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PROTEIN DATA BANK

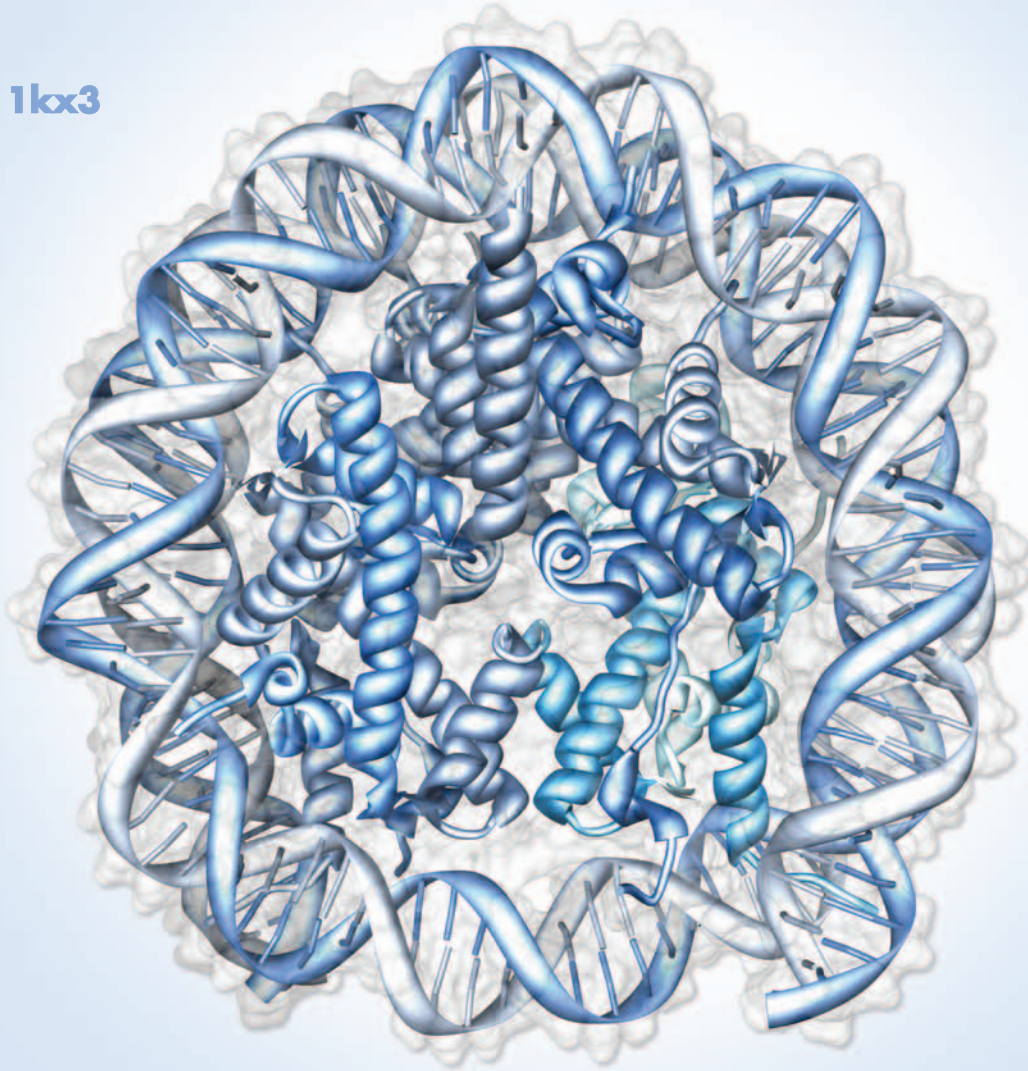


2008

Calendar

# DNA

1kx3



DNA is one of the most familiar molecules. As the central icon of molecular biology, it is easily recognized by everyone. Information is stored in the way that the bases match one another on opposite sides of the double helix – adenine with thymine, guanine with cytosine – to form a set of complementary hydrogen bonds (1bna).

Nucleosomes package DNA and also modify the activity of the genes that they store (1kx3). Each nucleosome is composed of eight "histone" proteins tightly bundled

and encircled by two loops of DNA. The histone proteins have long tails (not shown), which extend outward from the compact nucleosome, reaching out to neighboring nucleosomes and binding them tightly together. The nucleus contains regulatory enzymes that chemically modify these tails to weaken their interactions. In this way, the cell makes particular genes more accessible to polymerases, which lets their particular information be copied and used to build new proteins.

1bna



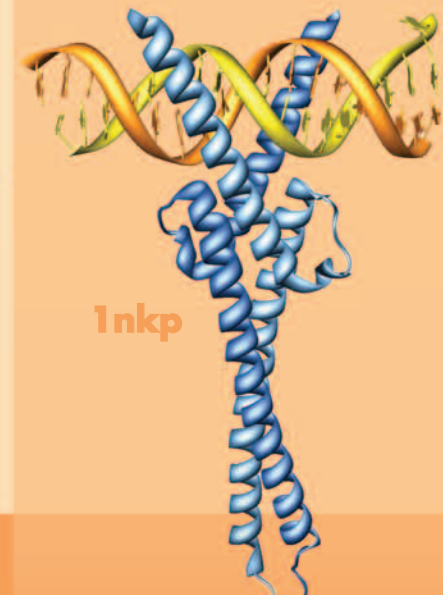
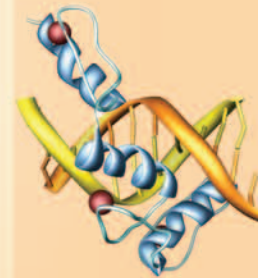
# January 2008

**1bnx:** H.R. Drew, R.M. Wing, T. Takano, C. Broka, S. Tanaka, K. Itakura, R.E. Dickerson (1981) Structure of a B-DNA dodecamer: conformation and dynamics. *Proc.Natl.Acad.Sci.USA* **78**:2179-2183.

**1kx3:** C.A. Davey, D.F. Sargent, K. Luger, A.W. Maeder, T.J. Richmond (2002) Solvent mediated interactions in the structure of the nucleosome core particle at 1.9 Å resolution. *J.Mol.Biol.* **319**:1097-1113.

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# Transcription Factors



Transcription factors are proteins that regulate gene expression by binding to DNA regulatory sequences. Examples of four structurally distinct types of transcription factors are shown:

Zinc finger proteins have a relatively simple modular structure. They recognize a diverse set of DNA sequences with key contacts made by a few residues from each finger (1a1h).

Helix-turn-helix motifs formed by two short connected helices are common in pro-

teins regulating DNA transcription and replication. Shown is one variant, a “winged helix-turn-helix” (1bc8).

$\beta$ -strands of “histone-like” integration host factor sharply bend DNA by opening up the minor groove (1owf).

Leucine zippers are dimeric motifs in which long helices are held by intertwined leucine residues. Both leucine zipper and helix-turn-helix motifs feature helices that fit snugly into the DNA major groove (1nkp).

# February 2008

**1owf:** T.W. Lynch, E.K. Read, A.N. Mattis, J.F. Gardner, P.A. Rice (2003) Integration Host Factor: putting a twist on protein-DNA recognition. *J.Mol.Biol.* 330:493-502.

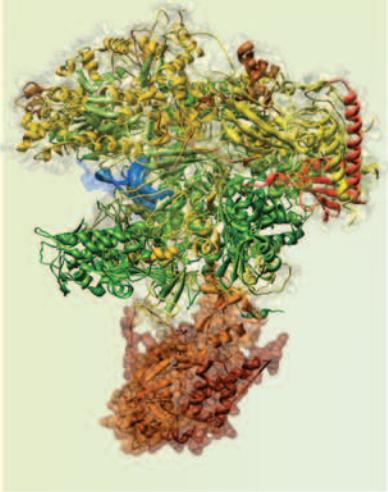
**1a1h:** M. Elrod-Erickson, T.E. Benson, C.O. Pabo, (1998) High-resolution structures of variant Zif268-DNA complexes: implications for understanding zinc finger-DNA recognition. *Structure* 6:451-464.

**1bc8:** Y. Mo, B. Vaessen, K. Johnston, R. Marmorstein, (1998) Structures of SAP-1 bound to DNA targets from the E74 and c-fos promoters: insights into DNA sequence discrimination by Ets proteins. *Mol. Cell* 2:201-212.

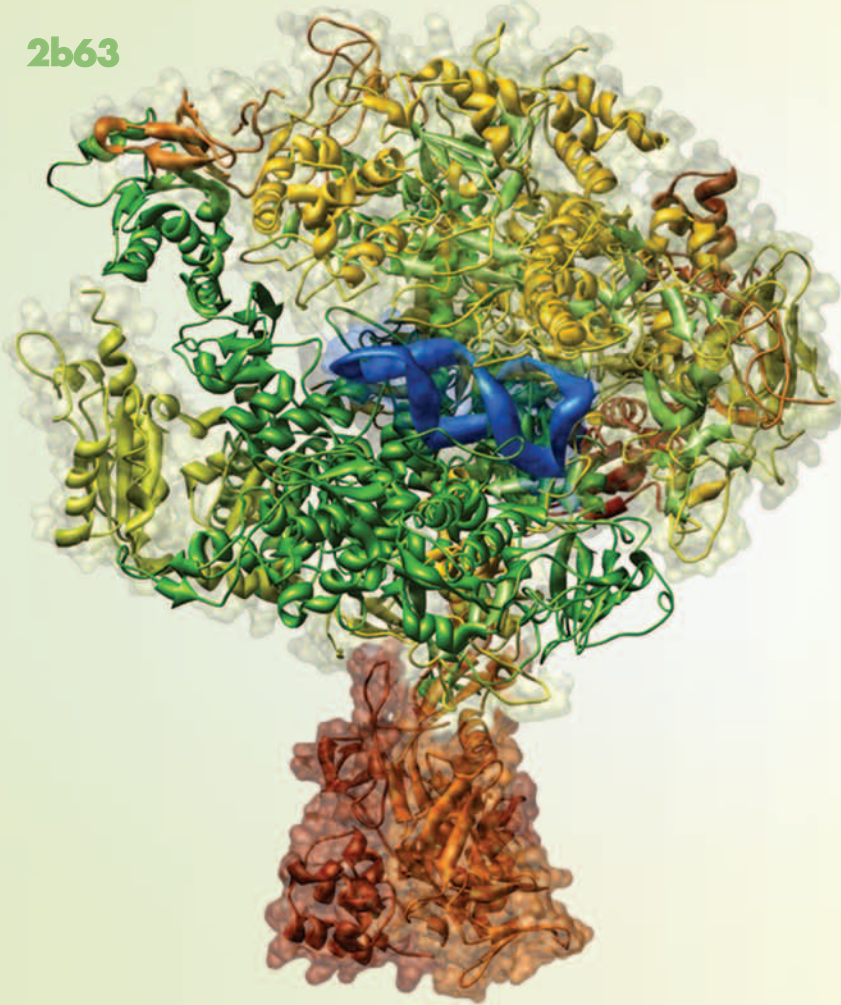
**1nkp:** S.K. Nair, S.K. Burley, (2003) X-ray structures of Myc-Max and Mad-Max recognizing DNA: Molecular bases of regulation by proto-oncogenic transcription factors. *Cell* 112:193-205.

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# RNA Polymerase



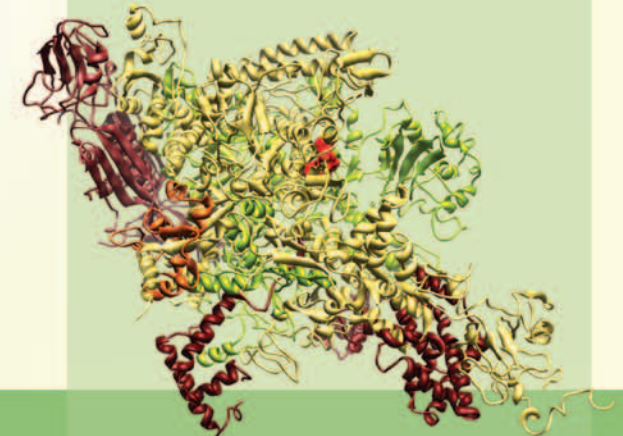
2b63



1lb2



1zyr



RNA polymerase is a complex and extremely precise nano-machine consisting of twelve different proteins. These proteins work in concert to unwind the DNA double helix, read the sequence from the anti-coding strand, and synthesize the messenger RNA.

The structure shown in 2b63 reports all twelve proteins of the yeast polymerase II enzyme in complex with a stem-loop RNA aptamer (in blue) that inhibits DNA

reading, but not RNA elongation, bound to the active center cleft. An example of an RNA polymerase found in bacteria is shown in 1zyr.

The complex shown in structure 1lb2 is the *E. coli* transcription factor catabolite activator protein interacting with one small piece of bacterial RNA polymerase to start transcription.

# March 2008

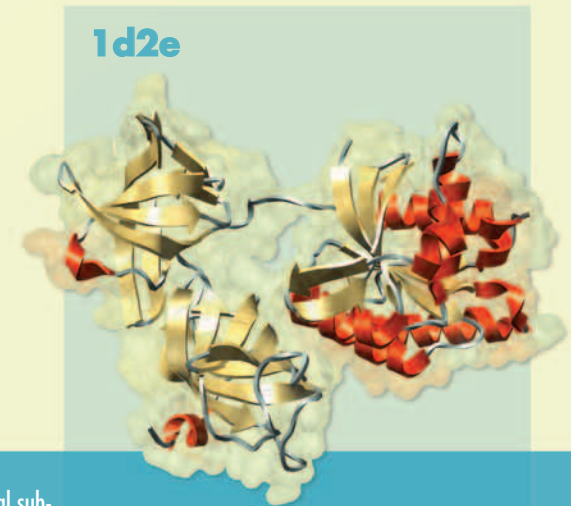
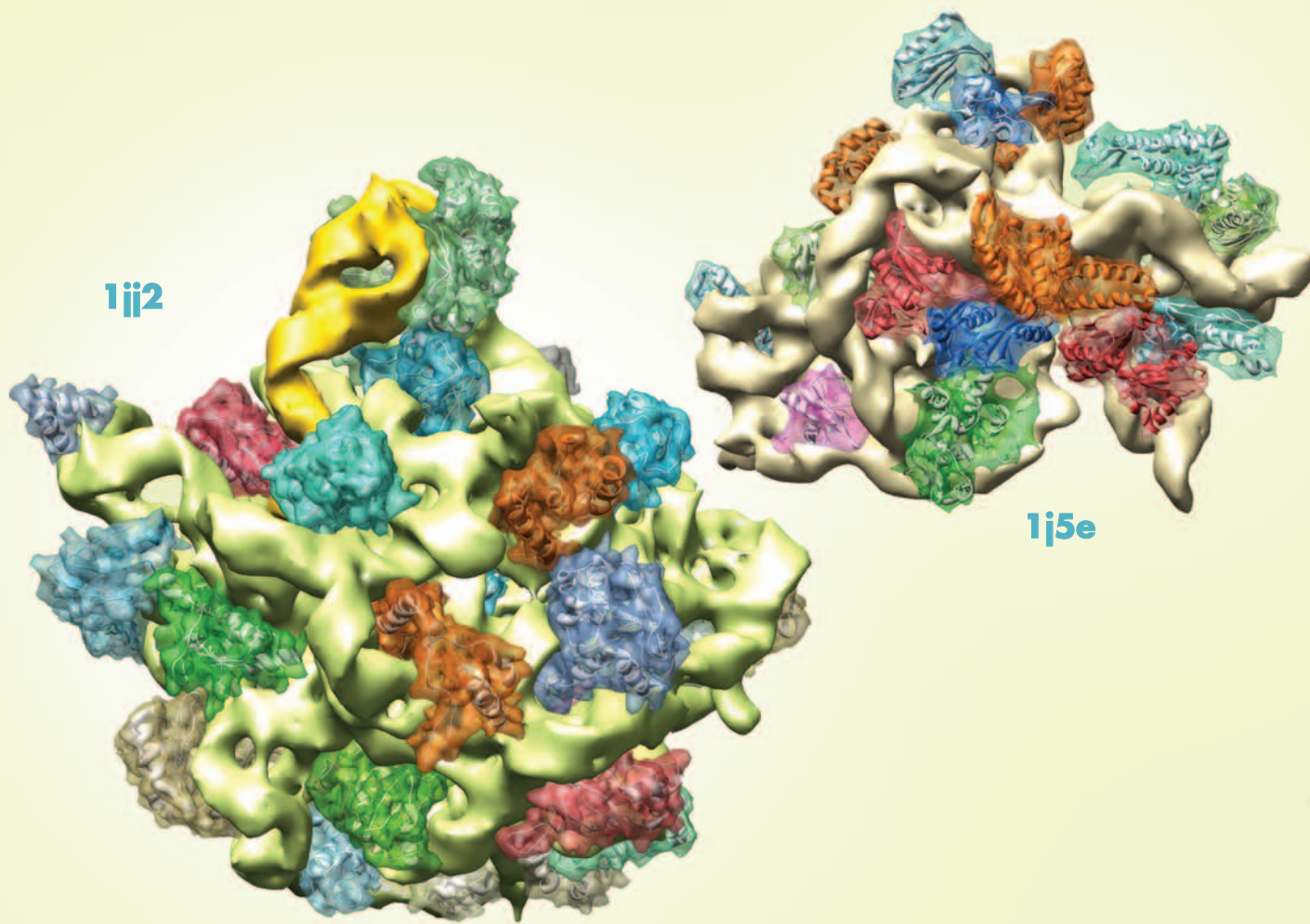
**11b2:** B. Benoff, H. Yang, C. L. Lawson, G. Parkinson, J. Liu, E. Blatter, Y.W. Ebright, H.M. Berman, R.H. Ebright (2002) Structural basis of transcription activation: the CAP-alpha CTD-DNA complex. *Science* **297**:1562-1566.

**2b63:** H. Kettenberger, A. Eisenfuehr, F. Brueckner, M. Theis, M. Famulok, P. Cramer (2006) Structure of an RNA polymerase II-RNA inhibitor complex elucidates transcription regulation by noncoding RNAs. *Nat.Struct.Mol.Biol.* **13**:44-48.

**1z9r:** S. Tuske, S.G. Sarafianos, X. Wang, B. Hudson, E. Sineva, J. Mukhopadhyay, J.J. Birktoff, O. Leroy, S. Ismail, A.D. Clark, C. Dharia, A. Napoli, O. Laptenko, J. Lee, S. Borukhov, R.H. Ebright, E. Arnold (2005) Inhibition of bacterial RNA polymerase by streptolydigin: stabilization of a straight-bridge-helix active-center conformation. *Cell* **122**:541-552.

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# Ribosomes



Protein synthesis is a major task performed by living cells. The translation of the genetic information from messenger RNA into protein is performed by the ribosome, a large nucleoprotein complex comprising two subunits. The small subunit is denoted 30S in bacteria (1j5e) and the large subunit, 50S (1ij2). Both subunits are composed of long strands of RNA, shown in light yellow, and about fifty small proteins shown in bright colors. When synthesizing a new protein, the two subunits assemble into a full ribo-

some, with a messenger RNA trapped in the space between. The large ribosomal subunit catalyzes peptide bond formation and binds initiation, termination, and elongation factors (such as the mitochondrial elongation factor Tu (EF-Tu) in 1d2e). The ribosome reads the messenger RNA in steps of three nucleotides, since each nucleotide triplet codes for one amino acid. Amino acid building blocks are carried to the ribosomal machine bound to transfer RNA (6tna).



# April 2008

**1j5e:** B.T. Wimberly, D.E. Brodersen, W.M. Clemons Jr., R.J. Morgan-Warren, A.P. Carter, C. Vonnheim, T. Hartsch, V. Ramakrishnan (2000) Structure of the 30S ribosomal subunit. *Nature* **407**:327-339.

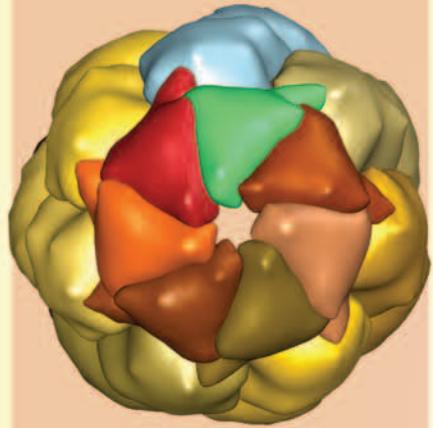
**1jj2:** D.J. Klein, T.M. Schmeing, P.B. Moore, T.A. Steitz (2001) The kink-turn: a new RNA secondary structure motif. *EMBO J.* **20**:4214-4221; N. Ban, P. Nissen, J. Hansen, P.B. Moore, T.A. Steitz (2000) The complete atomic structure of the large ribosomal subunit at 2.4 Å resolution. *Science* **289**:905-920.

**6tna:** J.L. Sussman, S.R. Holbrook, R.W. Warrant, G.M. Church, S.H. Kim (1978) Crystal structure of yeast phenylalanine transfer RNA. I. Crystallographic refinement. *J.Mol.Biol.* **123**:607-630.

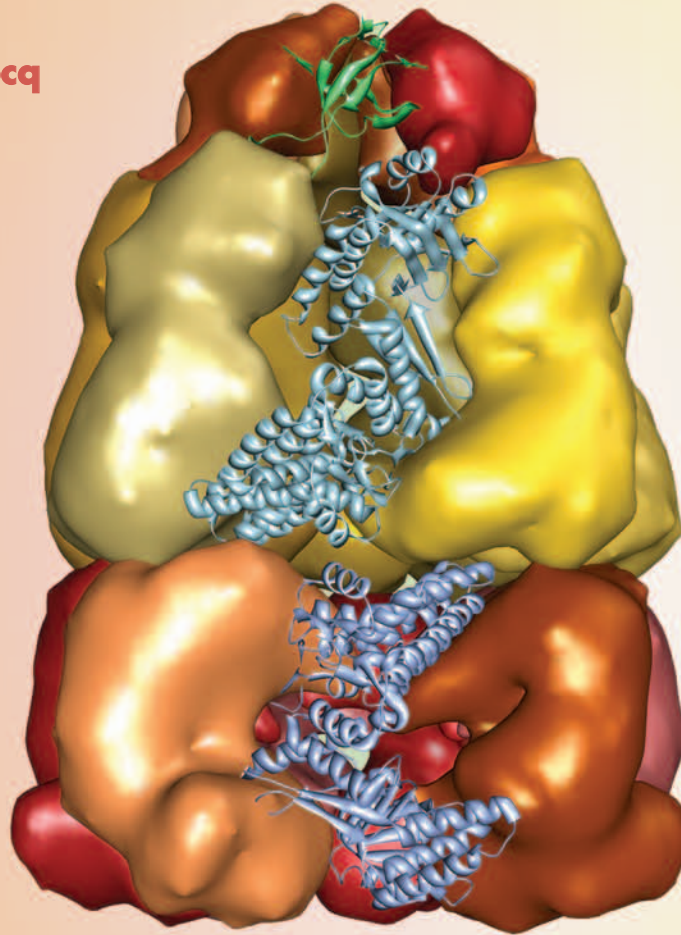
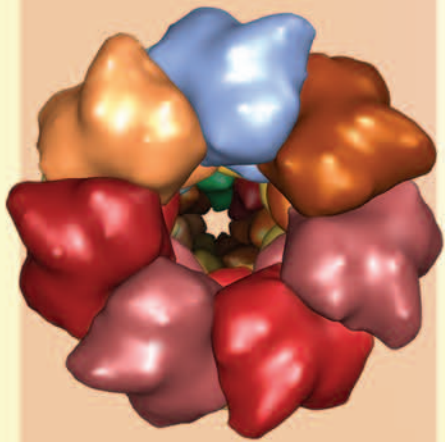
**1d2e:** G.R. Andersen, S. Thirup, L.L. Spremulli, J. Nyborg (2000) High resolution crystal structure of bovine mitochondrial EF-Tu in complex with GDP. *J.Mol.Biol.* **297**:421-436.

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# Chaperones



1pcq



1g31



Proteins are synthesized as formless chains and need to be folded into precise three-dimensional structures. Correct folding is critical for protein function. Chaperones are multimeric machines that help proteins fold correctly. Misfolded or unfolded proteins can lead to disease; Alzheimer's disease, for example, is caused by the unnatural aggregation of proteins into cell-clogging fibrils.

The GroEL/GroES chaperone complex helps to fold proteins by isolating them in a cage, away from other molecules present in the cytosol environment.

GroEL/GroES is composed of two stacked cylinders of GroEL proteins and a cap of GroES (1pcq, in red). The GroEL proteins form a hollow cylinder with a protein-sized cavity inside. Unfolded proteins enter this cavity and fold up inside. The heptamer of the Gp31 protein from bacteriophage T4 functionally substitutes for the bacterial co-chaperonin GroES in assisted protein folding reactions and effectively increases the size and the hydrophilicity of the folding cavity of native bacterial GroEL/GroES complex (1g31).

# May 2008

**1g31**: J.F. Hunt, S.M. van der Vies, L. Henry, J. Deisenhofer (1997) Structural adaptations in the specialized bacteriophage T4 co-chaperonin Gp31 expand the size of the Anfinsen cage. *Cell* **90**:361-371.

**1pcq**: C. Chaudhry, G.W. Farr, M. Todd, H.S. Rye, A.T. Brunger, P.D. Adams, A.L. Horwich, P.B. Sigler (2003) Role of the gamma-phosphate of ATP in triggering protein folding by GroEL-GroES: function, structure and energetics. *EMBO J.* **22**:4877-4887.

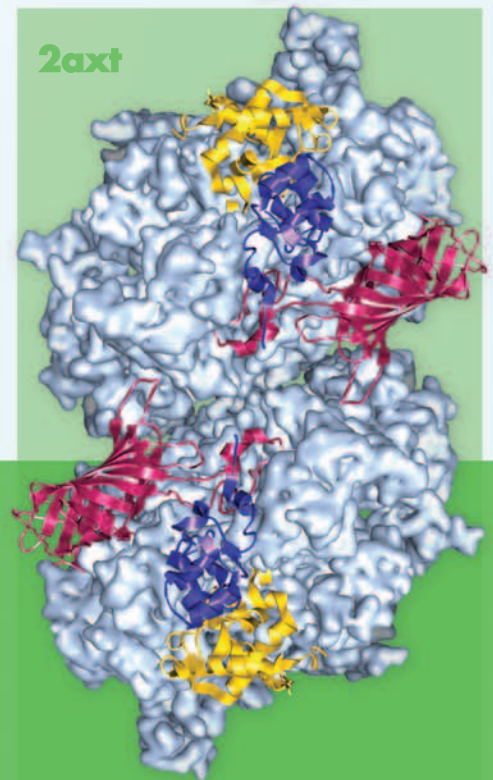
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# Photosynthesis

1jb0



1prc



2axt

The dominant energy source for living systems is sunlight. Photosynthesis turns sunlight into chemical energy. The photosynthetic proteins that perform this task form molecular complexes composed of more than a dozen proteins, tens of organic molecules ("cofactors"), and many metal cations. Photons from visible light excite electrons in green chlorophylls. The electrons are transferred to a series of other cofactors and finally reduce a carrier molecule,  $\text{NADP}^+$  to  $\text{NADPH}$ , which delivers them to enzymes that build sugar from water and carbon dioxide.

Photosystem II (2axt) and the photosynthetic reaction centre (1prc) capture photons and use their energy to extract electrons from water molecules. The electrons from Photosystem II are passed down a chain of electron-carrying proteins, getting an additional boost from Photosystem I (1jb0). As these electrons flow down the chain, they are used to pump hydrogen ions across the thylakoid membrane, providing power for ATP synthesis.

# June 2008

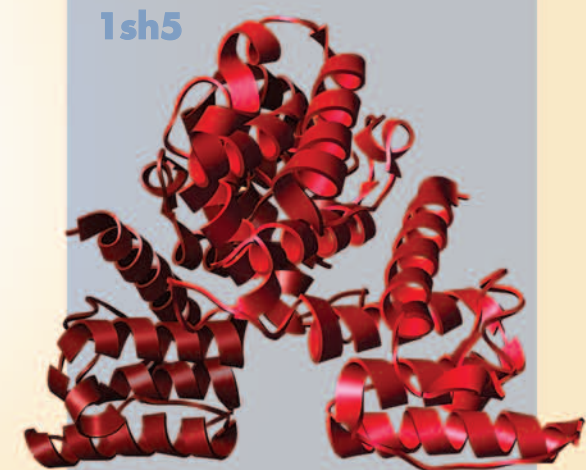
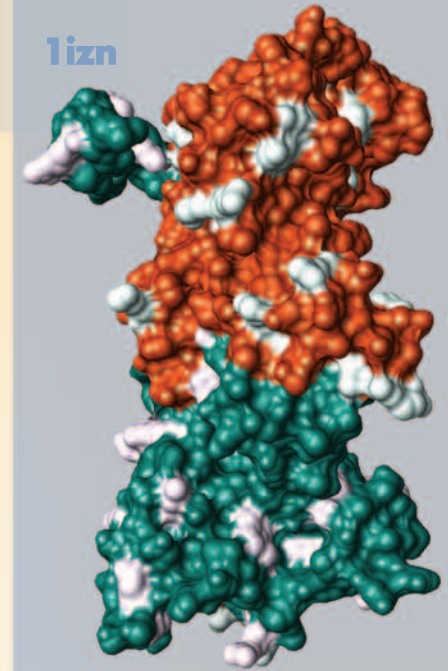
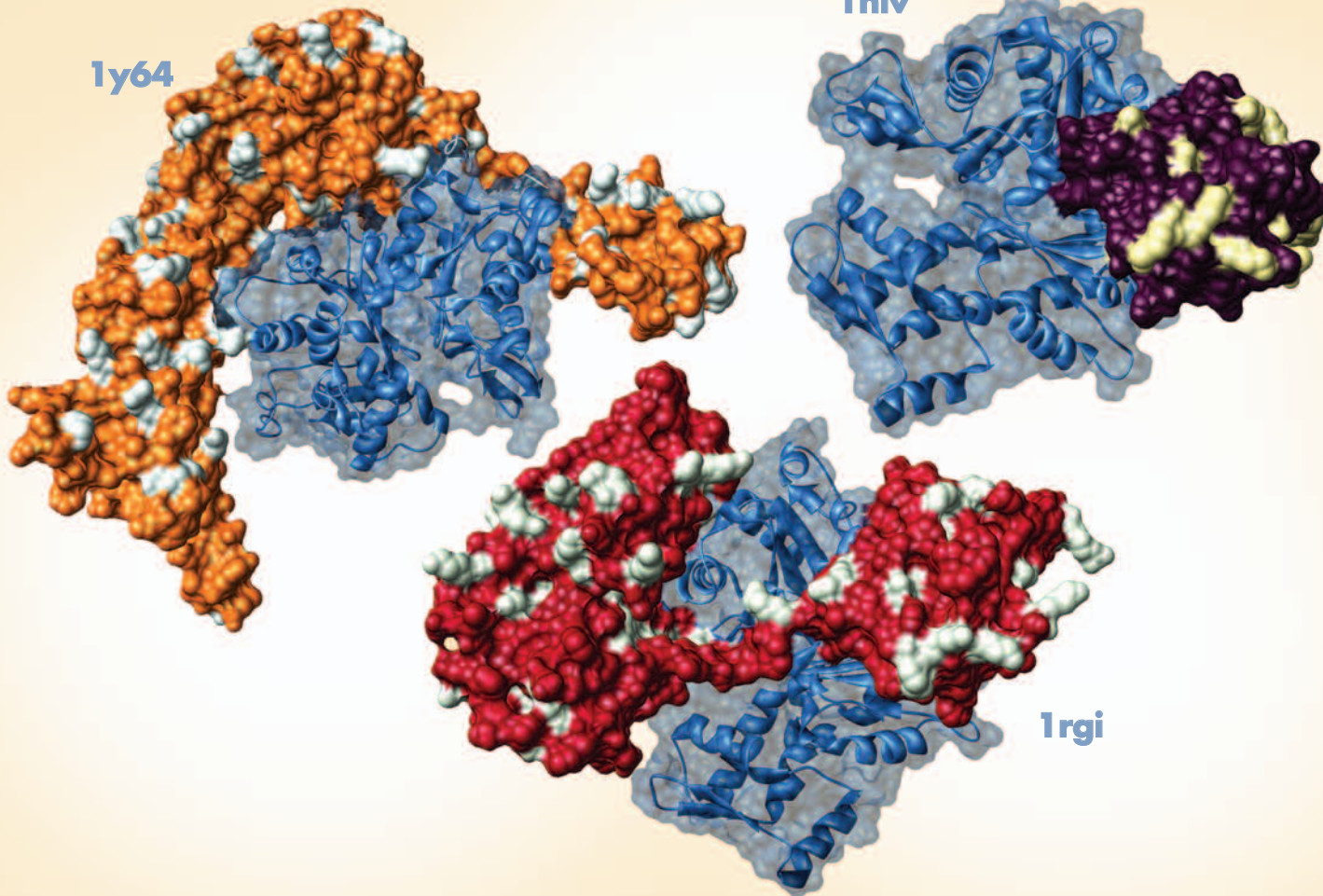
**1prc:** J. Deisenhofer, O. Epp, I. Sinning, H. Michel (1995) Crystallographic refinement at 2.3 Å resolution and refined model of the photosynthetic reaction centre from *Rhodospseudomonas viridis*. *J.Mol.Biol.* **246**:429-457.

**1jbo:** P. Jordan, P. Fromme, H.T. Witt, O. Klukas, W. Saenger, N. Krauss (2001) Three-dimensional structure of cyanobacterial photosystem I at 2.5 Å resolution. *Nature* **411**:909-917.

**2axt:** B. Loll, J. Kern, W. Saenger, A. Zouni, J. Biesiadka (2005) Towards complete cofactor arrangement in the 3.0 Å resolution structure of photosystem II. *Nature* **438**:1040-1044.

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# Actin



The cytoskeleton is an intracellular maze of filaments that supports and shapes the cell. The most plentiful type of filament is composed of actin, shown here in blue. The cytoskeleton, however, is not a static structure, since it must respond to the changing needs of the cell.

The proteins shown here help to reshape the cytoskeleton by assembling or disassembling actin filaments as necessary. A molecule of ATP, which is

bound inside each actin molecule, is important in this process. When it is hydrolyzed to ADP, the filament becomes unstable and falls apart.

Gelsolin breaks down actin filaments by assisting the hydrolysis of ATP and blocking the sites of interaction with other actin proteins. Two different fragments of gelsolin are shown in 1nlv and 1rgi bound to actin.

The protein CapZ forms a cap on the actin filaments shown in 1izn, which

limits assembly.

The protein formin assists the assembly of actin by aligning two actin proteins in the proper orientation which starts the process of filament growth. One domain of formin is shown bound to actin in 1y64.

Plectin links neighboring actin filaments into higher order structures. The actin-binding domain is shown in 1sh5.

# July 2008

**1nlv:** S.M.Vorobiev, B. Strokopytov, D.G. Drubin, C. Frieden, S. Ono, J. Condeelis, P.A. Rubenstein, S.C. Almo. The structure of non-vertebrate actin: Implications for the ATP hydrolytic mechanism (2003) *Proc.Natl.Acad.Sci. USA* **100**:5760-5765.

**1rgj:** L.D.Burtnick, D. Urosev, E. Irobi, K. Narayan, R.C. Robinson (2004) Structure of the N-terminal half of gelsolin bound to actin: roles in severing, apoptosis and FAF *EMBO J.* **23**:2713-2722.

**1tzn:** A.Yamashita, K. Maeda, Y. Maeda (2003) Crystal structure of CapZ: structural basis for actin filament barbed end capping *EMBO J.* **22**:1529-1538.

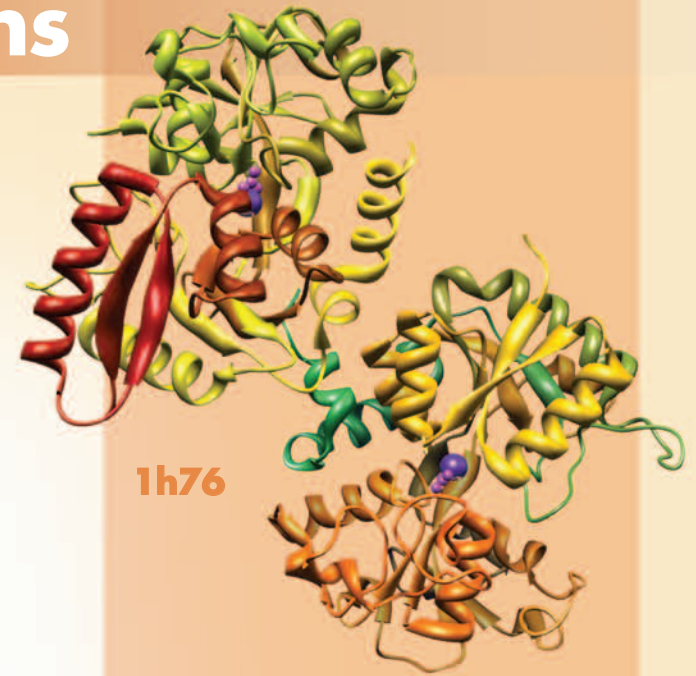
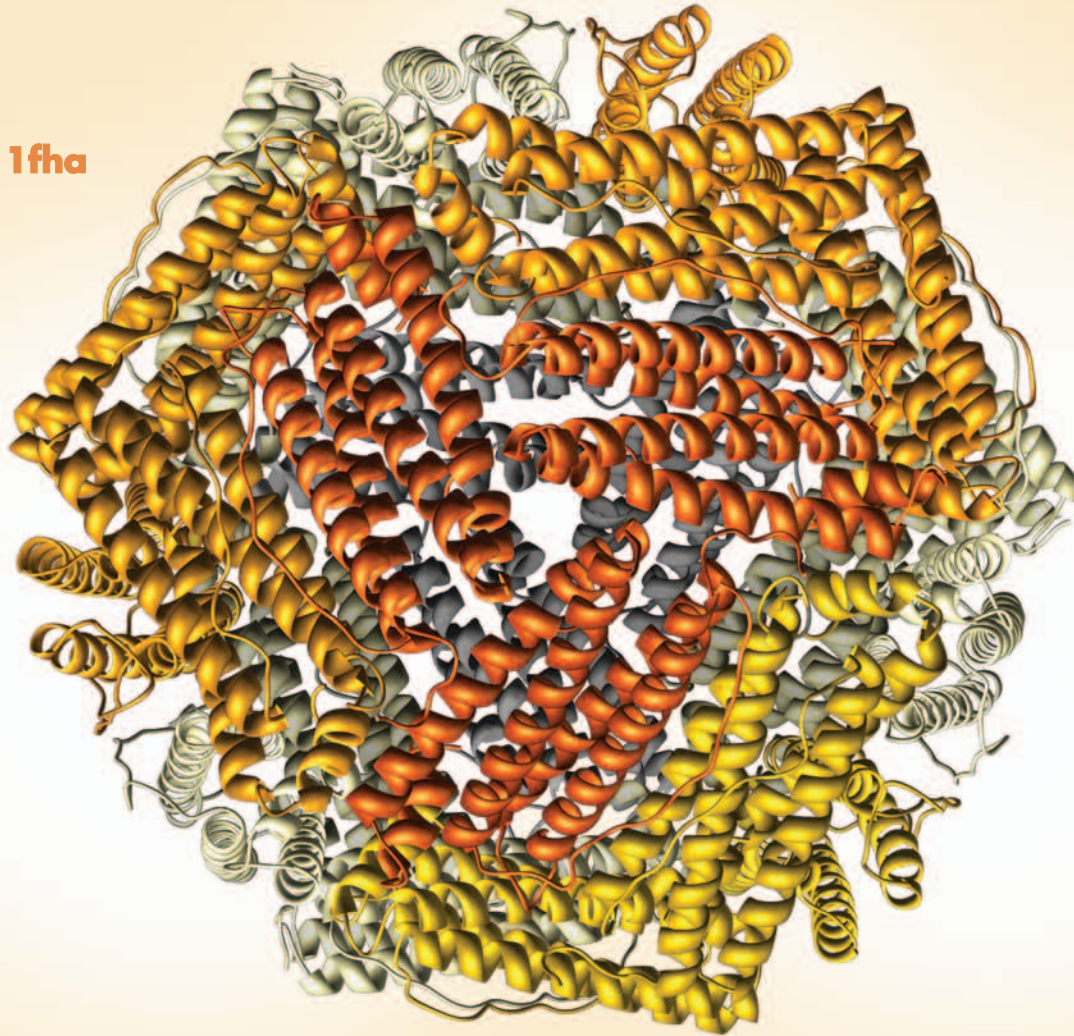
**1sh5:** J. Sevak, L. Urbanikova, J. Kostan, L. Janda, G. Wiche (2004) Actin-binding domain of mouse plectin: crystal structure and binding to vimentin *Eur.J.Biochem.* **271**:873-1884.

**1y64:** T. Otomo, D.R. Tomchick, C.Otomo, S.C. Panchal, M. Machius, M.K. Rosen (2005) Structural basis of actin filament nucleation and processive capping by a formin homology 2 domain. *Nature* **433**:488-494.

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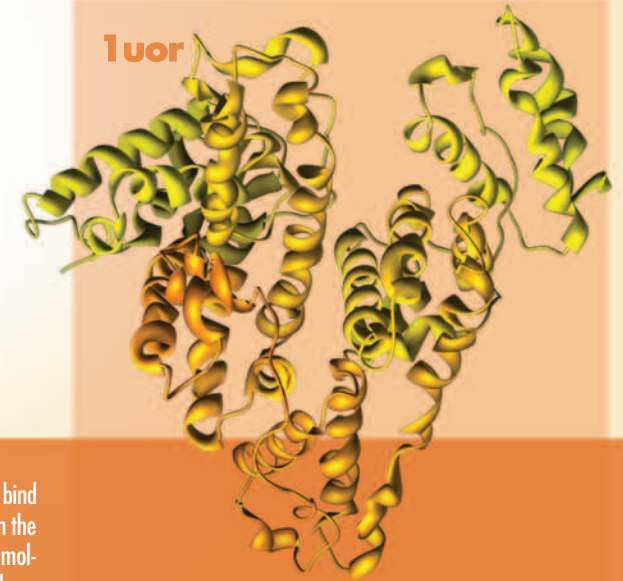
# Transport and Storage Proteins

1fha



1h76

1uor



Many transport proteins carry insoluble molecules from place to place through the bloodstream. Iron, for instance, is not very soluble in water, but it plays essential roles in enzymes, hemoglobin, and myoglobin throughout the body. It is transported through the blood by the protein transferrin (1h76), which is picked up by special receptors on cell surfaces and transferred inside. Once the iron is released inside cells, it is stored inside ferritin (1fha). Ferritin's twenty-four chains assemble into a hollow shell that provides an iron-storage cavity for up to 4500 iron ions.

Serum albumin (shown in 1uor) carries fatty acids in the bloodstream. They bind in deep crevices in the protein, burying their carbon-rich tails safely away from the surrounding water. Serum albumin also binds to many other water-insoluble molecules. In particular, serum albumin binds to many drug molecules, such as ibuprofen, and can strongly affect the way they are delivered through the body.



# August 2008

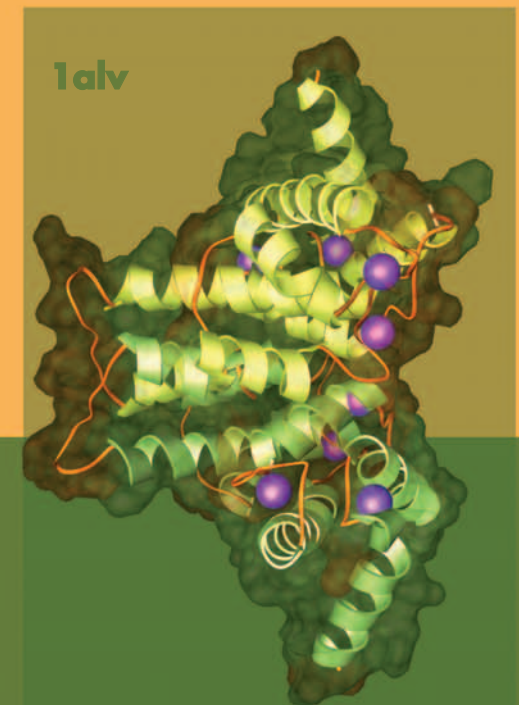
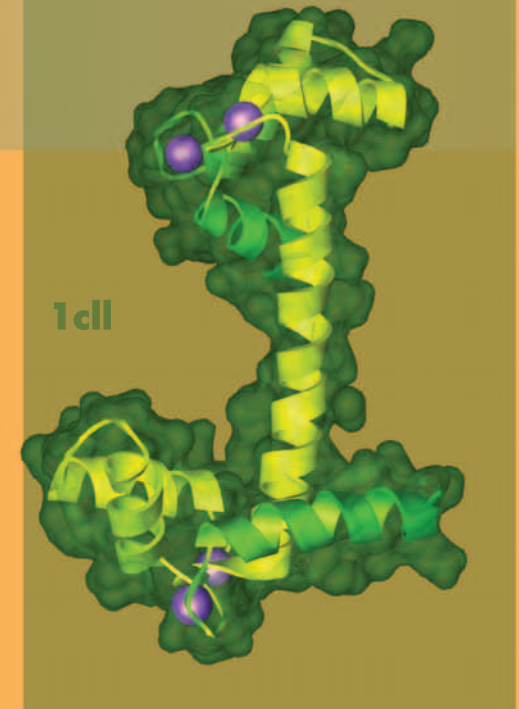
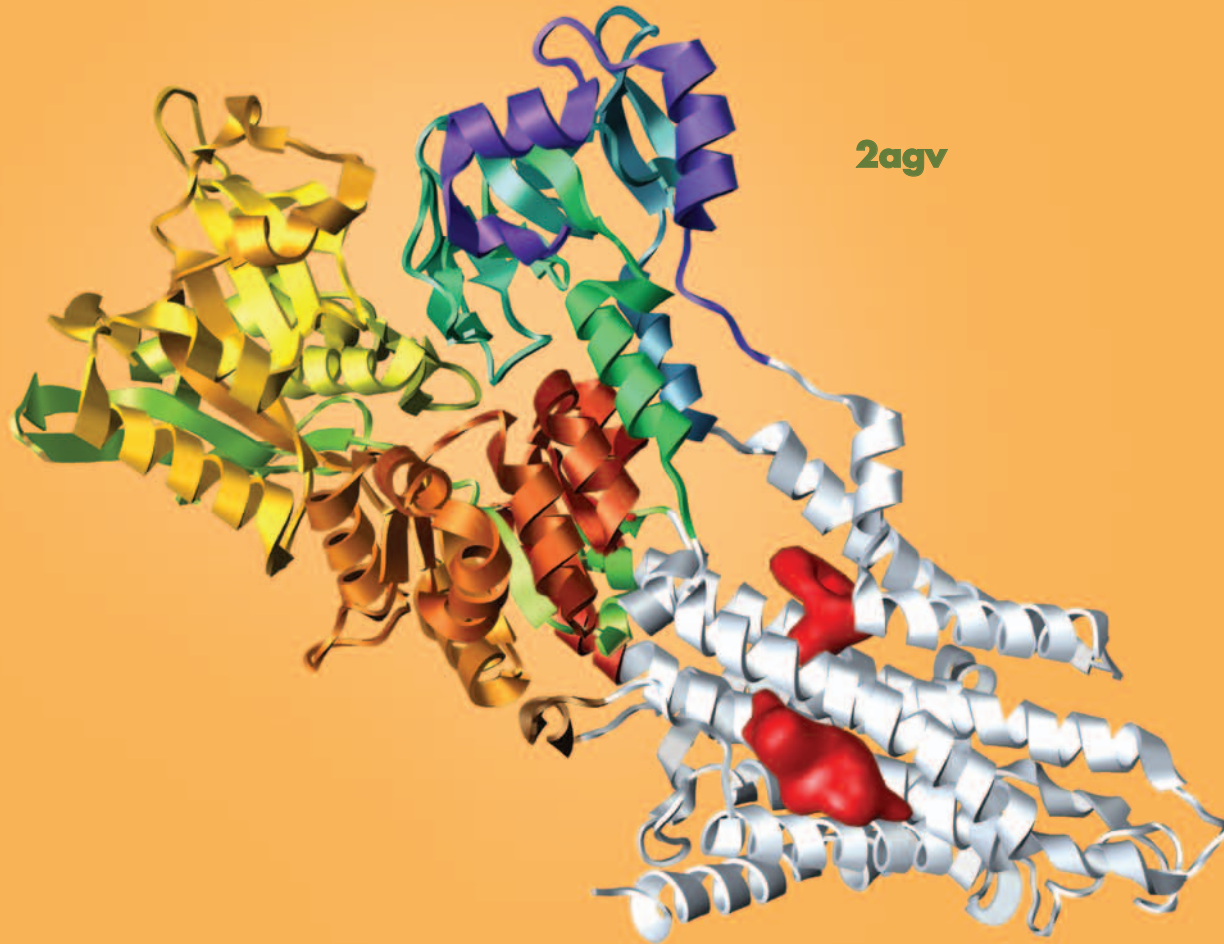
**1fha**: D.M. Lawson, P.J. Artymiuk, S.J. Yewdall, J.M. Smith, J.C. Livingstone, A. Treffry, A. Luzzago, S. Levi, P. Arosio, G. Cesareni, C.D. Thomas, W.V. Shaw, P.M. Harrison (1991) Solving the structure of human H ferritin by genetically engineering intermolecular crystal contacts. *Nature* **349**:541-544.

**1h76**: D.R. Hall, J.M. Hadden, G.A. Leonard, S. Bailey, M. Neu, M. Winn, P.F. Lindley (2002) The crystal and molecular structures of diferric porcine and rabbit serum transferrins at resolutions of 2.15 and 2.60 Å, respectively. *Acta Crystallogr., Sect.D* **58**:70-80.

**1uor**: X.M.He, D.C. Carter (1992) Atomic structure and chemistry of human serum albumin. *Nature* **358**:209-215.

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# Calcium Sensing and Transport



Calcium cations (shown here in purple) are essential for cells to exchange signals, such in nerve signaling and muscle contraction.

When a muscle cell is given the signal to contract, a flood of calcium ions are released and contractions follow. The calcium pump (shown in 2agv) uses ATP to pump calcium ions back to the sarcoplasmic reticulum. This allows muscles to relax after this frenzied wave of calcium-induced contraction.

Calmodulin is a CALcium MODULated protein (1cll). Calmodulin acts as an interme-

diary protein that senses calcium levels and relays signals to various calcium-sensitive enzymes, ion channels and other proteins. Calmodulin is a small dumbbell-shaped protein composed of two globular domains connected together by a flexible linker. Each end binds to two calcium ions.

Calpain is a calcium-dependent protease. The structure of domain VI (in entry 1alv), which senses calcium, is shown here.

# September 2008

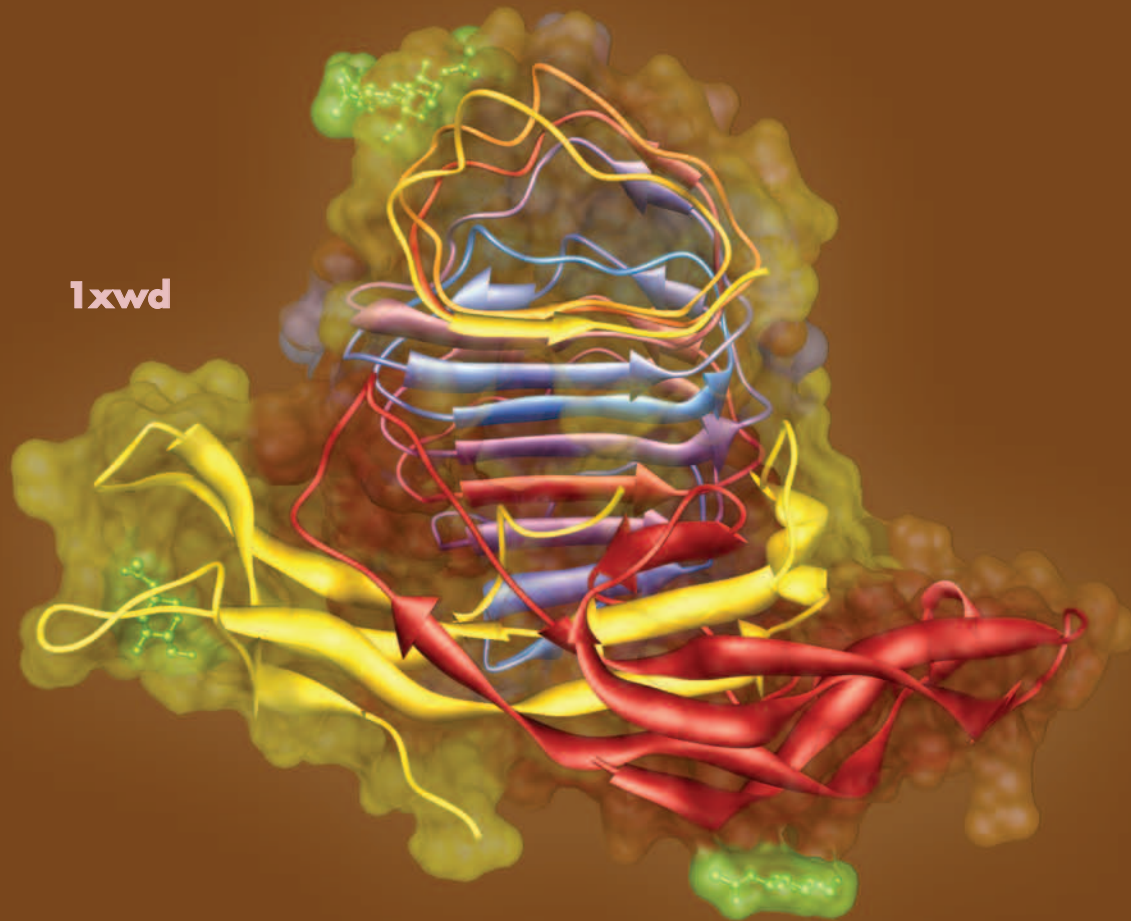
**2agv:** K. Obara, N. Miyashita, C. Xu, I. Toyoshima, Y. Sugita, G. Inesi, C. Toyoshima (2005) Structural role of countertransport revealed in Ca<sup>2+</sup> pump crystal structure in the absence of Ca<sup>2+</sup>. *Proc.Natl.Acad.Sci.USA* 102:14489-14496.

**1cll:** R. Chattopadhyaya, W.E. Meador, A.R. Means, F.A. Quijcho (1992) Calmodulin structure refined at 1.7 Å resolution. *J.Mol.Biol.* 228:1177-1192.

**1alv:** G.D. Lin, D. Chattopadhyay, M. Maki, K.K. Wang, M. Carson, L. Jin, P.W. Yuen, E. Takano, M. Hatanaka, L.J. DeLucas, S.V. Narayana (1997) Crystal structure of calcium bound domain VI of calpain at 1.9 Å resolution and its role in enzyme assembly, regulation, and inhibitor binding. *Nat.Struct.Biol.* 4:539-547.

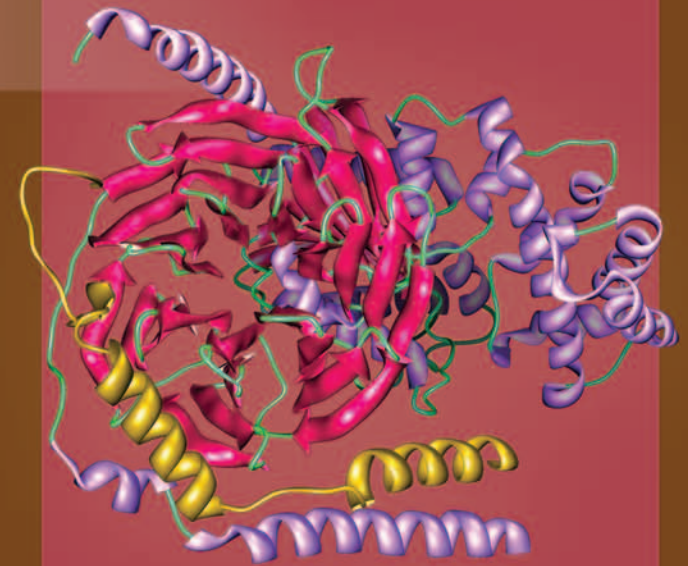
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# Receptors and G-Proteins



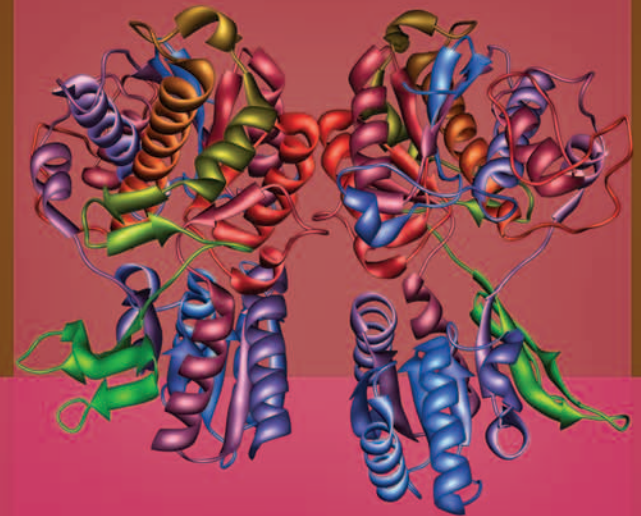
Messages between cells are mediated by many different molecules. When a message reaches its target cell, it must be recognized, transmitted across the cellular membrane, and translated to an appropriate response. This process is achieved by extracellular receptors and intracellular signaling proteins.

1xwd shows a complex of human follicle-stimulating hormone (strands on the bottom in dark red and yellow) bound to the extracellular hormone-binding domain of its receptor; the hormone has an elongated, curved shape.



1gg2

1ewk



G proteins, such as the example shown in 1gg2, are molecular switches that use the exchange of GDP (guanosine diphosphate) for GTP (guanosine triphosphate) to control their signaling cycle.

Glutamate acts as an excitatory neurotransmitter in the central nervous system. Dimeric metabotropic glutamate receptors, as shown in 1ewk, are coupled with G-proteins. Thus, neurotransmitter binding leads to cell signalling via G-proteins and finally release of calcium ions from intracellular stores.

# October 2008

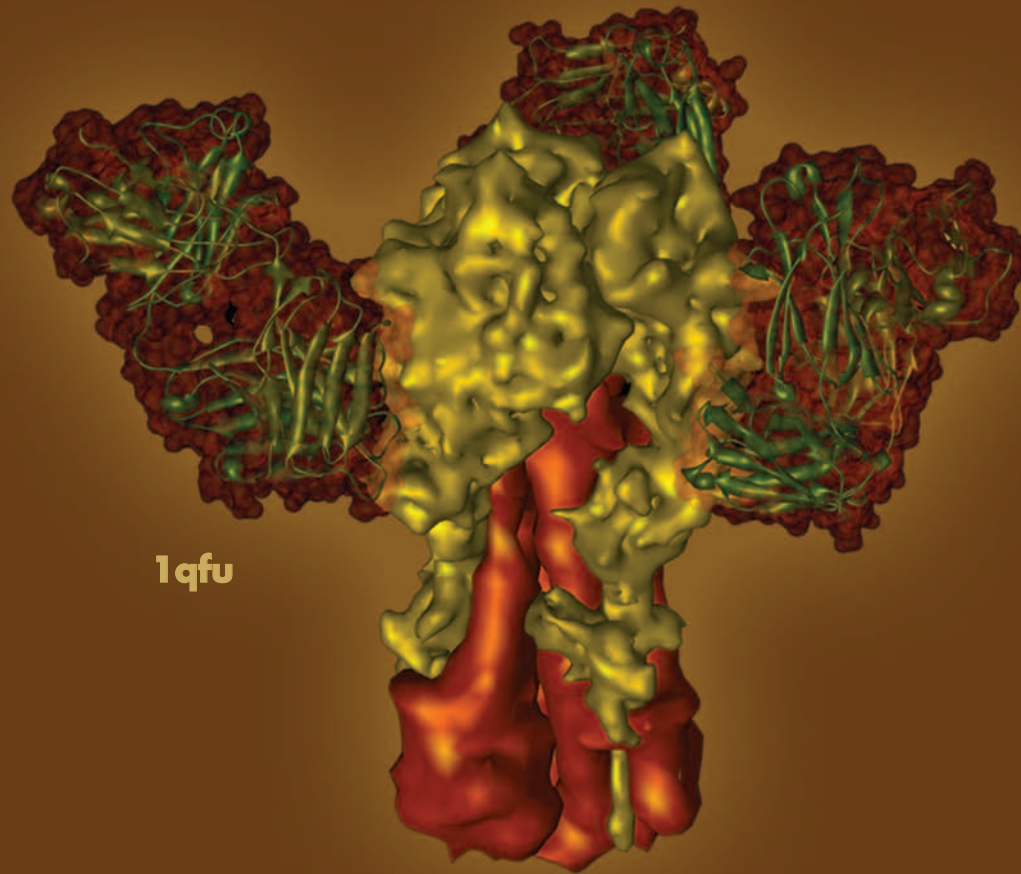
**1xwd:** Q.R.Fan, W.A. Hendrickson (2005) Structure of human follicle-stimulating hormone in complex with its receptor *Nature* **433**:269-277.

**1gg2:** M.A. Wall, D.E. Coleman, E. Lee, J.A. Iniguez-Lluhi, B.A. Posner, A.G. Gilman, S.R. Sprang (1995) The structure of the G protein heterotrimer  $G_{i\alpha 1}\beta\gamma 2$ . *Cell* **83**:1047-1058.

**1ewk:** N. Kunishima, Y. Shimada, Y. Tsuji, T. Sato, M. Yamamoto, T. Kumasaka, S. Nakanishi, H. Jingami, Morikawa, K. (2000) Structural basis of glutamate recognition by a dimeric metabotropic glutamate receptor. *Nature* **407**:971-977.

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# Antibodies



When a foreign molecule is found in the blood, many different antibodies may bind to it, attacking at different angles.

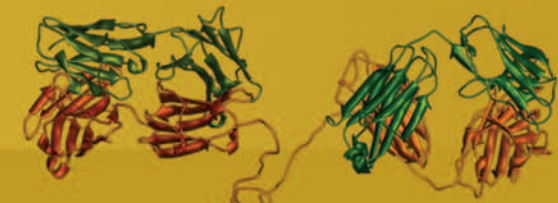
Antibodies, as exemplified with 1igt, are composed of two long heavy chains (brown) and two shorter light chains (green). The specific binding site is found at the tips of the two arms (each termed "Fab" for "antigen-binding fragment") in a pocket formed between the light and heavy chain. The binding site is composed of several loops in the protein chain that have very different lengths and amino acid composition. Differences in these "hypervariable loops" form the many types of pockets in different antibodies, each of which bind specifically to a different target. The rest of the antibody--the rest of the arms and the large constant domain that ties the two arms together--is relatively uniform in structure, providing a conven-

ient handle when antibodies interact with the rest of the immune system.

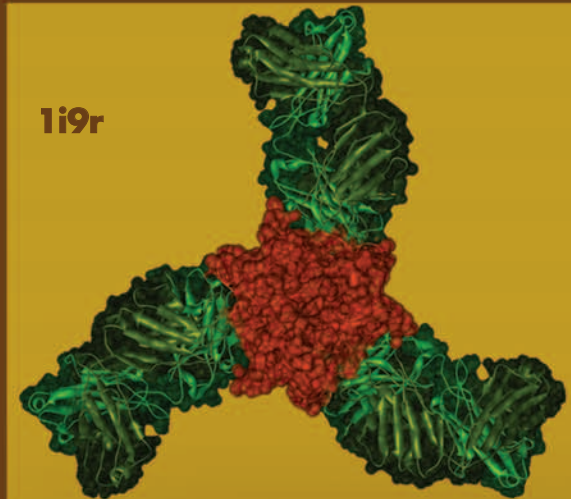
1qfu shows the structure of a complex of influenza hemagglutinin with neutralizing antibodies. Three copies of light and heavy Fab fragments of the antibody (in green) bind to the base of the stalk of hemagglutinin (in gold and brown).

Neuraminidase, another protein on the surface of influenza virus (brown), is complexed with four Fab fragments of monoclonal antibody Mem5 (green) in 2aep.

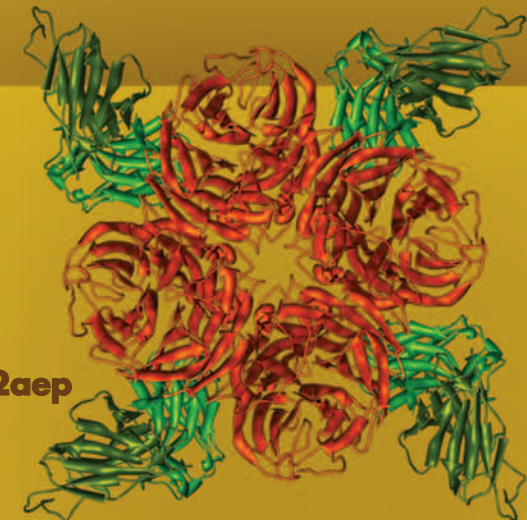
1i9r shows a member of the tumor necrosis factor family (brown) neutralized by three Fab fragments of 5c8 antibody (green).



**1igt**



**1i9r**



**2aep**

# November 2008

**1qfu:** D. Fleury, B. Barrere, T. Bizebard, R.S. Daniels, J.J. Skehel, M. Knossow (1999) A complex of influenza hemagglutinin with a neutralizing antibody that binds outside the virus receptor binding site. *Nat.Struct.Biol.* **6**:530-534.

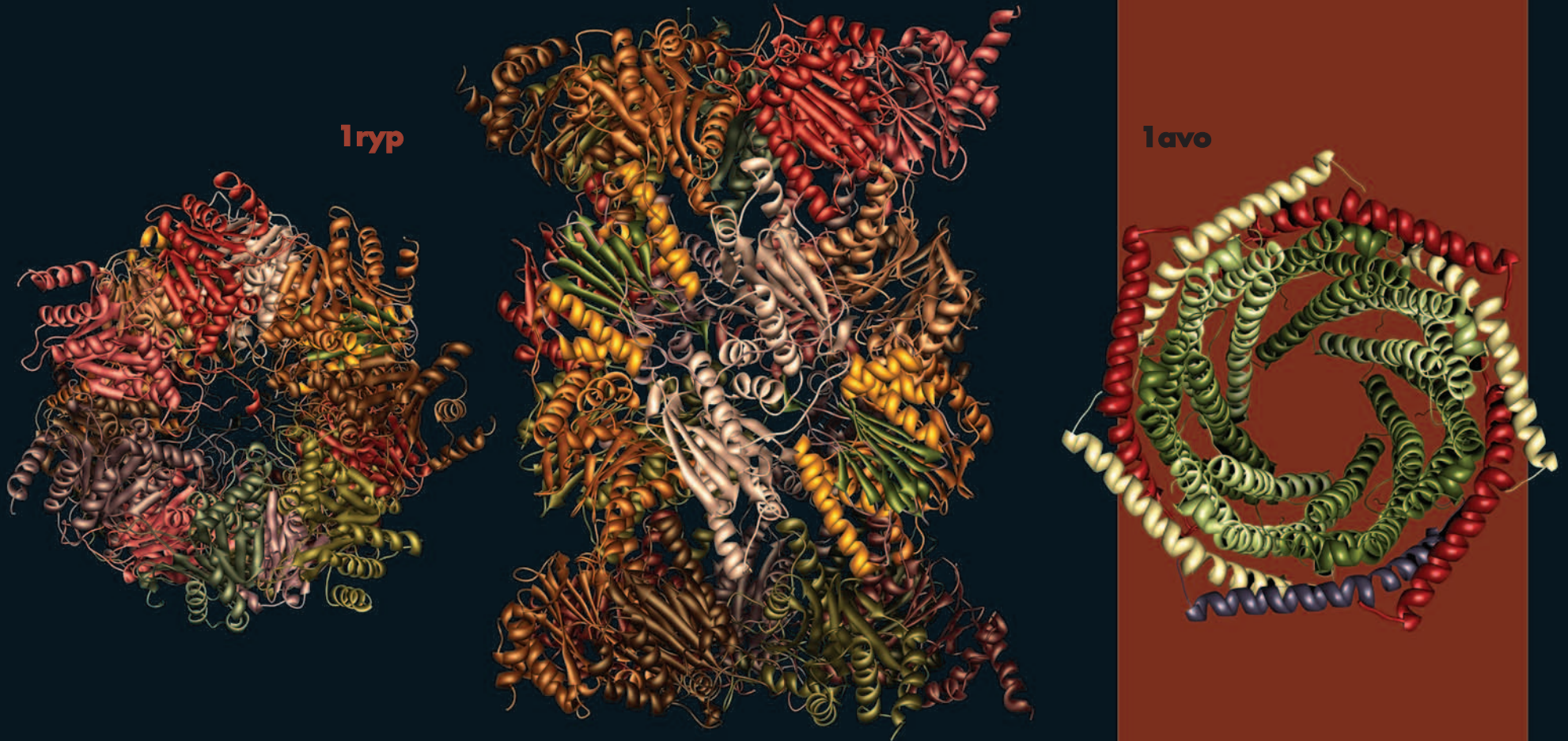
**2aep:** L. Venkatramani, E. Bochkareva, J.T. Lee, U. Gulati, W.G. Laver, A. Bochkarev, G.M. Air (2006) An epidemiologically significant epitope of a 1998 human influenza virus neuraminidase forms a highly hydrated interface in the NA-antibody complex. *J.Mol.Biol.* **356**:651-663.

**1i9r:** M. Karpusas, J. Lucci, J. Ferrant, C. Benjamin, F.R. Taylor, K. Strauch, E. Garber, Y.M. Hsu (2001) Structure of CD40 ligand in complex with the Fab fragment of a neutralizing humanized antibody. *Structure* **9**:321-329.

**1igt:** L.J. Harris, S.B. Larson, K.W. Hasel, A. McPherson (1997) Refined structure of an intact IgG2a monoclonal antibody. *Biochemistry* **36**:1581-1597.

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# Proteasomes



Proteasomes degrade damaged and misfolded proteins by breaking them into short peptides about seven amino acids long. These peptides are then further degraded into individual amino acids that can be re-used. Proteasomes are involved in cellular processes, including the cell division cycle and the regulation of gene expression.

The proteasome is a large barrel-like protein particle. It is formed by four stacked

rings, each formed by seven distinct protein chains (1ryp).

The action of the proteasome must be regulated so that it does not degrade proteins indiscriminately. The 11S regulator REGalpha, shown in 1avo, binds at each end of the proteasome. This opens up the entry channel and activates the proteasome.



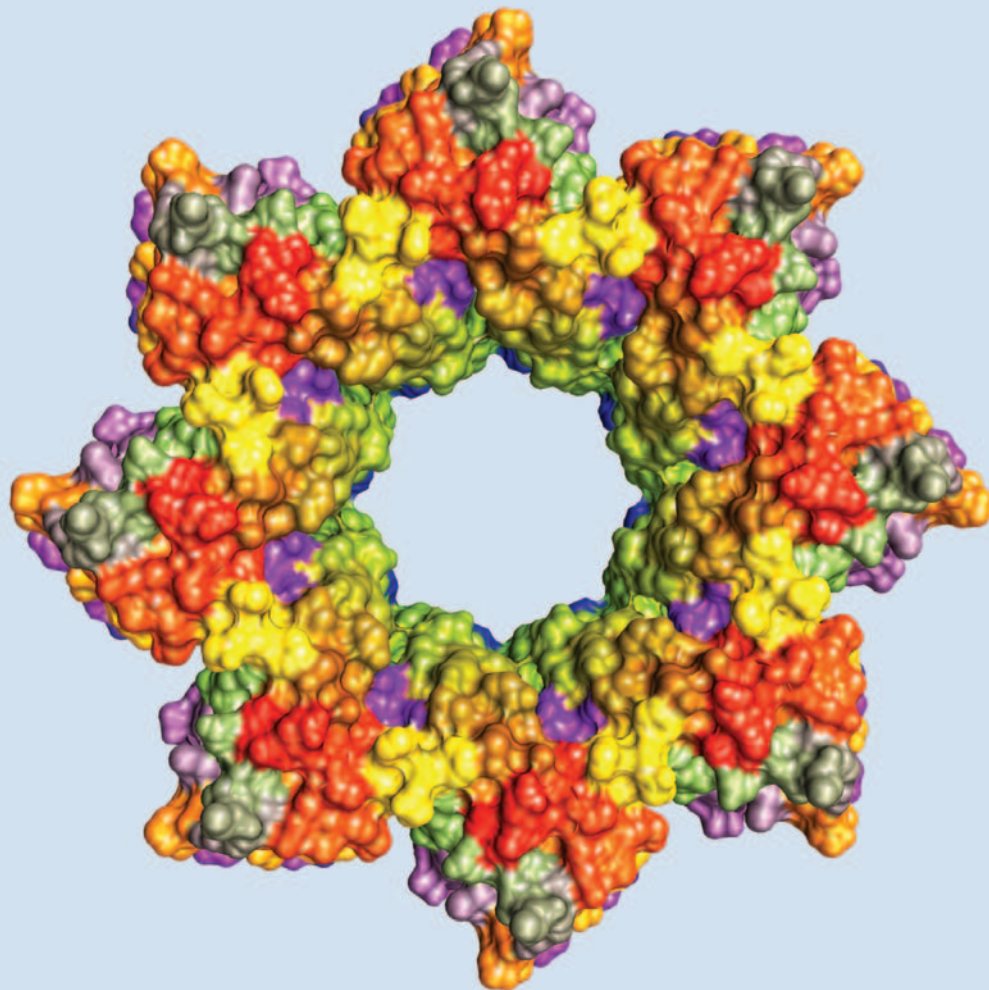
# December 2008

**Tryp:** M. Grall, L. Ditzel, J. Lowe, D. Stock, M. Bochtler, H.D. Bartunik, R. Huber (1997)  
Structure of 20S proteasome from yeast at 2.4 Å resolution. *Nature* **386**:463-471.

**Tavo:** J.R. Knowlton, S.C. Johnston, F.G. Whitby, C. Realini, Z. Zhang, M. Rechsteiner, C.P. Hill (1997) Structure of the proteasome activator REGalpha (PA28alpha) *Nature* **390**:639-643.

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# On the Cover



**1n0g:** S.Chen, J. Jancrick, H. Yokota, R. Kim, S.-H. Kim (2004) Crystal structure of a protein associated with cell division from *Mycoplasma pneumoniae* (GI: 13508053): a novel fold with a conserved sequence motif. *Proteins* **55**:785-791.

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# 2009 Calendar

## JANUARY

S	M	T	W	T	F	S
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## FEBRUARY

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## MARCH

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## APRIL

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## AUGUST

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