	Bare (Ex)		Coated (600 nm ^a)	
	μ_s	μ_d	μ_s	μ_d
PET	0.57 ± 0.02^D	0.52 ± 0.03^H	0.38 ± 0.01^C	0.33 ± 0.01^G
OPP	0.18 ± 0.01^A	0.17 ± 0.00^F	0.30 ± 0.02^B	0.18 ± 0.01^F
OPA	0.79 ± 0.02^E	0.74 ± 0.03^J	0.32 ± 0.01^{BC}	0.32 ± 0.02^G
CELL	0.62 ± 0.03^D	0.57 ± 0.01^I	0.38 ± 0.02^C	0.34 ± 0.01^G

^a Thickness of CNs coating

^A to ^E, different letters mean that static COFs are significantly different ($p < 0.01$);

^F to ^J, different letters mean that dynamic COFs are significantly different ($p < 0.01$).

Table 3 Transparency at 550 nm and haze of bare and CNs-coated films

Substrates	Transparency (%)		Haze (%)	
	Bare	CNs-coated	Bare	CNs-coated
PET	87.5±0.3	89.0±0.3	2.9±0.1	3.3±0.2
OPP	91.8±0.0	90.8±0.2	2.1±0.0	3.4±0.3
OPA	90.2±0.5	89.8±0.0	3.0±0.1	4.0±0.2
Cellophane	87.3±0.1	88.3±0.1	3.2±0.1	3.8±0.2

Table 4a Static contact angles of bare and CNs-coated films

Films	Static CA ^a		
	$\theta_{\text{Water}}(^{\circ})$	$\theta_{\text{DIM}}(^{\circ})^{\text{b}}$	$\theta_{\text{FOM}}(^{\circ})^{\text{c}}$
Bare PET	57.44±5.84	22.41±2.73	39.55±1.77
Bare OPP	63.03±1.00	52.22±1.61	41.39±1.70
CNs-PET	12.32±1.33	37.15±3.02	7.41±1.33
CNs-OPP	12.08±0.95	36.37±1.70	8.78±0.73

Table 4b Advancing and receding contact angles of bare and CNs-coated films

Films	Advancing CA			Receding CA		
	$\theta_{\text{Water}}(^{\circ})$	$\theta_{\text{DIM}}(^{\circ})^{\text{b}}$	$\theta_{\text{FOM}}(^{\circ})^{\text{c}}$	$\theta_{\text{Water}}(^{\circ})$	$\theta_{\text{DIM}}(^{\circ})^{\text{b}}$	$\theta_{\text{FOM}}(^{\circ})^{\text{c}}$
Bare PET	72.57±2.96	29.00±2.12	50.27±2.97	25.34±2.12	8.16±0.82	14.30±2.76
Bare OPP	90.06±1.84	79.72±0.69	52.99±2.65	17.18±3.41	22.72±1.26	11.23±1.38
CNs-PET	26.29±3.27	43.91±2.03	17.10±0.74	8.56±2.23	8.20±0.34	6.79±2.01
CNs-OPP	27.33±3.07	39.56±2.50	17.30±2.94	5.63±0.45	7.21±4.63	5.58±2.33

^a static contact angle values recorded at 60th second

^b DIM, diiodomethane

^c FOM, formamide,