

Contents lists available at [ScienceDirect](http://ScienceDirect.com)

Biochimica et Biophysica Acta

journal homepage: www.elsevier.com/locate/bbamem

Review

Key roles of aquaporins in tumor biology[☆]

Marios C. Papadopoulos, Samira Saadoun^{*}*Academic Neurosurgery Unit, St. George's, University of London, London SW17 0RE, UK*

ARTICLE INFO

Article history:

Received 27 July 2014

Received in revised form 25 August 2014

Accepted 1 September 2014

Available online 7 September 2014

Keywords:

Aquaporin

Cancer

Review

Tumor

Water channel

ABSTRACT

Aquaporins are protein channels that facilitate the flow of water across plasma cell membranes in response to osmotic gradients. This review summarizes the evidence that aquaporins play key roles in tumor biology including tumor-associated edema, tumor cell migration, tumor proliferation and tumor angiogenesis. Aquaporin inhibitors may thus be a novel class of anti-tumor agents. However, attempts to produce small molecule aquaporin inhibitors have been largely unsuccessful. Recently, monoclonal human IgG antibodies against extracellular aquaporin-4 domains have become available and could be engineered to kill aquaporin-4 over-expressing cells in the malignant brain tumor glioblastoma. We conclude this review by discussing future directions in aquaporin tumor research. This article is part of a Special Issue entitled: Membrane channels and transporters in cancers.

© 2014 Elsevier B.V. All rights reserved.

Contents

1. Introduction	2576
2. AQP expression	2577
2.1. Normal tissue	2577
2.2. Tumors	2577
3. AQP functions.	2577
3.1. Tissue water balance	2578
3.2. Cell migration	2578
3.3. Cell proliferation	2579
3.4. Cell adhesion.	2580
4. AQP-based tumor therapeutics	2580
5. Future directions	2580
6. Conclusion	2581
Acknowledgments	2581
References.	2581

1. Introduction

Aquaporins (AQPs) are a family of water channel proteins, which are found in the plasma cell membranes of various cells [1]. There are 14 AQPs in mammals, at least eight of which have been shown to transport water. AQP3, AQP7 and AQP9 also transport glycerol and are termed aquaglyceroporins [2]. Though some AQPs transport gases and ions in

artificial systems, gas and ion transport are probably not relevant under physiological conditions [3,4]. To date, the key function of AQPs remains water transport.

AQPs assemble in cell membranes as homo-tetramers [5]. Each monomer is about 30 kDa and has its own water pore. In general, AQPs increase the water permeability of the plasma cell membrane 5–20 fold. AQP0 and AQP4 have unique properties; their tetramers assemble into higher order structures that form orthogonal arrays of particles. AQP4 exists as two isoforms, termed M1-AQP4 and M23-AQP4; M1 is the full-length protein with the sequence starting at methionine 1, whereas M23 lacks the first 22 amino acid sequence and starts at methionine 23. M23-AQP4 assembles in large orthogonal arrays, but

[☆] This article is part of a Special Issue entitled: Membrane channels and transporters in cancers.

^{*} Corresponding author. Tel.: +44 2087254179.
E-mail address: ssaadoun@sgul.ac.uk (S. Saadoun).

M1-AQP4 exists as individual tetramers. M1 can hetero-tetramerize with M23 to form orthogonal arrays, the size of which increases with increasing M23:M1 ratio [6]. Though *in vitro* studies show that AQPs freely diffuse in the plasma cell membrane, *in vivo* AQPs may be anchored to parts of the plasma cell membrane [7,8]. For example, in the central nervous system AQP4 is concentrated in perivascular astrocyte endfeet [9]. Intracellular membrane proteins such as alpha syntrophin [10] and extracellular matrix proteins such as agrin may form part of the AQP4 anchoring mechanism [11], but further research is required to define the molecular anchor.

In this review we discuss the functions of AQPs in tumor cells and how elucidating these functions has identified novel therapeutic targets in cancer biology.

2. AQP expression

2.1. Normal tissue

AQPs are widely distributed in human tissues and are generally preserved in mammals including rodents and humans [1]. In some organs, such as the kidney, several AQPs are expressed and play a major role in normal function [1,12]. For example, in response to antidiuretic hormone (vasopressin), AQP2 (which is found in intracellular vesicles) becomes expressed in the apical plasma cell membrane of collecting duct epithelial cells and increases the reabsorption of urine by the kidney. Humans with AQP2 mutations have congenital nephrogenic non-X-linked diabetes insipidus thus confirming the key role of AQP2 in water reabsorption by the kidney. In other organs, such as the brain, AQPs do not appear to play a major role in normal function, but become important in pathological conditions [1]. In mice that lack AQP4, which is normally expressed in perivascular astrocyte foot processes, the brain is phenotypically normal at baseline. In cerebral ischemia, brain tumors, bacterial meningitis and other conditions AQP4 becomes upregulated in astrocytes and facilitates brain edema formation and elimination. Recently, however, AQP4 was proposed to play a role in the clearance of solutes from the interstitial fluid of normal brain through a paravascular pathway termed the glymphatic system [13]. In some locations, such as the stomach, AQPs do not seem to be important for normal functions or in pathological conditions. Although AQP4 is expressed in the basolateral plasma cell membrane of gastric parietal cells, which are responsible for secreting hydrochloric acid, AQP4 deletion in mice does not affect gastric acid secretion. Aquaglyceroporins are involved in cellular metabolism [1,2]. AQP3 is expressed in the stratum corneum layer of the skin. AQP3 deletion in mice impairs skin hydration, elasticity, barrier recovery and wound healing due to lack of glycerol in the cells. These deficiencies in AQP3 null mice can be reversed by glycerol administration. AQP7, expressed in adipocytes, controls glycerol movement into and out of the cell [1]. AQP7 deletion in mice is associated with adipocyte hypertrophy, likely due to impaired glycerol exit from the adipocytes.

Several studies report regulation of AQP expression [1]. Though AQPs become upregulated or downregulated in response to different pathologies, it is impossible to determine from these descriptive studies the roles of AQP in these pathological conditions.

2.2. Tumors

Several authors have suggested a role for AQPs in cancer [14–20]. Table 1 summarizes AQP expression in different tumors. In general, tumor cells overexpress AQPs including AQPs that are normally found in their cell of origin as well as AQPs not present in the originating cell. In tumor cells, AQPs are expressed in the plasma cell membrane as well as the cytoplasm. There is often a strong correlation between the level of AQP expression and tumor grade. An example is diffuse astrocytoma, which is an infiltrating brain tumor that arises from astrocytes. Diffuse astrocytomas are histologically classified as grades II, III

Table 1
AQP expression in human tumors.

Tumor type	Aquaporins	AQP level	References
Astrocytoma	AQP1, 4, 78, 9	high	[21–32]
Breast cancer	AQP5	high	[77–79]
	AQP1	high	[79]
	AQP4	low	[79]
	AQP1	low	[80]
Cholangiocarcinoma	AQP1	high	[81]
	AQP1, 3, 5	high	[60,82–85]
Colorectal cancer	AQP8	low	[85,86]
	AQP1, 3	high	[87]
Cervical cancer	AQP5	high	[88]
	AQP1	high	[89]
Choroid plexus tumor	AQP1	high	[90]
Hemangioblastoma	AQP1	high	[91]
Laryngeal cancer	AQP1	high	[92]
Leukaemia	AQP5	high (CML)	[93]
Liver cancer	AQP3, 5	high	[94]
	AQP8, 9	low	[94]
	AQP1, 3, 5	high	[64,95,96]
Lung cancer	AQP1, 4	high	[97]
	AQP4	high	[98,99]
Meningioma	AQP1	high	[100]
Nasopharyngeal cancer	AQP3, 5	high	[101,102]
Oesophageal cancer	AQP1, 5, 9	high	[103,104]
Ovarian cancer	AQP3	high	[105]
Renal	AQP3	high	[106]
Skin, SCC	AQP3	high	[107]
Stomach cancer	AQP5	high	[108]
	AQP4	low	[109]
	AQP3	high	[110]
	AQP4	high (papillary)	[110]
Thyroid cancer	AQP3, 4	low (undifferentiated)	[111]
	AQP7	high	[111]
	AQP3, 5	high (SCC)	[112]
Tongue cancer	AQP3	low	[113,114]
Urinary bladder	AQP3	low	[113,114]

CML, chronic myelogenous leukaemia; SCC, squamous cell carcinoma; AQP level indicates level of AQP expression in the tumor compared with the normal tissue.

or IV, with the most malignant grade IV also termed glioblastoma. Astrocytes normally express AQP4 in their perivascular foot processes. Normal astrocytes express AQP4, but little or no AQP1 and AQP9. We initially reported, using immunohistochemistry, strong AQP4 [21] and AQP1 [22] expression in diffuse astrocytomas with the level of expression positively correlating with tumor grade. There is now substantial evidence from different investigators that AQP1, AQP4 and AQP9 are strongly expressed in human astrocytomas [21–33]. Another example of human tumors that strongly express AQPs is epithelial ovarian tumors. By immunoblot there is substantially higher AQP7 and AQP9 protein expression in malignant and borderline tumors compared with benign tumors and normal ovarian tissue with AQP9 expression level positively correlating with tumor grade [34]. Though some tumor types show reduced AQP expression compared with their cell of origin, this is only seen for individual AQPs; when considering several AQPs, there appears to be increased overall AQP expression in all tumors listed in Table 1. In some cases there is correlation between increased AQP expression in the tumor and patient prognosis. Potential caveats of these studies include reporting bias for positive studies, failure to distinguish tumor cells from other cell types found within the tumor (e.g. reactive astrocytes, fibroblasts or leukocytes) and the poor specificity of some antibodies used for immunostaining e.g. anti-AQP9. Notwithstanding these caveats, the large number of tumor AQP expression studies raises the intriguing hypothesis that AQPs contribute to carcinogenesis in a range of tumor types.

3. AQP functions

In addition to the well-established role of AQPs in maintaining tissue water balance, other roles of AQPs include facilitating cell migration, cell proliferation and cell adhesion (Fig. 1). Under each section below, we first discuss the role of AQPs in normal cells, followed by their role in tumor cells. The functional data discussed here are largely derived

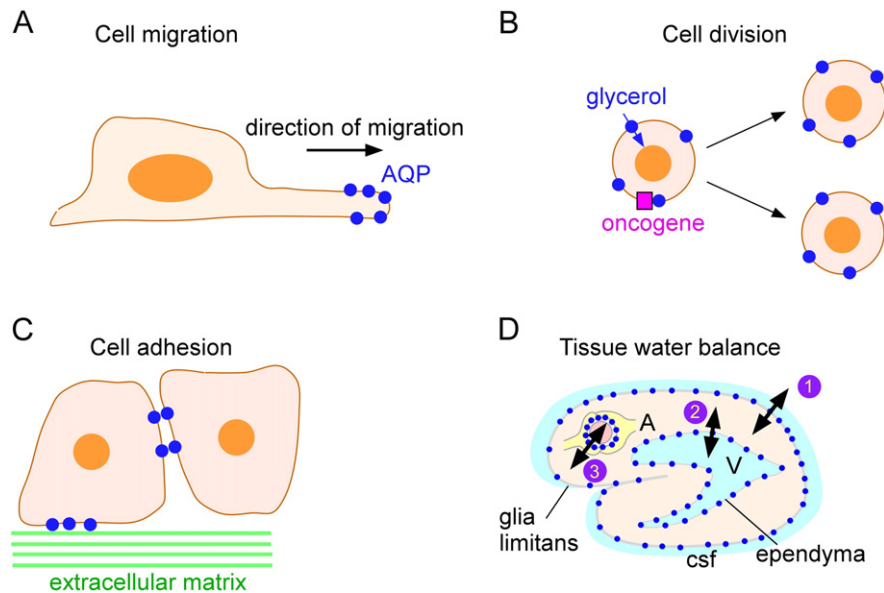


Fig. 1. Roles of AQPs in cancer. A. Cell migration. AQP polarizes to the leading end of the cell and facilitates formation of the lamellipodium. B. Cell proliferation. Aquaglyceroporin facilitates glycerol entry into the cell, which is essential for biosynthesis. AQP may directly interact with oncogenes. C. Cell adhesion. AQP0–AQP0 or AQP4–AQP4 binding increases cell–cell adhesion. AQP may increase adhesion of the cell to the extracellular matrix. D. Tissue water balance. Example of brain where AQP4 controls water flow between the brain and major fluid compartments: 1) glia limitans (brain–subarachnoid cerebrospinal fluid), 2) ependyma (brain–ventricular cerebrospinal fluid), 3) astrocyte foot processes (brain–blood). A, astrocyte; csf, cerebrospinal fluid; V, ventricle.

from experiments comparing wildtype vs. AQP null mice and should be interpreted with caution because of compensatory effects that may have developed in these mice as a result of longstanding AQP deletion. To minimize compensatory effects, some authors have created central nervous system specific AQP4 deletion in mice [35]. Ideally, conditionally inducible AQP knockout mice are required to eliminate the confounding effect of long-term adaptation to AQP deletion. Such mice are not currently available, but may provide more definitive information about AQP4 function. Another way to elucidate AQP function is to inhibit AQPs; this is not currently possible because of a lack of non-toxic, specific AQP inhibitors [36].

3.1. Tissue water balance

There is now substantial evidence that AQPs play a major role in maintaining water balance in several tissues. One of the first organs to be investigated following the discovery of AQPs was the kidney, in which multiple AQPs (AQP1, AQP2, AQP3 and AQP4) are responsible for water absorption and elimination [12]. Key roles for AQPs have also been defined in the central nervous system, with AQP4 playing a central role in edema fluid accumulation and elimination, and AQP1 in cerebrospinal fluid production [37]. The mechanisms of AQP4 involvement in brain edema are discussed in detail elsewhere [37]. In the salivary gland, AQP5 is found in the apical plasma cell membrane of acinar epithelial cells and plays a role in osmotic water transport across the cells into the sweat duct [38]. In some tissues, loss of AQP expression in mice does not influence function, e.g. deletion of AQP1, AQP3, AQP4, and AQP5 in mouse lacrimal glands does not affect tear secretion, probably due to the low level of transepithelial water transport rates in these tissues [39]. In general, AQPs facilitate water flow across cells in response to osmotic gradients produced by salt transport.

A major role for aquaporins in tumor edema was first postulated by Saadoun et al. who showed that astrocytomas express high levels of AQP1 [22] and AQP4 [21] with the level of AQP expression positively correlating with the presence of brain tumor edema on computed tomography. The most malignant astrocytomas also express AQP9 [23, 28, 29]. AQP4 is expressed by astrocytoma cells, but also by reactive astrocytes in and around the tumor [21]. Subsequent experiments showed that AQP4 deletion in mice increases edema around B16F10 brain

melanoma [40]. In this mouse model there is prominent reactive gliosis around the melanoma. These findings suggest that the increased AQP4 expression by reactive astrocytes in and around the tumor facilitate elimination of brain edema fluid. Whether AQP4 expressed in tumor cells also plays a role in brain edema remains unknown and could be defined by implanting AQP4-expressing and non-expressing tumor cells in mice and quantifying the water content of the tumor and brain. Because the brain is surrounded by the non-distensible skull, brain tumor edema is a major clinical problem that causes increased intracranial pressure, brain ischemia, herniation and, ultimately, brain death. To date, a role for AQPs in tissue edema associated with tumors outside the central nervous system has not been investigated.

3.2. Cell migration

An unanticipated role for AQPs in facilitating cell migration was first suggested by Loitto et al. who studied AQP9 in neutrophils [41]. Subsequently, Saadoun et al. showed that several AQPs facilitate cell migration in different cell types including AQP1 in aortic endothelial cells [42] and AQP4 in astrocytes [43]. The overall conclusion from several studies *in vitro* and *in vivo* is that AQP expression enhances cell migration towards a chemotactic stimulus [41–47]. The exact mechanism remains unclear, but may involve targeted water entry into the leading edge of a migrating cell, which enhances formation of the lamellipodium (a flattened protrusion at the leading end of a migrating cell, which is essential for cell motility) [48]. The idea that AQPs facilitate formation of the lamellipodium is consistent with the polarization of AQPs to the leading end of migrating cells. It has been suggested that AQPs also facilitate the rapid changes in cell shape that take place as a migrating cell squeezes through the tortuous extracellular space [14]. Such changes in cell volume are likely to require rapid flow of water into and out of the cell. Some authors have recently suggested that cells may utilize directed water permeation mediated by AQPs to create a net inflow of water and ions at the cell leading edge and a net outflow of water and ions at the trailing edge leading to net cell displacement [49]. This mechanism, termed the osmotic engine model, may allow cell migration through confined micro-spaces without the need for actin depolymerization–polymerization or myosin II-mediated contractility. It is important to note that lack of AQPs does not entirely inhibit cell migration, but renders

migration towards a chemotactic stimulus less efficient. This may explain why AQP-null mice develop normally *in utero* even though cell migration is an important component of embryogenesis.

Tumor cell migration is a fundamental property of different cancers and contributes to tumor cell infiltration into surrounding tissue as well as the metastatic spread. B16F10 melanoma cells and 4T1 breast cancer cells transfected to express AQP1 were more likely to extravasate after tail vein injection in mice [50]. The resulting lung tumors were more diffusely infiltrating into the surrounding alveolar tissue compared with tumor cells lacking AQP1. AQP1 is important for endothelial cell migration that takes place during angiogenesis, which is vital to permit solid tumors to grow rapidly. Melanoma tumors produced by subcutaneous implantation of B16F10 cells grow faster in wildtype than AQP1 null mice [42]. Histological examination of these tumors revealed that AQP1 deficiency is associated with impaired tumor angiogenesis. Tumor vascular endothelial cells express AQP1, which plays a major role in tumor angiogenesis by facilitating endothelial cell migration. These findings are supported by several follow-on studies. One study found reduced proliferation of implanted melanoma cells in mice treated with AQP1 siRNA [51]. AQP1 deficiency in mice that spontaneously develop well-differentiated breast adenomas with lung metastases reduced total tumor mass and volume compared with wildtype mice, due to impaired angiogenesis in the AQP null mice [52]. A key role for AQP1 in enhancing angiogenesis has also been shown in several other non-tumor pathologies including liver cirrhosis [53,54] and hypoxia-inducible angiogenesis in the retina [55]. Together, these findings suggest that, by enhancing cell migration and angiogenesis, AQPs may facilitate tumor growth, local infiltration and metastasis.

Further work is required to define the role of AQPs in tumor cell infiltration. This could be achieved by mapping the location of AQP-expressing and non-expressing astrocytoma cells in human tumor specimens. If AQP expression increases tumor cell infiltration, then we hypothesize that the AQP-expressing cells will be located in infiltrative parts of the tumor whereas non-AQP expressing cells that should be mostly within the tumor core. A recent study reported AQP1 expression in astrocytoma cells in areas of tumor infiltration, distant from the necrotic tumor core, thus supporting this hypothesis [25]. Experiments are also required to investigate whether it is the water transport, or another, unknown AQP function that contributes to tumor cell infiltration and spread.

The mechanism by which AQPs facilitate cell migration remains unknown. Here we propose a novel hypothesis, that AQPs do not increase the speed of migrating cells, but by polarizing to the leading edge, AQPs ensure that the lamellipodium forms in the direction of the chemotactic gradient. This effect may enhance the directionality of migration i.e. cells expressing AQPs follow a less tortuous route towards their target compared with cells lacking AQPs (Fig. 2). A detailed description of the molecular basis of cell migration is beyond the scope of this paper and is the subject of other reviews [56,57]. A useful analogy for a migrating cell is a moving car. The wheels of the car are the integrins, which are transmembrane proteins that allow cell–cell and cell–extracellular matrix interaction during cell migration. The petrol in the car is the ATP that provides the energy for cell migration. The car engine is the actin cytoskeleton that forms the propulsion system for cell migration. The road is the extracellular space. A map is also needed, which is provided by several proteins that facilitate detection of the chemotactic gradient. Here we propose that the AQPs are the steering wheel, which ensures that the moving car heads towards its destination. Further experiments are required to test these ideas, by determining the effect of AQP expression on the speed, directionality and tortuosity of cells in migrating chemotactic chambers.

3.3. Cell proliferation

AQP3, which is expressed in the epidermis, enhances the proliferation rate of basal keratinocytes [46]. AQP3 null mice have impaired

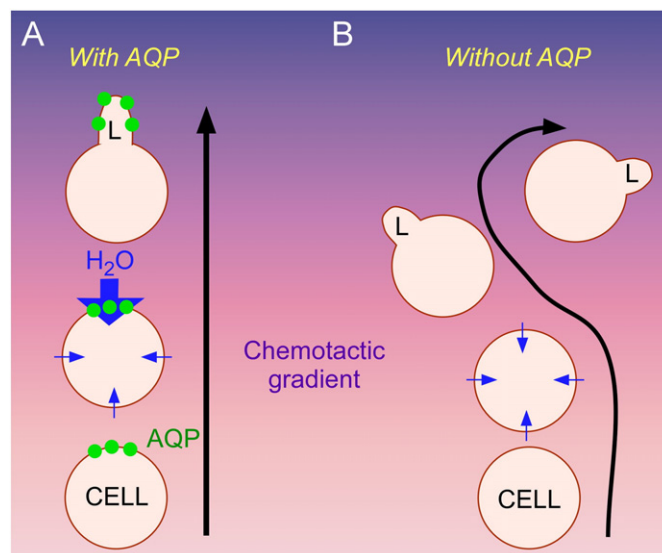


Fig. 2. Proposed role of AQP in increasing directionality of migrating cell. A. AQP polarizes to the leading end of the cell thus ensuring that the lamellipodium forms in the direction of the chemotactic gradient. The cell migrates toward the chemotactic gradient. B. Without AQP, the lamellipodium does not form in the direction of the chemotactic gradient and, therefore, the cell migrates in a more tortuous route.

wound healing, due to reduced glycerol and ATP content in the keratinocytes, which are required for biosynthesis [58,59]. Little is known about the role of aquaglyceroporins 7 and 9 in the proliferation of normal cells.

There is direct and indirect evidence that AQP3 and AQP5 play a role in tumor cell proliferation. AQP5 might interact with the Ras pathway in colon cancer [60]. Ras activation switches on other proteins that ultimately turn on genes involved in cell growth, differentiation and survival. Another study showed AQP5-facilitated lung cancer cell proliferation and migration, possibly through activation of the EGFR/ERK/p38 MAPK signaling pathway [61]. These AQP5–oncogene interactions may represent novel AQP functions, which are unrelated to water transport. AQP3 null mice are remarkably resistant to the development of skin tumors following exposure to the tumor initiator and promoter, phorbol ester [62]. Glycerol supplementation corrected the reduced proliferation in AQP3 deficiency, with cellular glycerol, ATP, and proliferative ability being closely correlated. There is, therefore, an established link between AQP3 expression in the epidermis and skin cancer. It is worth noting here that some moisturizing creams (such as Eucerin®, Be+®, HydrAction®) are marketed to improve skin hydration by increasing AQP3 expression in keratinocytes. At least one investigator has cautioned the cosmetics industry that products that increase AQP3 expression in the skin might be carcinogenic [63]. AQP3 expression is high in non-small cell lung cancer [64] and, in a mouse model, AQP3 knock-down suppressed tumor growth and reduced angiogenesis in human non-small cell lung cancer xenografts [65].

Though a role for some AQPs in facilitating tumor cell proliferation seems likely, further studies are required to define the link between the expression level of some AQPs, notably AQP3 and AQP5, and tumor cell proliferation. This could be achieved in a variety of human tumors by doubly immunostaining for the AQP of interest and the cell proliferation marker Ki67. If AQP increases cell proliferation, then a greater proportion of AQP⁺ than AQP[−] tumor cells should also be Ki67⁺. It is unclear whether and how some AQPs directly interact with oncogenes or whether the increased proliferation in AQP expressing tumor cells is a secondary effect of increasing the glycerol content in the cell. It is also unclear whether the water transporting function of AQPs is necessary to increase cell proliferation.

3.4. Cell adhesion

A role for AQP0 in cell–cell adhesion is well established. AQP0 is expressed in lens fiber cells in the eye where it constitutes about 50% of the fiber cell membrane protein. Studies using AQP0 null mice revealed that AQP0 is important for maintaining the structure of interlocking protrusions that is critical to the integrity and transparency of the lens. A role for AQP4 in cell–cell adhesion was also proposed based on structural considerations [66]. AQP4 contains a short helix in an extracellular loop, which mediates weak interactions between AQP4 molecules in adjoining plasma cell membranes, in effect binding adjacent cells to each other. Expression of AQP4 in L-cells (which lack endogenous adhesion molecules) resulted in clustering of the cells thus supporting the idea that AQP4 may play a role in cell–cell adhesion. There is also evidence against a significant effect of AQP4 expression on adhesion in several different cell types, including L-cells [67]. Recent experiments show that during cell migration, M1-AQP4 isoforms (which exist as individual tetramers) polarize to the leading edge of the cell, to support cell migration [68]. However, the larger M23-AQP4 rich orthogonal arrays do not enter the lamellipodium, but become bound with adhesion complexes, suggesting a role for M23-AQP4 but not M1-AQP4 in cell adhesion to the extracellular matrix. We conclude that data regarding the role of AQP4 in cell adhesion are contradictory. It is thus unclear whether AQP4 plays any role in tumor cell adhesion.

4. AQP-based tumor therapeutics

If AQPs play a role in tumor cell infiltration, metastasis, proliferation and possibly cell adhesion, then AQP modulators may be useful anti-cancer agents. For effects that are dependent on the water transport property of AQPs such as tumor cell migration, inhibitors of AQP transport are required. For other effects that are independent of water transport through the AQP channel other drugs are required, e.g. disruptors of AQP-oncogene interaction to reduce cell proliferation.

To illustrate the therapeutic potential of AQP inhibitors, we consider glioblastoma, which is the most common primary brain tumor with fatal prognosis. The median survival from diagnosis is about a year even with aggressive treatment (radical surgery, radiotherapy and temozolomide chemotherapy) [69,70]. What makes glioblastoma so aggressive is its ability to infiltrate extensively into the brain, which renders the tumor impossible to excise surgically. Drugs that inhibit water flow through the AQP4 pore may reduce tumor cell infiltration thus converting a tumor that is not surgically excisable into a tumor with well-defined margins that can be surgically resected. Currently, chemotherapy and radiotherapy target the rapidly dividing cells. By eliminating the most infiltrative cells, AQP4 inhibitors would offer a novel therapeutic option, which targets the main cause of tumor malignancy in glioblastoma. Vascular endothelial cells within glioblastoma express AQP1 and, therefore, AQP1 inhibitors may reduce angiogenesis, which would secondarily inhibit glioblastoma growth.

A recent review has summarized attempts to develop AQP-based therapeutics and concluded that such attempts have been largely unsuccessful [36]. Heavy metals such as mercury inhibit AQP1 but are too toxic for clinical use. There are several reports of AQP4 inhibitors including acetazolamide, anti-epileptic drugs, bumetanide, sumatriptan and thiazazole [71–73]. Unfortunately, follow-on studies on these compounds by other investigators using different assays have failed to confirm AQP4 inhibition. For a detailed discussion on the issues related to discovering AQP inhibitors please refer to the recent review by Verkman et al. [36]. One interesting development is the discovery of an autoantibody against AQP4 found in patients with an inflammatory demyelinating disease of the central nervous system termed neuromyelitis optica, termed AQP4-IgG or NMO-IgG [74]. AQP4-IgG is pathogenic, by binding AQP4 on astrocytes and causing complement-dependent astrocyte damage, followed by leukocyte infiltration into the lesion. Monoclonal AQP4-IgG can now be produced artificially [75]. AQP4-IgG linked to a

toxin could be used for destroying AQP4-expressing glioblastoma cells, most of which express large amounts of AQP4. However, it is unclear whether eliminating the AQP4-expressing subpopulation of tumor cells will render the glioblastoma less aggressive. Local AQP4-IgG delivery in human glioblastoma is possible by using wafers positioned against the resection cavity wall or by convection-enhanced delivery. The side effect of AQP4-IgG treatment may be neuromyelitis optica type symptoms caused by AQP4-IgG-mediated damage to normal astrocytes. A more elegant approach is to link AQP4-IgG to a toxin that becomes activated when AQP4-IgG is internalized (Fig. 3). There is evidence that intact astrocytes (which express AQP4 in the perivascular foot processes) do not internalize AQP4-IgG, but cells that express AQP4 throughout the plasma cell membrane (such as glioblastoma) internalize AQP4-IgG [76]. This observation suggests that, compared with normal astrocytes, AQP4-expressing glioblastoma cells may be selectively vulnerable to damage by AQP4-IgG internalization, thus reducing the side effects of this treatment.

5. Future directions

Several roles of AQPs in tumor biology are beginning to emerge. There is a need to define further the molecular mechanisms responsible for AQP-mediated cell migration and cell proliferation. It is unclear whether the water transporting property of AQPs is important or whether there are as yet unidentified interactions between AQPs and oncogenes. Further research is also required to discover non-toxic AQP inhibitors, which can be used to define AQP functions and as novel cancer therapeutics. AQP inhibitors, which could target tumor infiltration, metastasis and angiogenesis, might be used in combination with current cancer therapeutics that target the proliferating tumor cells.

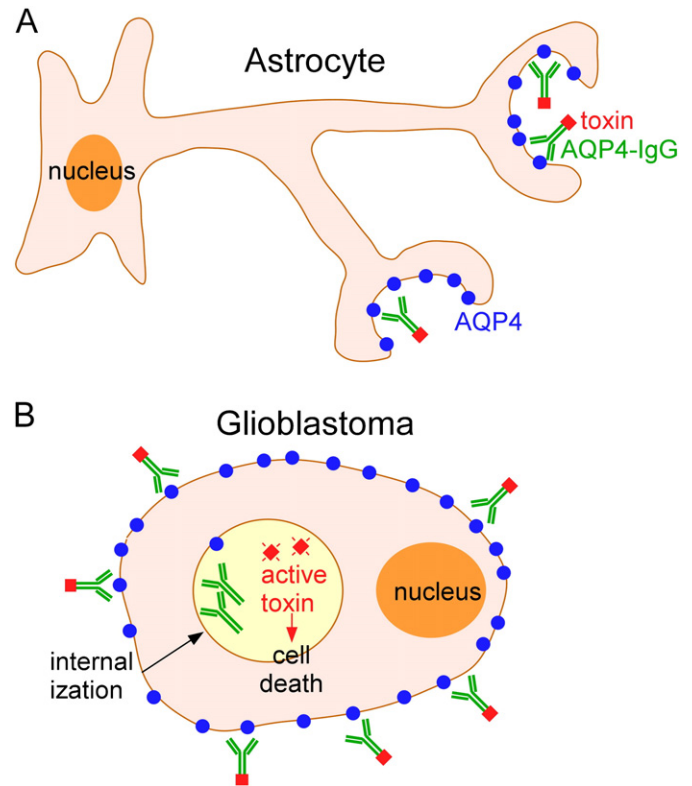


Fig. 3. AQP4-IgG linked to toxin as a potential treatment for glioblastoma. A. AQP4-IgG binds AQP4 on astrocyte foot processes and is not internalized. Toxin remains linked to AQP4-IgG. B. AQP4-IgG binds AQP4 on glioblastoma cell and becomes internalized. Toxin is released from AQP4-IgG and kills the cell.

6. Conclusion

Over the last few years there emerged a surprising link between AQP's and cancer. AQP's appear to play a key role in several tumor-related processes including tumor edema, tumor cell migration, tumor proliferation and angiogenesis. AQP inhibitors may thus be useful anti-cancer drugs. Unfortunately, no such inhibitors are available to date.

Acknowledgments

MCP is funded by the Guthy Jackson Charitable Foundation and the Neurosciences Research Foundation.

References

- [1] A.S. Verkman, Aquaporins in clinical medicine, *Annu. Rev. Med.* 63 (2012) 303–316.
- [2] A. Rojek, J. Praetorius, J. Frokiaer, S. Nielsen, R.A. Fenton, A current view of the mammalian aquaglyceroporins, *Annu. Rev. Physiol.* 70 (2008) 301–327.
- [3] B. Wu, E. Beitz, Aquaporins with selectivity for unconventional permeants, *Cell. Mol. Life Sci.* 64 (2007) 2413–2421.
- [4] M. Herrera, J.L. Garvin, Aquaporins as gas channels, *Pflugers Arch.* 462 (2011) 623–630.
- [5] T. Walz, Y. Fujiyoshi, A. Engel, The AQP structure and functional implications, *Handb. Exp. Pharmacol.* (2009) 31–56.
- [6] B.J. Jin, A. Rossi, A.S. Verkman, Model of aquaporin-4 supramolecular assembly in orthogonal arrays based on heterotetrameric association of M1–M23 isoforms, *Biophys. J.* 100 (2011) 2936–2945.
- [7] J.M. Crane, A.S. Verkman, Long-range nonanomalous diffusion of quantum dot-labeled aquaporin-1 water channels in the cell plasma membrane, *Biophys. J.* 94 (2008) 702–713.
- [8] A.S. Verkman, A. Rossi, J.M. Crane, Live-cell imaging of aquaporin-4 supramolecular assembly and diffusion, *Methods Enzymol.* 504 (2012) 341–354.
- [9] J.E. Rash, T. Yasumura, C.S. Hudson, P. Agre, S. Nielsen, Direct immunogold labeling of aquaporin-4 in square arrays of astrocyte and ependymocyte plasma membranes in rat brain and spinal cord, *Proc. Natl. Acad. Sci. U. S. A.* 95 (1998) 11981–11986.
- [10] J.D. Neely, M. Amiry-Moghadam, O.P. Ottersen, S.C. Froehner, P. Agre, M.E. Adams, Syntrophin-dependent expression and localization of aquaporin-4 water channel protein, *Proc. Natl. Acad. Sci. U. S. A.* 98 (2001) 14108–14113.
- [11] S. Noell, P. Fallier-Becker, U. Deutsch, A.F. Mack, H. Wolburg, Agrin defines polarized distribution of orthogonal arrays of particles in astrocytes, *Cell Tissue Res.* 337 (2009) 185–195.
- [12] Y. Noda, E. Sohara, E. Ohta, S. Sasaki, Aquaporins in kidney pathophysiology, *Nat. Rev. Nephrol.* 6 (2010) 168–178.
- [13] L. Yang, B.T. Kress, H.J. Weber, M. Thiyagarajan, B. Wang, R. Deane, H. Benveniste, J.J. Iliff, M. Nedergaard, Evaluating glymphatic pathway function utilizing clinically relevant intrathecal infusion of CSF tracer, *J. Transl. Med.* 11 (2013) 107.
- [14] M.C. Papadopoulos, S. Saadoun, A.S. Verkman, Aquaporins and cell migration, *Pflugers Arch.* 456 (2008) 693–700.
- [15] A.S. Verkman, M. Hara-Chikuma, M.C. Papadopoulos, Aquaporins—new players in cancer biology, *J. Mol. Med. (Berlin)* 86 (2008) 523–529.
- [16] A.J. Yool, E.A. Brown, G.A. Flynn, Roles for novel pharmacological blockers of aquaporins in the treatment of brain oedema and cancer, *Clin. Exp. Pharmacol. Physiol.* 37 (2010) 403–409.
- [17] E. Monzani, A.A. Shtil, C.A. La Porta, The water channels, new druggable targets to combat cancer cell survival, invasiveness and metastasis, *Curr. Drug Targets* 8 (2007) 1132–1137.
- [18] B. Nico, D. Ribatti, Aquaporins in tumor growth and angiogenesis, *Cancer Lett.* 294 (2010) 135–138.
- [19] D. Ribatti, G. Ranieri, T. Annese, B. Nico, Aquaporins in cancer, *Biochim. Biophys. Acta* 1840 (2014) 1550–1553.
- [20] M.C. Papadopoulos, S. Saadoun, D.K. Binder, G.T. Manley, S. Krishna, A.S. Verkman, Molecular mechanisms of brain tumor edema, *Neuroscience* 129 (2004) 1011–1020.
- [21] S. Saadoun, M.C. Papadopoulos, D.C. Davies, S. Krishna, B.A. Bell, Aquaporin-4 expression is increased in oedematous human brain tumours, *J. Neurol. Neurosurg. Psychiatry* 72 (2002) 262–265.
- [22] S. Saadoun, M.C. Papadopoulos, D.C. Davies, B.A. Bell, S. Krishna, Increased aquaporin 1 water channel expression in human brain tumours, *Br. J. Cancer* 87 (2002) 621–623.
- [23] S. Jelen, B. Parm Ulhoi, A. Larsen, J. Frokiaer, S. Nielsen, M. Rutzler, AQP9 expression in glioblastoma multiforme tumors is limited to a small population of astrocytic cells and CD15(+) / CalB(+) leukocytes, *PLoS One* 8 (2013) e75764.
- [24] S.J. Zhu, K.J. Wang, S.W. Gan, J. Xu, S.Y. Xu, S.Q. Sun, Expression of aquaporin8 in human astrocytomas: correlation with pathologic grade, *Biochem. Biophys. Res. Commun.* 440 (2013) 168–172.
- [25] N. El Hindy, A. Bankfalvi, A. Herring, M. Adamzik, N. Lambert, Y. Zhu, W. Siffert, U. Sure, I.E. Sandalcioglu, Correlation of aquaporin-1 water channel protein expression with tumor angiogenesis in human astrocytoma, *Anticancer Res.* 33 (2013) 609–613.
- [26] P. Deb, S. Pal, V. Dutta, D. Boruah, V.M. Chandran, H.S. Bhatoo, Correlation of expression pattern of aquaporin-1 in primary central nervous system tumors with tumor type, grade, proliferation, microvessel density, contrast-enhancement and perilesional edema, *J. Cancer Res. Ther.* 8 (2012) 571–577.
- [27] R.K. Dua, B.I. Devi, T.C. Yasha, Increased expression of Aquaporin-4 and its correlation with contrast enhancement and perilesional edema in brain tumors, *Br. J. Neurosurg.* 24 (2010) 454–459.
- [28] G. Tan, S.Q. Sun, D.L. Yuan, Expression of the water channel protein aquaporin-9 in human astrocytic tumours: correlation with pathological grade, *J. Int. Med. Res.* 36 (2008) 777–782.
- [29] A. Warth, M. Mittelbronn, P. Hulper, B. Erdlenbruch, H. Wolburg, Expression of the water channel protein aquaporin-9 in malignant brain tumors, *Appl. Immunohistochem. Mol. Morphol.* 15 (2007) 193–198.
- [30] A. Warth, S. Kroger, H. Wolburg, Redistribution of aquaporin-4 in human glioblastoma correlates with loss of agrin immunoreactivity from brain capillary basal laminae, *Acta Neuropathol.* 107 (2004) 311–318.
- [31] A. Warth, P. Simon, D. Capper, B. Goepfert, G. Tabatabai, H. Herzog, K. Dietz, F. Stubenvoll, R. Ajaaj, R. Becker, M. Weller, R. Meyermann, H. Wolburg, M. Mittelbronn, Expression pattern of the water channel aquaporin-4 in human gliomas is associated with blood–brain barrier disturbance but not with patient survival, *J. Neurosci. Res.* 85 (2007) 1336–1346.
- [32] K. Oshio, D.K. Binder, Y. Liang, A. Bollen, B. Feuerstein, M.S. Berger, G.T. Manley, Expression of the aquaporin-1 water channel in human glial tumors, *Neurosurgery* 56 (2005) 375–381 (discussion 375–381).
- [33] B. Nico, D. Mangieri, R. Tamma, V. Longo, T. Annese, E. Crivellato, B. Pollo, E. Maderna, D. Ribatti, A. Salmaggi, Aquaporin-4 contributes to the resolution of peritumoral brain oedema in human glioblastoma multiforme after combined chemotherapy and radiotherapy, *Eur. J. Cancer* 45 (2009) 3315–3325.
- [34] J.H. Yang, C.X. Yan, X.J. Chen, Y.S. Zhu, Expression of aquaglyceroporins in epithelial ovarian tumours and their clinical significance, *J. Int. Med. Res.* 39 (2011) 702–711.
- [35] N.N. Haj-Yasein, G.F. Vindedal, M. Eilert-Olsen, G.A. Gundersen, O. Skare, P. Laake, A. Klungland, A.E. Thoren, J.M. Burkhardt, O.P. Ottersen, E.A. Nagelhus, Glial-conditional deletion of aquaporin-4 (AQP4) reduces blood–brain water uptake and confers barrier function on perivascular astrocyte endfeet, *Proc. Natl. Acad. Sci. U. S. A.* 108 (2011) 17815–17820.
- [36] A.S. Verkman, M.O. Anderson, M.C. Papadopoulos, Aquaporins: important but elusive drug targets, *Nat. Rev. Drug Discov.* 13 (2014) 259–277.
- [37] M.C. Papadopoulos, A.S. Verkman, Aquaporin water channels in the nervous system, *Nat. Rev. Neurosci.* 14 (2013) 265–277.
- [38] T. Ma, Y. Song, A. Gillespie, E.J. Carlson, C.J. Epstein, A.S. Verkman, Defective secretion of saliva in transgenic mice lacking aquaporin-5 water channels, *J. Biol. Chem.* 274 (1999) 20071–20074.
- [39] M. Moore, T. Ma, B. Yang, A.S. Verkman, Tear secretion by lacrimal glands in transgenic mice lacking water channels AQP1, AQP3, AQP4 and AQP5, *Exp. Eye Res.* 70 (2000) 557–562.
- [40] M.C. Papadopoulos, G.T. Manley, S. Krishna, A.S. Verkman, Aquaporin-4 facilitates reabsorption of excess fluid in vasogenic brain edema, *FASEB J.* 18 (2004) 1291–1293.
- [41] V.M. Loitto, T. Forslund, T. Sundqvist, K.E. Magnusson, M. Gustafsson, Neutrophil leukocyte motility requires directed water influx, *J. Leukoc. Biol.* 71 (2002) 212–222.
- [42] S. Saadoun, M.C. Papadopoulos, M. Hara-Chikuma, A.S. Verkman, Impairment of angiogenesis and cell migration by targeted aquaporin-1 gene disruption, *Nature* 434 (2005) 786–792.
- [43] S. Saadoun, M.C. Papadopoulos, H. Watanabe, D. Yan, G.T. Manley, A.S. Verkman, Involvement of aquaporin-4 in astroglial cell migration and glial scar formation, *J. Cell Sci.* 118 (2005) 5691–5698.
- [44] J. Ruiz-Ederra, A.S. Verkman, Aquaporin-1-facilitated keratocyte migration in cell culture and in vivo corneal wound healing models, *Exp. Eye Res.* 89 (2009) 159–165.
- [45] M. Hara-Chikuma, A.S. Verkman, Aquaporin-1 facilitates epithelial cell migration in kidney proximal tubule, *J. Am. Soc. Nephrol.* 17 (2006) 39–45.
- [46] M. Hara-Chikuma, A.S. Verkman, Aquaporin-3 facilitates epidermal cell migration and proliferation during wound healing, *J. Mol. Med. (Berlin)* 86 (2008) 221–231.
- [47] K.I. Auguste, S. Jin, K. Uchida, D. Yan, G.T. Manley, M.C. Papadopoulos, A.S. Verkman, Greatly impaired migration of implanted aquaporin-4 deficient astroglial cells in mouse brain toward a site of injury, *FASEB J.* 21 (2007) 108–116.
- [48] S. Bisi, A. Disanza, C. Malinverno, E. Frittoli, A. Palamidessi, G. Scita, Membrane and actin dynamics interplay at lamellipodia leading edge, *Curr. Opin. Cell Biol.* 25 (2013) 565–573.
- [49] K.M. Stroka, H. Jiang, S.H. Chen, Z. Tong, D. Wirtz, S.X. Sun, K. Konstantopoulos, Water permeation drives tumor cell migration in confined microenvironments, *Cell* 157 (2014) 611–623.
- [50] J. Hu, A.S. Verkman, Increased migration and metastatic potential of tumor cells expressing aquaporin water channels, *FASEB J.* 20 (2006) 1892–1894.
- [51] G.P. Nicchia, C. Stigliano, A. Sparaneo, A. Rossi, A. Frigeri, M. Svelto, Inhibition of aquaporin-1 dependent angiogenesis impairs tumour growth in a mouse model of melanoma, *J. Mol. Med. (Berlin)* 91 (2013) 613–623.
- [52] C. Esteva-Font, B.J. Jin, A.S. Verkman, Aquaporin-1 gene deletion reduces breast tumor growth and lung metastasis in tumor-producing MMTV-PyVT mice, *FASEB J.* 28 (2014) 1446–1453.
- [53] H. Yokomori, M. Oda, K. Yoshimura, F. Kaneko, T. Hibi, Aquaporin-1 associated with hepatic arterial capillary proliferation on hepatic sinusoid in human cirrhotic liver, *Liver Int.* 31 (2011) 1554–1564.

- [54] R.C. Huebert, K. Jagavelu, H.I. Hendrickson, M.M. Vasdev, J.P. Arab, P.L. Splinter, C.E. Trussoni, N.F. Larusso, V.H. Shah, Aquaporin-1 promotes angiogenesis, fibrosis, and portal hypertension through mechanisms dependent on osmotically sensitive microRNAs, *Am. J. Pathol.* 179 (2011) 1851–1860.
- [55] J. Ruiz-Ederra, A.S. Verkman, Aquaporin-1 independent microvessel proliferation in a neonatal mouse model of oxygen-induced retinopathy, *Invest. Ophthalmol. Vis. Sci.* 48 (2007) 4802–4810.
- [56] X. Trepat, Z. Chen, K. Jacobson, Cell migration, *Compr. Physiol.* 2 (2012) 2369–2392.
- [57] R.J. Petrie, K.M. Yamada, At the leading edge of three-dimensional cell migration, *J. Cell Sci.* 125 (2012) 5917–5926.
- [58] M. Hara-Chikuma, K. Takahashi, S. Chikuma, A.S. Verkman, Y. Miyachi, The expression of differentiation markers in aquaporin-3 deficient epidermis, *Arch. Dermatol. Res.* 301 (2009) 245–252.
- [59] M. Hara, T. Ma, A.S. Verkman, Selectively reduced glycerol in skin of aquaporin-3-deficient mice may account for impaired skin hydration, elasticity, and barrier recovery, *J. Biol. Chem.* 277 (2002) 46616–46621.
- [60] S.K. Kang, Y.K. Chae, J. Woo, M.S. Kim, J.C. Park, J. Lee, J.C. Soria, S.J. Jang, D. Sidransky, C. Moon, Role of human aquaporin 5 in colorectal carcinogenesis, *Am. J. Pathol.* 173 (2008) 518–525.
- [61] Z. Zhang, Z. Chen, Y. Song, P. Zhang, J. Hu, C. Bai, Expression of aquaporin 5 increases proliferation and metastasis potential of lung cancer, *J. Pathol.* 221 (2010) 210–220.
- [62] M. Hara-Chikuma, A.S. Verkman, Prevention of skin tumorigenesis and impairment of epithelial cell proliferation by targeted aquaporin-3 gene disruption, *Mol. Cell. Biol.* 28 (2008) 326–332.
- [63] A.S. Verkman, A cautionary note on cosmetics containing ingredients that increase aquaporin-3 expression, *Exp. Dermatol.* 17 (2008) 871–872.
- [64] Y.L. Liu, T. Matsuzaki, T. Nakazawa, S. Murata, N. Nakamura, T. Kondo, M. Iwashina, K. Mochizuki, T. Yamane, K. Takata, R. Katoh, Expression of aquaporin 3 (AQP3) in normal and neoplastic lung tissues, *Hum. Pathol.* 38 (2007) 171–178.
- [65] H. Xia, Y.F. Ma, C.H. Yu, Y.J. Li, J. Tang, J.B. Li, Y.N. Zhao, Y. Liu, Aquaporin 3 knock-down suppresses tumour growth and angiogenesis in experimental non-small cell lung cancer, *Exp. Physiol.* 99 (2014) 974–984.
- [66] Y. Hiroaki, K. Tani, A. Kamegawa, N. Gyobu, K. Nishikawa, H. Suzuki, T. Walz, S. Sasaki, K. Mitsuoka, K. Kimura, A. Mizoguchi, Y. Fujiyoshi, Implications of the aquaporin-4 structure on array formation and cell adhesion, *J. Mol. Biol.* 355 (2006) 628–639.
- [67] H. Zhang, A.S. Verkman, Evidence against involvement of aquaporin-4 in cell-cell adhesion, *J. Mol. Biol.* 382 (2008) 1136–1143.
- [68] A.J. Smith, B.J. Jin, J. Ratelade, A.S. Verkman, Aggregation state determines the localization and function of M1- and M23-aquaporin-4 in astrocytes, *J. Cell Biol.* 204 (2014) 559–573.
- [69] M.J. Tait, V. Petrik, A. Loosmore, B.A. Bell, M.C. Papadopoulos, Survival of patients with glioblastoma multiforme has not improved between 1993 and 2004: analysis of 625 cases, *Br. J. Neurosurg.* 21 (2007) 496–500.
- [70] U. Pichlmeier, A. Bink, G. Schackert, W. Stummer, Resection and survival in glioblastoma multiforme: an RTOG recursive partitioning analysis of ALA study patients, *Neuro Oncol.* 10 (2008) 1025–1034.
- [71] H. Igarashi, V.J. Huber, M. Tsujita, T. Nakada, Pretreatment with a novel aquaporin 4 inhibitor, TGN-020, significantly reduces ischemic cerebral edema, *Neurosci. Sci.* 32 (2011) 113–116.
- [72] V.J. Huber, M. Tsujita, T. Nakada, Aquaporins in drug discovery and pharmacotherapy, *Mol. Aspects Med.* 33 (2012) 691–703.
- [73] V.J. Huber, M. Tsujita, T. Nakada, Identification of aquaporin 4 inhibitors using in vitro and in silico methods, *Bioorg. Med. Chem.* 17 (2009) 411–417.
- [74] M.C. Papadopoulos, A.S. Verkman, Aquaporin 4 and neuromyelitis optica, *Lancet Neurol.* 11 (2012) 535–544.
- [75] J.L. Bennett, C. Lam, S.R. Kalluri, P. Saikali, K. Bautista, C. Dupree, M. Glogowska, D. Case, J.P. Antel, G.P. Owens, D. Gilden, S. Nessler, C. Stadelmann, B. Hemmer, Intrathecal pathogenic anti-aquaporin-4 antibodies in early neuromyelitis optica, *Ann. Neurol.* 66 (2009) 617–629.
- [76] J. Ratelade, J.L. Bennett, A.S. Verkman, Evidence against cellular internalization in vivo of NMO-IgG, aquaporin-4, and excitatory amino acid transporter 2 in neuromyelitis optica, *J. Biol. Chem.* 286 (2011) 45156–45164.
- [77] H.J. Jung, J.Y. Park, H.S. Jeon, T.H. Kwon, Aquaporin-5: a marker protein for proliferation and migration of human breast cancer cells, *PLoS One* 6 (2011) e28492.
- [78] S.J. Lee, Y.S. Chae, J.G. Kim, W.W. Kim, J.H. Jung, H.Y. Park, J.Y. Jeong, J.Y. Park, H.J. Jung, T.H. Kwon, AQP5 expression predicts survival in patients with early breast cancer, *Ann. Surg. Oncol.* 21 (2014) 375–383.
- [79] Z. Shi, T. Zhang, L. Luo, H. Zhao, J. Cheng, J. Xiang, C. Zhao, Aquaporins in human breast cancer: identification and involvement in carcinogenesis of breast cancer, *J. Surg. Oncol.* 106 (2012) 267–272.
- [80] S. Aishima, Y. Kuroda, Y. Nishihara, K. Taguchi, T. Iguchi, A. Taketomi, Y. Maehara, M. Tsuneyoshi, Down-regulation of aquaporin-1 in intrahepatic cholangiocarcinoma is related to tumor progression and mucin expression, *Hum. Pathol.* 38 (2007) 1819–1825.
- [81] P.R. Mazal, M. Susani, F. Wrba, A. Haitel, Diagnostic significance of aquaporin-1 in liver tumors, *Hum. Pathol.* 36 (2005) 1226–1231.
- [82] T. Yoshida, S. Hojo, S. Sekine, S. Sawada, T. Okumura, T. Nagata, Y. Shimada, K. Tsukada, Expression of aquaporin-1 is a poor prognostic factor for stage II and III colon cancer, *Mol. Clin. Oncol.* 1 (2013) 953–958.
- [83] C. Moon, J.C. Soria, S.J. Jang, J. Lee, M. Obaidul Hoque, M. Sibony, B. Trink, Y.S. Chang, D. Sidransky, L. Mao, Involvement of aquaporins in colorectal carcinogenesis, *Oncogene* 22 (2003) 6699–6703.
- [84] X. Shi, S. Wu, Y. Yang, L. Tang, Y. Wang, J. Dong, B. Lu, G. Jiang, W. Zhao, AQP5 silencing suppresses p38 MAPK signaling and improves drug resistance in colon cancer cells, *Tumour Biol.* 35 (2014) 7035–7045.
- [85] W. Wang, Q. Li, T. Yang, G. Bai, D. Li, Q. Li, H. Sun, Expression of AQP5 and AQP8 in human colorectal carcinoma and their clinical significance, *World J. Surg. Oncol.* 10 (2012) 242.
- [86] H. Fischer, R. Stenling, C. Rubio, A. Lindblom, Differential expression of aquaporin 8 in human colonic epithelial cells and colorectal tumors, *BMC Physiol.* 1 (2001) 1.
- [87] R. Chen, Y. Shi, R. Amiduo, T. Tuokan, L. Suzuk, Expression and prognostic value of aquaporin 1, 3 in cervical carcinoma in women of Uygur ethnicity from Xinjiang, China, *PLoS One* 9 (2014) e98576.
- [88] T. Zhang, C. Zhao, D. Chen, Z. Zhou, Overexpression of AQP5 in cervical cancer: correlation with clinicopathological features and prognosis, *Med. Oncol.* 29 (2012) 1998–2004.
- [89] P. Longatti, L. Basaldella, E. Orvieto, A. Dei Tos, A. Martinuzzi, Aquaporin(s) expression in choroid plexus tumours, *Pediatr. Neurosurg.* 42 (2006) 228–233.
- [90] Y. Chen, O. Tachibana, M. Oda, R. Xu, J. Hamada, J. Yamashita, N. Hashimoto, J.A. Takahashi, Increased expression of aquaporin 1 in human hemangioblastomas and its correlation with cyst formation, *J. Neurooncol.* 80 (2006) 219–225.
- [91] B. Guan, D. Zhu, Z. Dong, Z. Yang, Expression and distribution of aquaporin 1 in laryngeal carcinoma, *Lin Chung Er Bi Yan Hou Tou Jing Wai Ke Za Zhi* 21 (2007) 269–272.
- [92] Y.K. Chae, S.K. Kang, M.S. Kim, J. Woo, J. Lee, S. Chang, D.W. Kim, M. Kim, S. Park, I. Kim, B. Keam, J. Rhee, N.H. Koo, G. Park, S.H. Kim, S.E. Jang, I.Y. Kwon, D. Sidransky, C. Moon, Human AQP5 plays a role in the progression of chronic myelogenous leukemia (CML), *PLoS One* 3 (2008) e2594.
- [93] X. Guo, T. Sun, M. Yang, Z. Li, Z. Li, Y. Gao, Prognostic value of combined aquaporin 3 and aquaporin 5 overexpression in hepatocellular carcinoma, *Biomed. Res. Int.* 2013 (2013) 206525.
- [94] E.M. Jablonski, M.A. Mattocks, E. Sokolov, L.G. Koniaris, F.M. Hughes Jr., N. Fausto, R.H. Pierce, I.H. McKillop, Decreased aquaporin expression leads to increased resistance to apoptosis in hepatocellular carcinoma, *Cancer Lett.* 250 (2007) 36–46.
- [95] Y. Machida, Y. Ueda, M. Shimasaki, K. Sato, M. Sagawa, S. Katsuda, T. Sakuma, Relationship of aquaporin 1, 3, and 5 expression in lung cancer cells to cellular differentiation, invasive growth, and metastasis potential, *Hum. Pathol.* 42 (2011) 669–678.
- [96] M.O. Hoque, J.C. Soria, J. Woo, T. Lee, J. Lee, S.J. Jang, S. Upadhyay, B. Trink, C. Monitto, C. Desmaze, L. Mao, D. Sidransky, C. Moon, Aquaporin 1 is overexpressed in lung cancer and stimulates NIH-3 T3 cell proliferation and anchorage-independent growth, *Am. J. Pathol.* 168 (2006) 1345–1353.
- [97] Y. Xie, X. Wen, Z. Jiang, H.Q. Fu, H. Han, L. Dai, Aquaporin 1 and aquaporin 4 are involved in invasion of lung cancer cells, *Clin. Lab.* 58 (2012) 75–80.
- [98] P. Wang, R.Y. Ni, M.N. Chen, K.J. Mou, Q. Mao, Y.H. Liu, Expression of aquaporin-4 in human supratentorial meningiomas with peritumoral brain edema and correlation of VEGF with edema formation, *Genet. Mol. Res.* 10 (2011) 2165–2171.
- [99] W.H. Ng, J.W. Hy, W.L. Tan, D. Liew, T. Lim, B.T. Ang, I. Ng, Aquaporin-4 expression is increased in edematous meningiomas, *J. Clin. Neurosci.* 16 (2009) 441–443.
- [100] Q. Li, B. Zhang, Expression of aquaporin-1 in nasopharyngeal cancer tissues, *J. Otolaryngol. Head Neck Surg.* 39 (2010) 511–515.
- [101] S. Liu, S. Zhang, H. Jiang, Y. Yang, Y. Jiang, Co-expression of AQP3 and AQP5 in esophageal squamous cell carcinoma correlates with aggressive tumor progression and poor prognosis, *Med. Oncol.* 30 (2013) 636.
- [102] M. Kusayama, K. Wada, M. Nagata, S. Ishimoto, H. Takahashi, M. Yoneda, A. Nakajima, M. Okura, M. Kogo, Y. Kamisaki, Critical role of aquaporin 3 on growth of human esophageal and oral squamous cell carcinoma, *Cancer Sci.* 102 (2011) 1128–1136.
- [103] J.H. Yang, Y.Q. Yu, C.X. Yan, Localisation and expression of aquaporin subtypes in epithelial ovarian tumours, *Histol. Histopathol.* 26 (2011) 1197–1205.
- [104] J.H. Yang, Y.F. Shi, Q. Cheng, L. Deng, Expression and localization of aquaporin-5 in the epithelial ovarian tumors, *Gynecol. Oncol.* 100 (2006) 294–299.
- [105] H. Kafe, J.M. Verbavatz, B. Cochand-Priollet, P. Castagnet, A. Vieillefond, Collecting duct carcinoma: an entity to be redefined? *Virchows Arch.* 445 (2004) 637–640.
- [106] D. Niu, T. Kondo, T. Nakazawa, T. Yamane, K. Mochizuki, T. Kawasaki, T. Matsuzaki, K. Takata, R. Katoh, Expression of aquaporin3 in human neoplastic tissues, *Histopathology* 61 (2012) 543–551.
- [107] T. Watanabe, T. Fujii, T. Oya, N. Horikawa, Y. Tabuchi, Y. Takahashi, M. Morii, N. Takeguchi, K. Tsukada, H. Sakai, Involvement of aquaporin-5 in differentiation of human gastric cancer cells, *J. Physiol. Sci.* 59 (2009) 113–122.
- [108] H. Xu, Y. Zhang, W. Wei, L. Shen, W. Wu, Differential expression of aquaporin-4 in human gastric normal and cancer tissues, *Gastroenterol. Clin. Biol.* 33 (2009) 72–76.
- [109] J. Chen, T. Wang, Y.C. Zhou, F. Gao, Z.H. Zhang, H. Xu, S.L. Wang, L.Z. Shen, Aquaporin 3 promotes epithelial-mesenchymal transition in gastric cancer, *J. Exp. Clin. Cancer Res.* 33 (2014) 38.
- [110] D. Niu, T. Kondo, T. Nakazawa, T. Kawasaki, T. Yamane, K. Mochizuki, Y. Kato, T. Matsuzaki, K. Takata, R. Katoh, Differential expression of aquaporins and its diagnostic utility in thyroid cancer, *PLoS One* 7 (2012) e40770.
- [111] L. Lacroix, V. Lazar, S. Michiels, H. Ripoché, P. Dessen, M. Talbot, B. Caillou, J.P. Levilain, M. Schlumberger, J.M. Bidart, Follicular thyroid tumors with the PAX8-PPARGamma1 rearrangement display characteristic genetic alterations, *Am. J. Pathol.* 167 (2005) 223–231.
- [112] S. Ishimoto, K. Wada, Y. Usami, N. Tanaka, T. Aikawa, M. Okura, A. Nakajima, M. Kogo, Y. Kamisaki, Differential expression of aquaporin 5 and aquaporin 3 in squamous cell carcinoma and adenoid cystic carcinoma, *Int. J. Oncol.* 41 (2012) 67–75.

- [113] P.C. Rubenwolf, W. Otto, S. Denzinger, F. Hofstadter, W. Wieland, N.T. Georgopoulos, Expression of aquaporin water channels in human urothelial carcinoma: correlation of AQP3 expression with tumour grade and stage, *World J. Urol.* 31 (2013) 991–997.
- [114] W. Otto, P.C. Rubenwolf, M. Burger, H.M. Fritsche, W. Rossler, M. May, A. Hartmann, F. Hofstadter, W.F. Wieland, S. Denzinger, Loss of aquaporin 3 protein expression constitutes an independent prognostic factor for progression-free survival: an immunohistochemical study on stage pT1 urothelial bladder cancer, *BMC Cancer* 12 (2012) 459.