

Optical Imaging for Neuroscience and Developmental Biology

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LEHIGH
UNIVERSITY.

Outline

•Introduction

- Biomedical Imaging Modalities
- Why Optical Imaging?
- Optical Biopsy with Optical Coherence Tomography (OCT) and Microscopy (OCM)

•Applications in Neuroscience and Developmental Biology

- 3D imaging of brain slices
- Evaluate heart function in fruit flies

•Summary

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BIOMEDICAL IMAGING MODALITIES

- ✓ X-Ray
 - ✓ Computed Tomography (CT)
 - ✓ Positron Emission Tomography (PET)
 - ✓ Magnetic Resonance Imaging (MRI)
 - ✓ Ultrasonography (US)
 - ✓ Optical Imaging
-

X-RAY

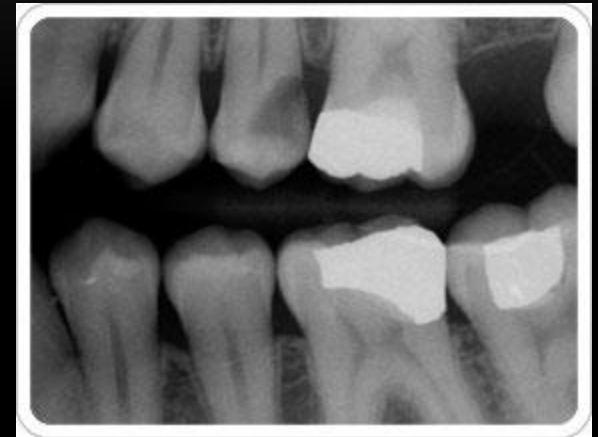
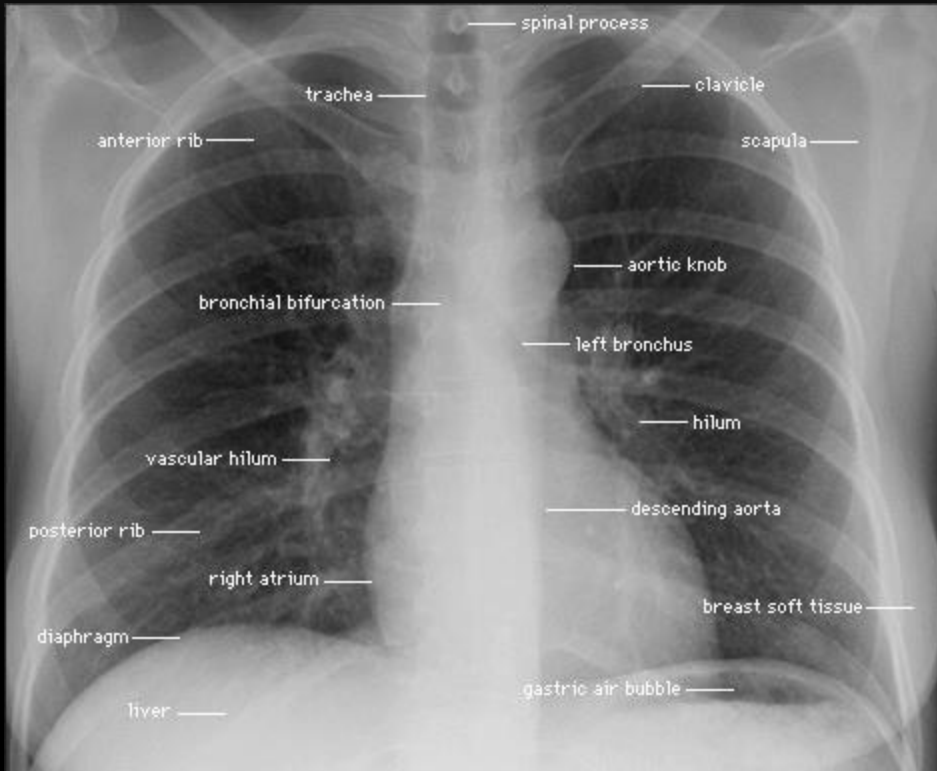


- ✓ Discovered in 1895 by Wilhelm Conrad Röntgen, who received the first Nobel Prize in Physics in 1901.

X-RAY



X-RAY



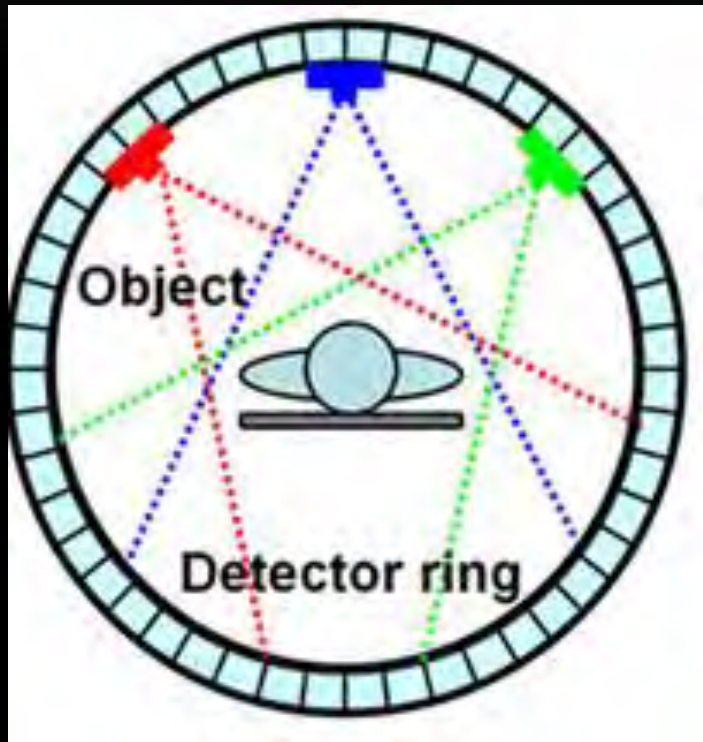
Ionizing radiation

COMPUTED TOMOGRAPHY (CT)

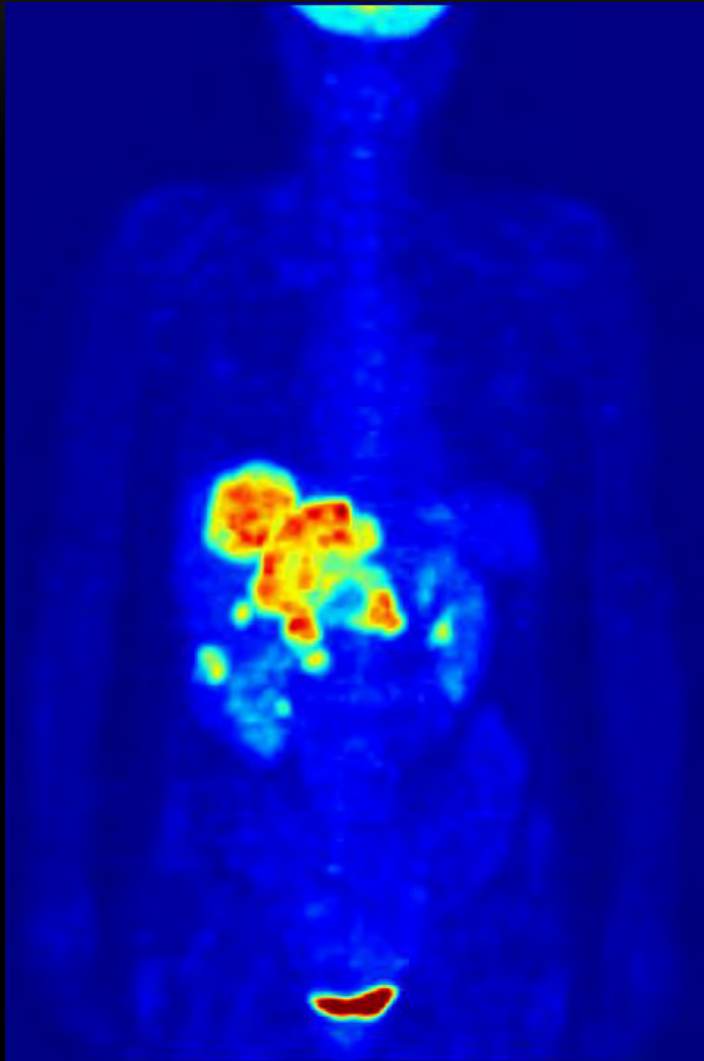


- ✓ Invented in 1971 by Allan Cormack and Godfrey Hounsfield, who shared the 1979 Nobel Prize for Physiology or Medicine

COMPUTED TOMOGRAPHY (CT)

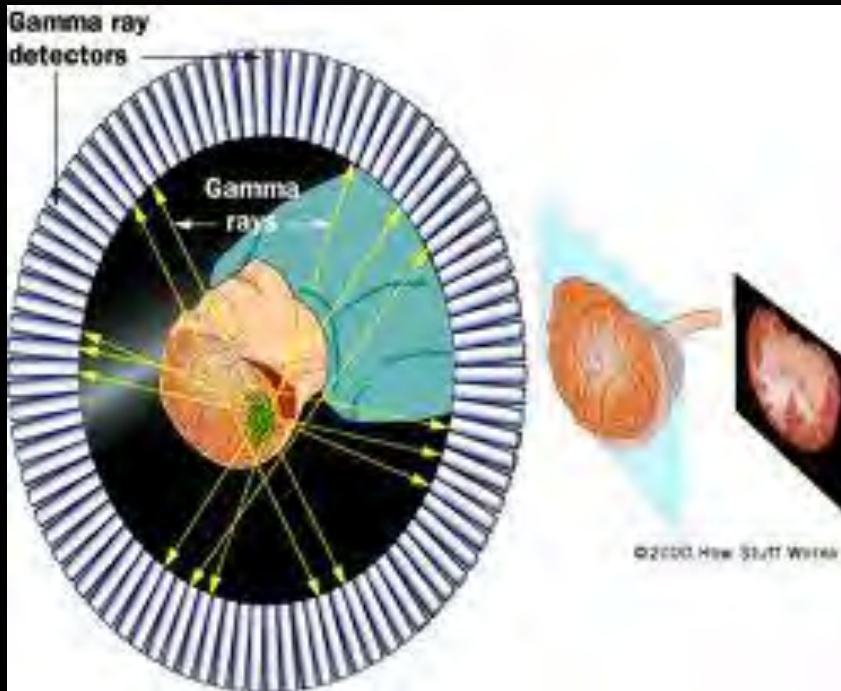


POSITRON EMISSION TOMOGRAPHY (PET)



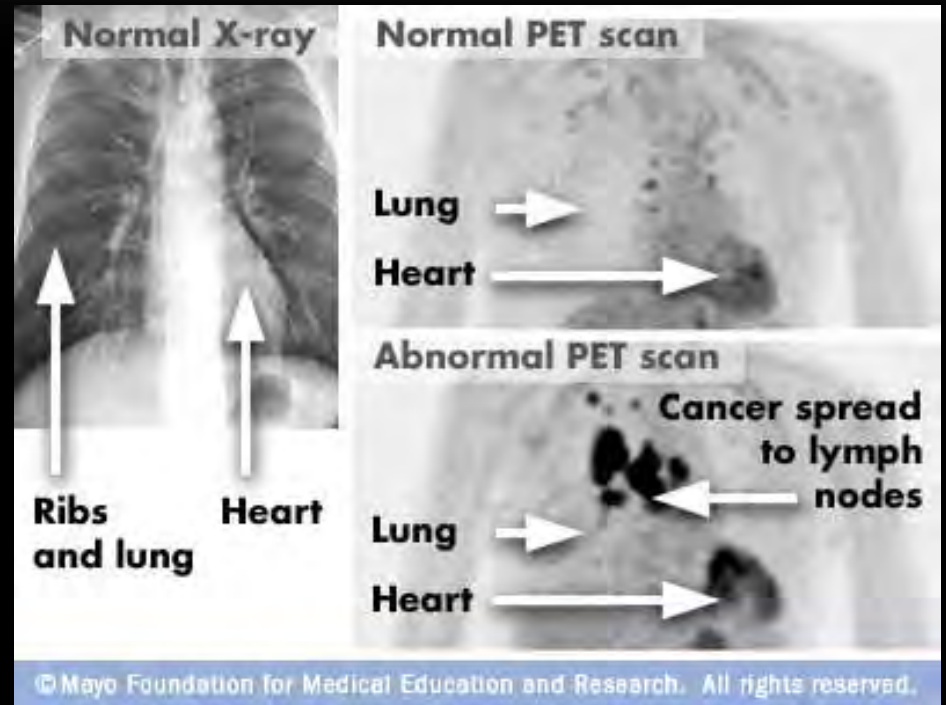
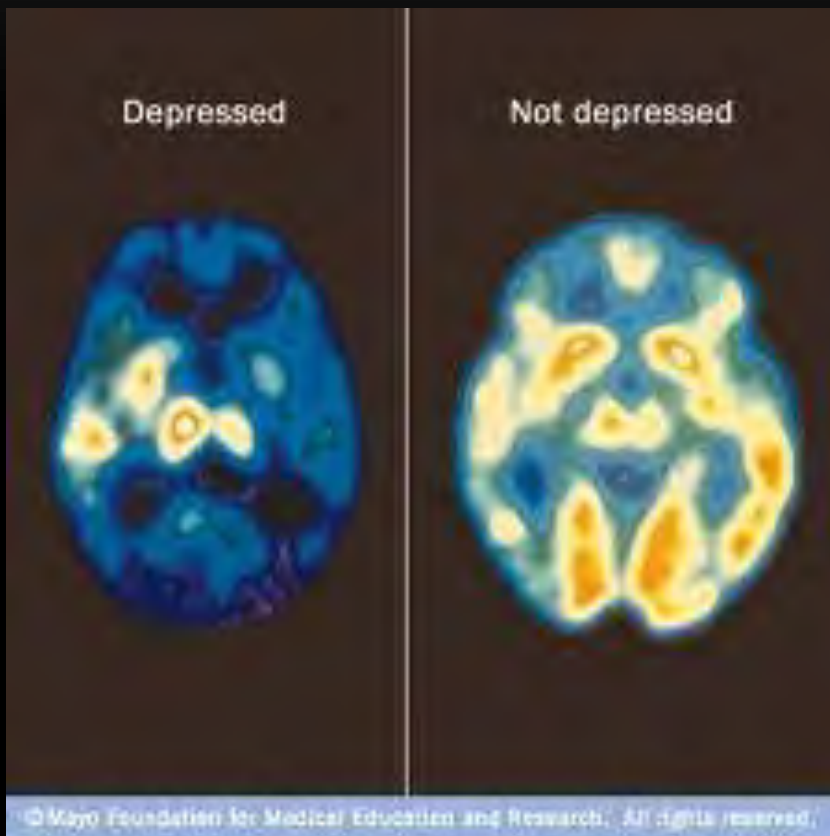
- ✓ Concept was introduced by David E. Kuhl, Luke Chapman and Roy Edwards in the late 1950s.
- ✓ Was further developed by Michel Ter-Pogossian, Michael E. Phelps and others.

POSITRON EMISSION TOMOGRAPHY (PET)



- ✓ Inject radioactive tracer, Fluorodeoxy-D-glucose (FDG), an analogue of glucose.
- ✓ Pairs of gamma rays emitted by the tracer were detected.
- ✓ The concentrations of tracer give tissue metabolic activity proportional to tissue glucose uptake.

POSITRON EMISSION TOMOGRAPHY (PET)

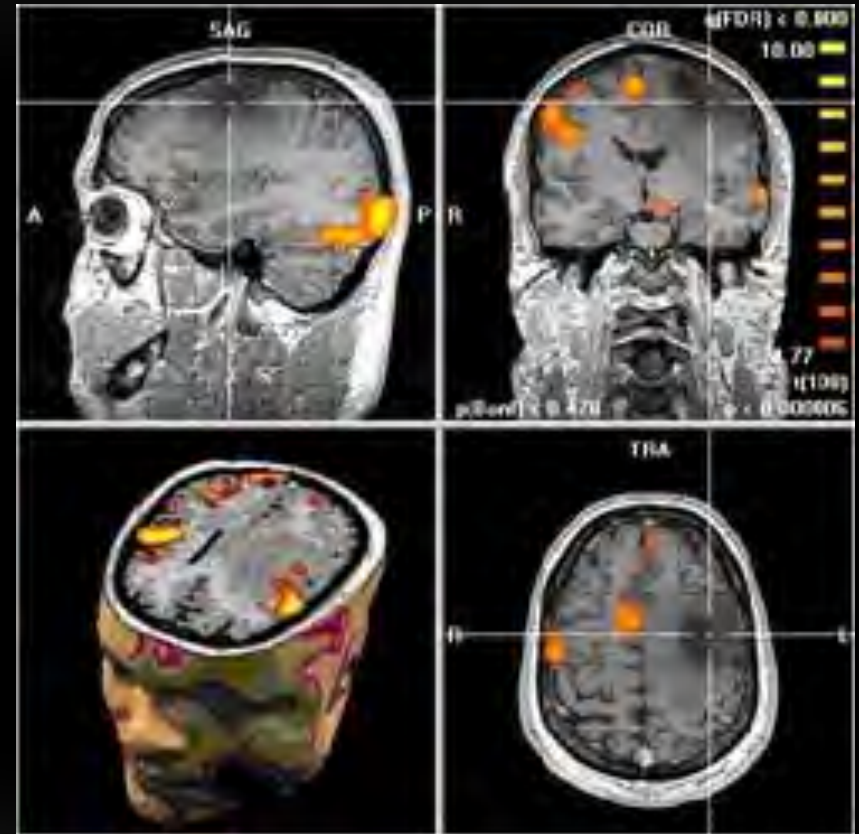
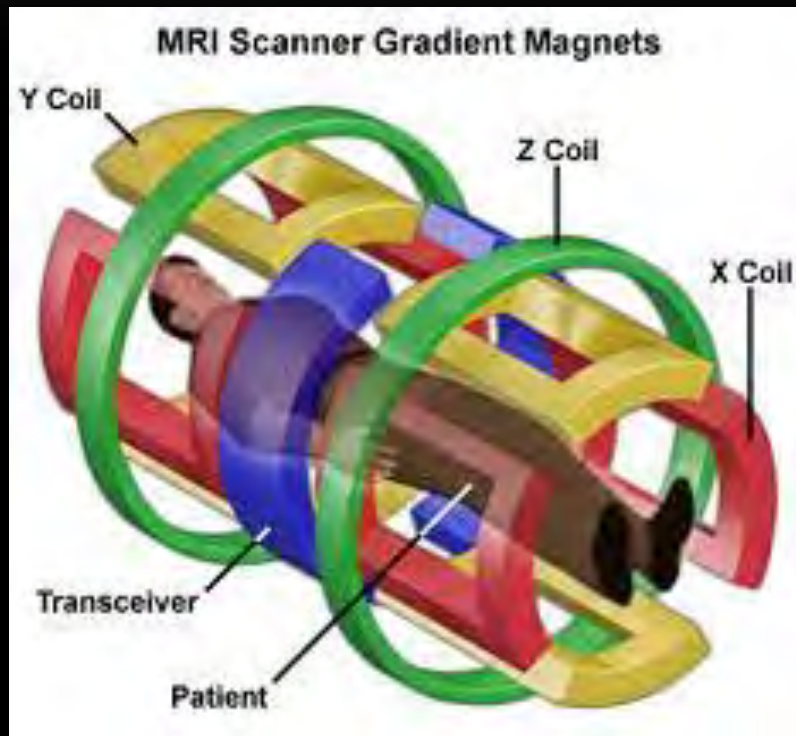


MAGNETIC RESONANCE IMAGING (MRI)

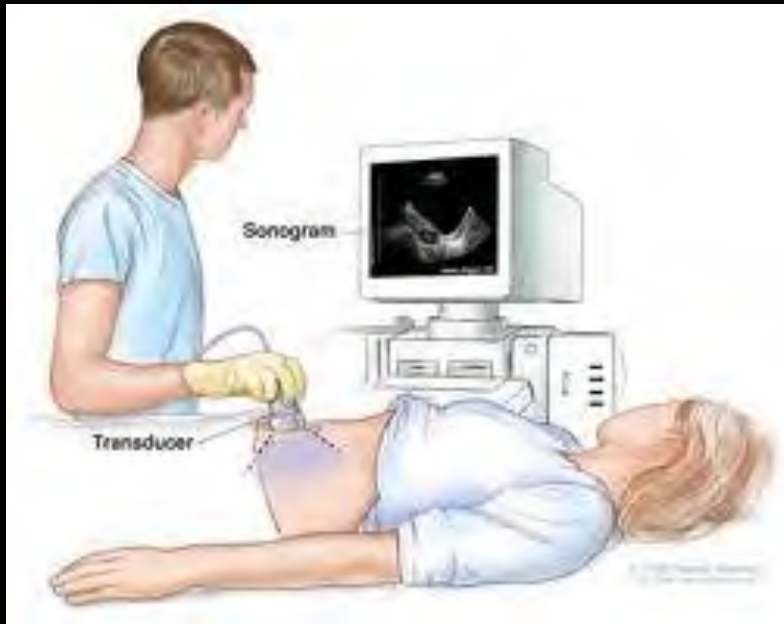


- ✓ Paul Lauterbur (University of Illinois) demonstrated first MRI image in living mouse in 1974.
- ✓ Peter Mansfield (University of Nottingham) demonstrated first MRI image in human in 1977.
- ✓ They won the Nobel Prize for Physiology or Medicine in 2003.

MAGNETIC RESONANCE IMAGING (MRI)



ULTRASONOGRAPHY (US)



- ✓ First applied to the human body by Dr. George Ludwig at the Naval Medical Research Institute in 1940s.
- ✓ Typically, 2 to 18 megahertz, though frequencies up to 50–100 megahertz have been used experimentally.

ULTRASONOGRAPHY (US)



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WHY OPTICAL IMAGING?

Characteristics	X-Ray	CT	PET	MRI	US	Optical Imaging
Ionizing Radiation	Yes	Yes	Yes	No	No	No
Spatial Resolution	mm-cm	mm-cm	cm	mm	100um - mm	Um to sub-um
Temporal Resolution	second	min	Tens of min	min	Sub-second	Sub-second
Contrast	Tissue density	Tissue density	Contrast agents	Tissue parametric al property	Tissue mechanical properties	Intrinsic contrast / contrast agents
Imaging Depth	Deep	Deep	Deep	Deep	Deep	Shallow
3D capability	No	Yes	Yes	Yes	Yes	Yes
Cost	\$	\$\$\$	\$\$\$	\$\$\$	\$\$	\$

Outline

•Introduction

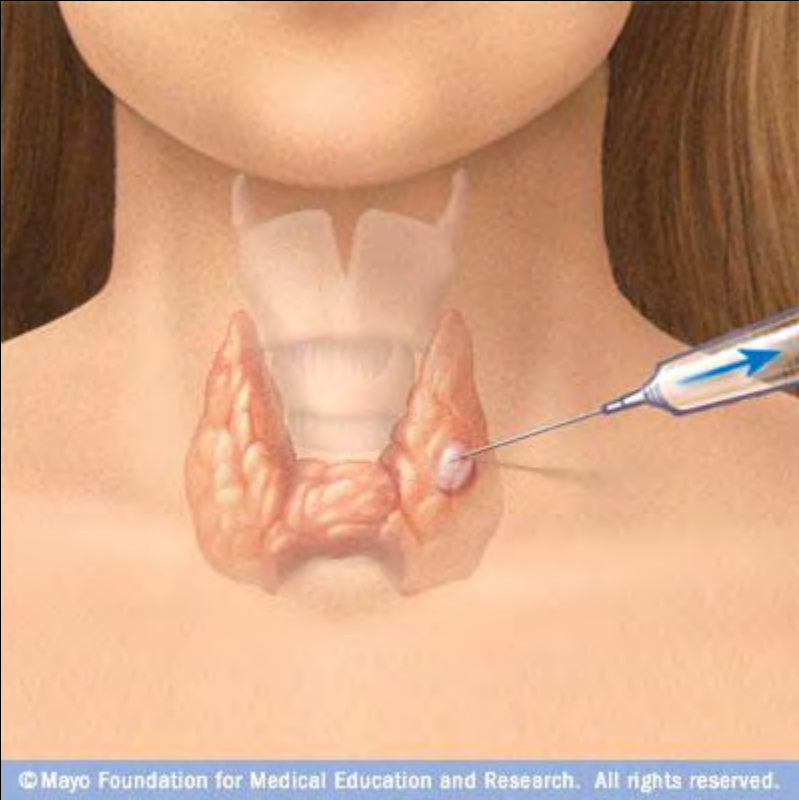
- Biomedical Imaging Modalities
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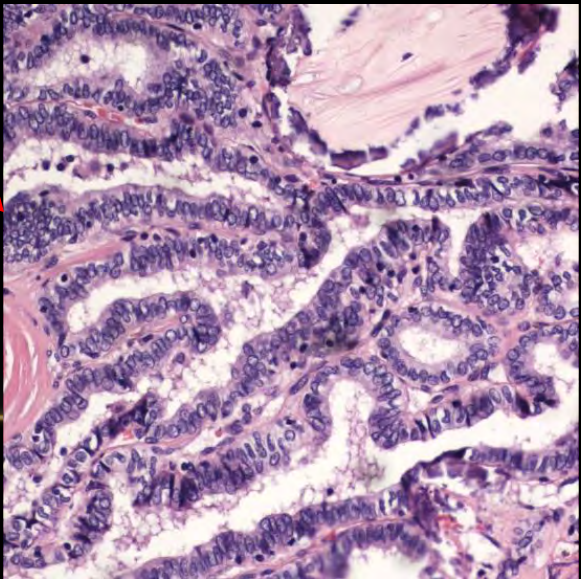
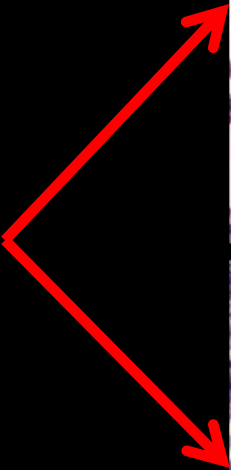
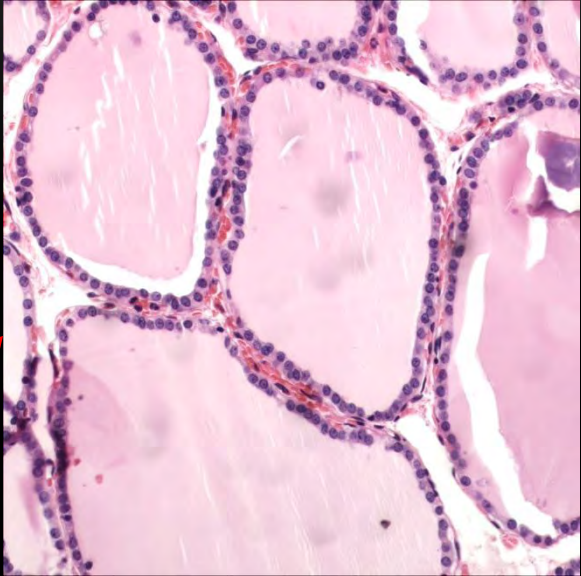
•Summary

BIOPSY



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Benign



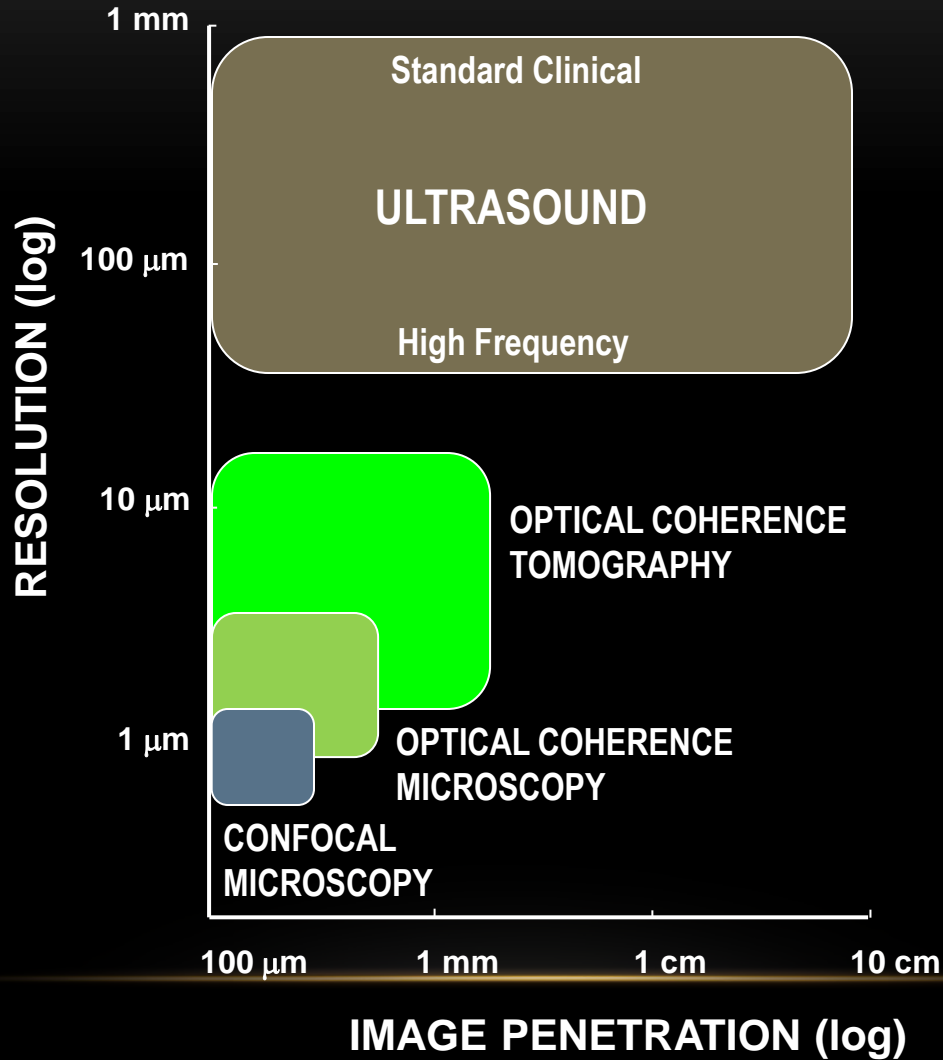
Several days to weeks!!!

Tumor

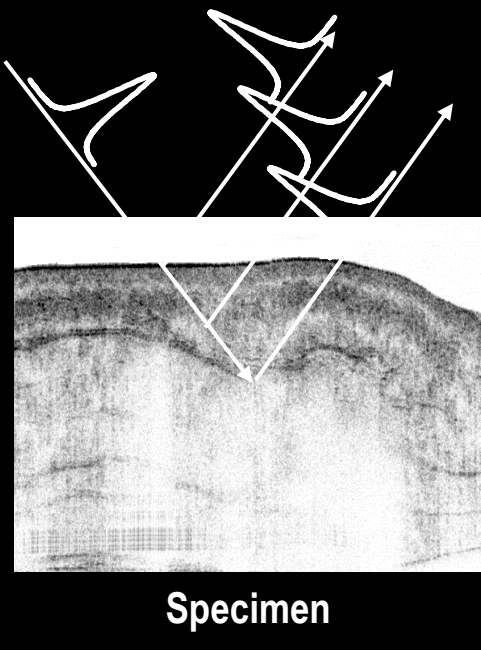
OBJECTIVE: OPTICAL BIOPSY

- ✓ *In situ, real-time* imaging of tissue microstructure with a resolution approaching that of histology, **without the need for tissue excision and processing.**
- ✓ Especially important in situations where excisional biopsy is either hazardous or impossible, *e.g.*, in ophthalmic or cardiovascular applications, neuroscience and developmental biology.

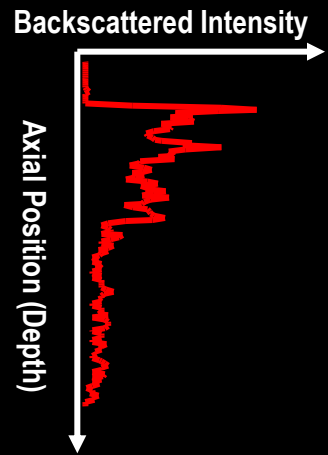
HIGH RESOLUTION SUBSURFACE IMAGING



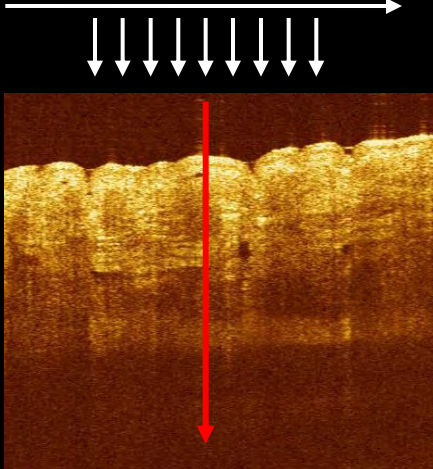
OPTICAL COHERENCE TOMOGRAPHY (OCT)



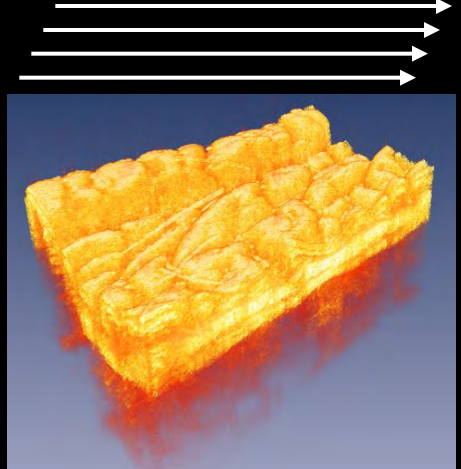
1 D
Axial (Z) Scanning



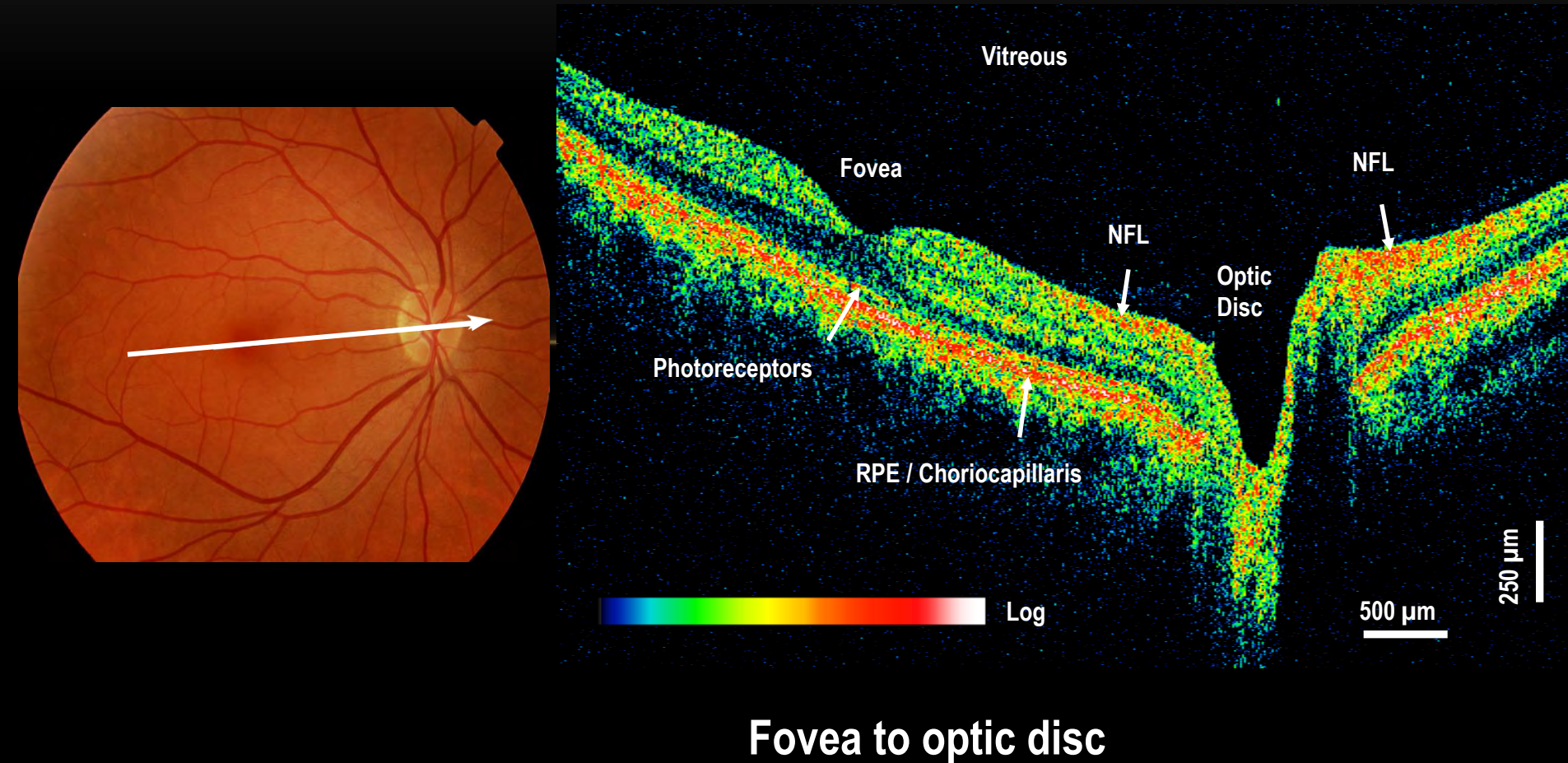
2 D
Axial (Z) Scanning
Transverse (X) Scanning



3 D
Axial (Z) Scanning
XY Scanning



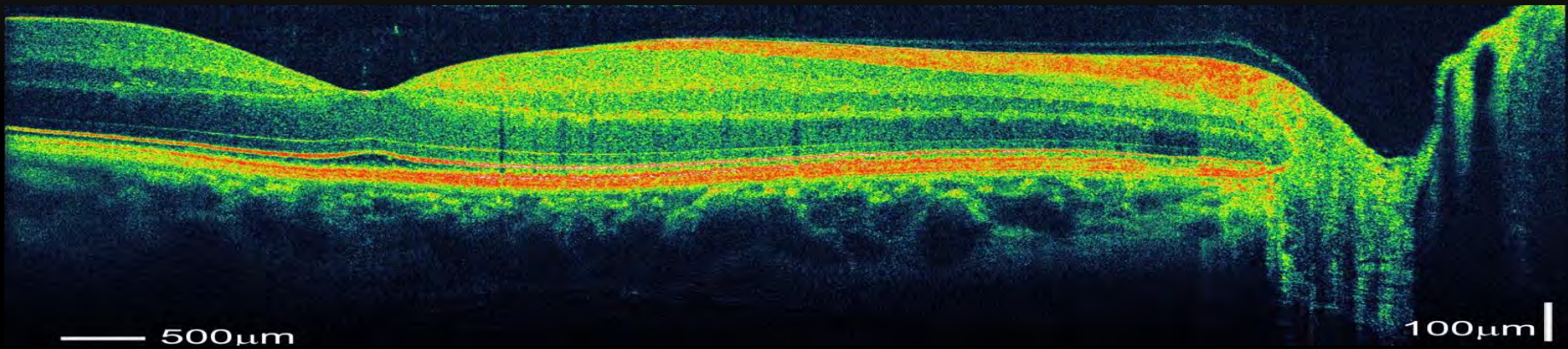
OCT IN OPHTHALMOLOGY



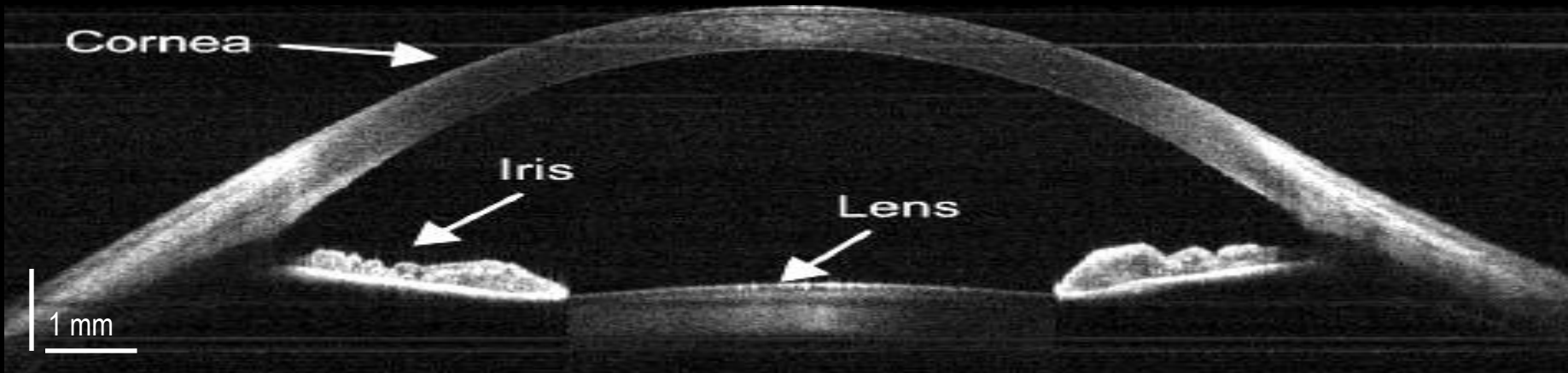
HIGH SPEED, ULTRAHIGH RESOLUTION OCT

(250,000 – 400,000 A-lines/s)

High Speed, Ultrahigh Resolution OCT of Human Retina



High Speed OCT of Human Anterior Segment



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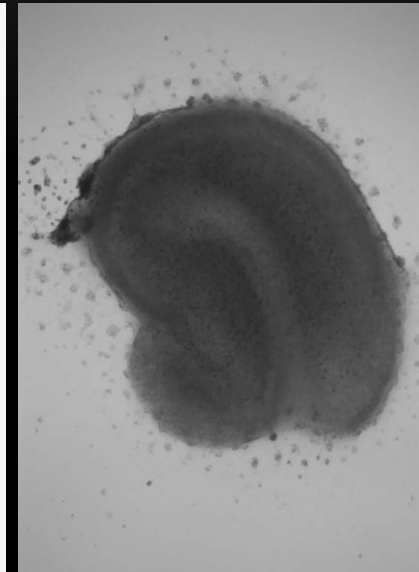
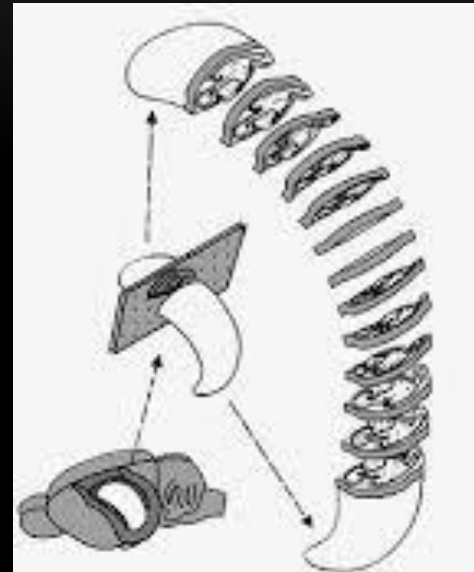
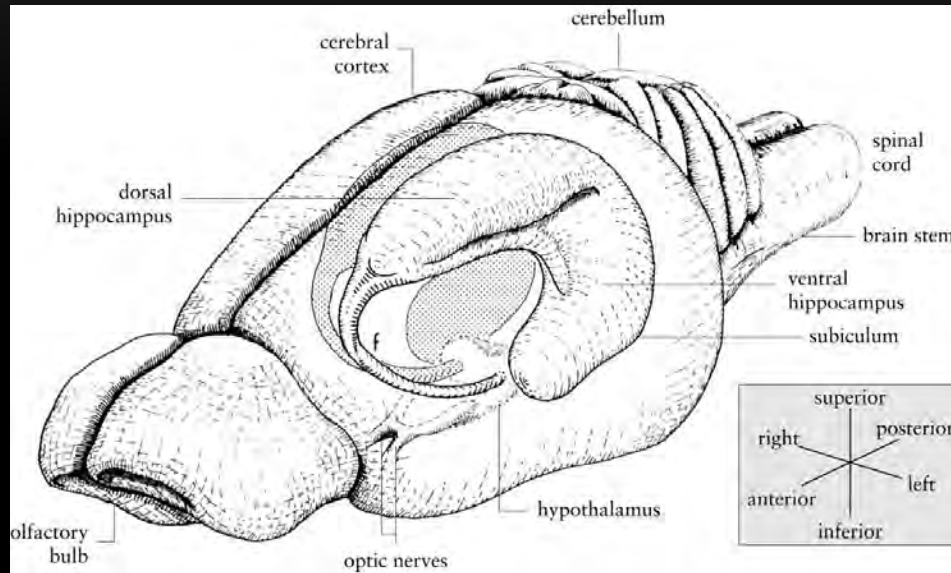
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Organotypic Hippocampal Slice Cultures

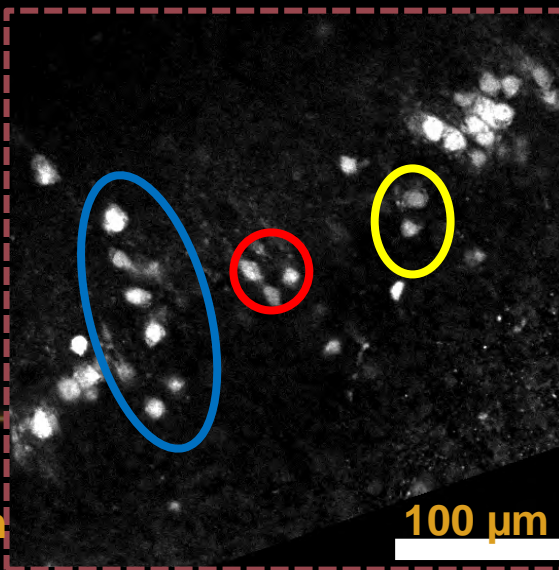
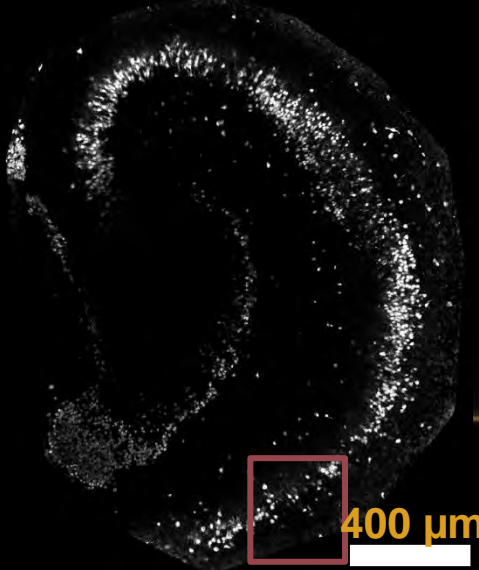
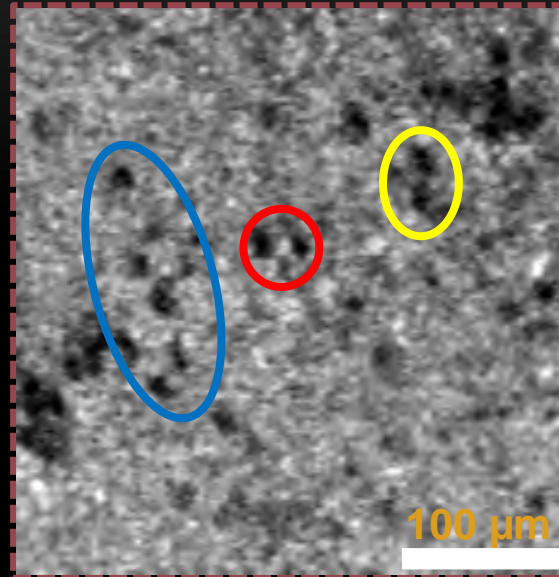
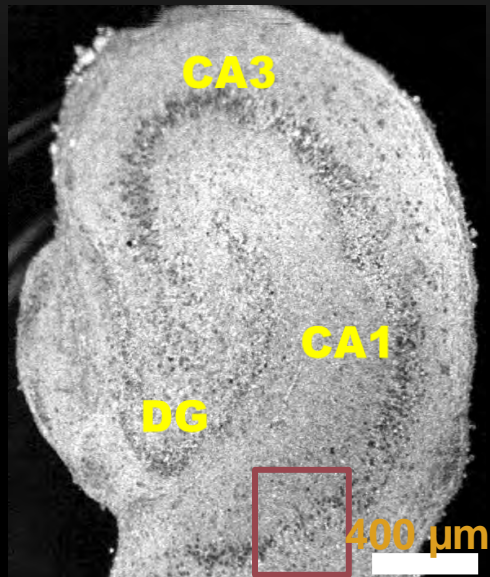


Cheung and Cardinal *BMC Neuroscience* 2005

Characteristics:

- Hippocampus of **7-day old Sprague-Dawley rats**
- ~300 μ m thick**

Comparison of OCM and Confocal Images

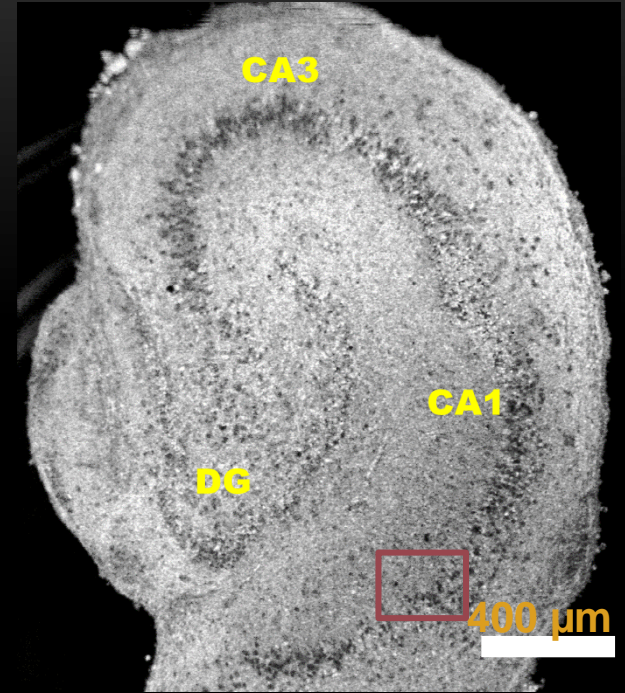


Nuclei: anti-NeuN

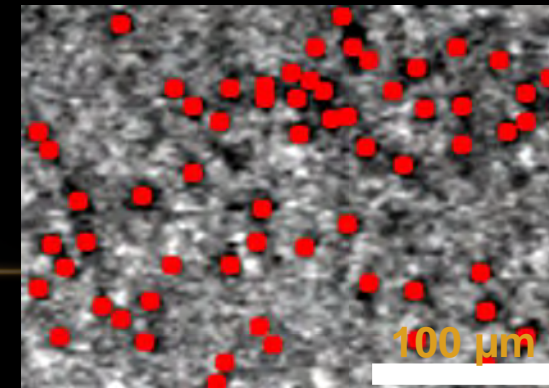
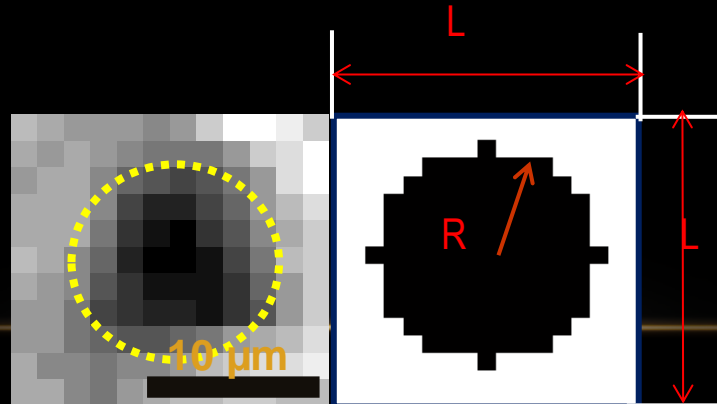
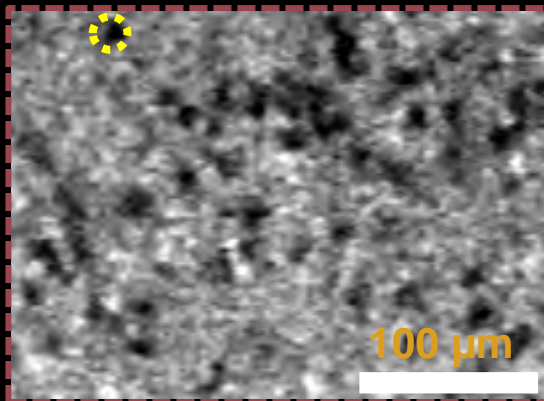
Quantify Neurons in 3D

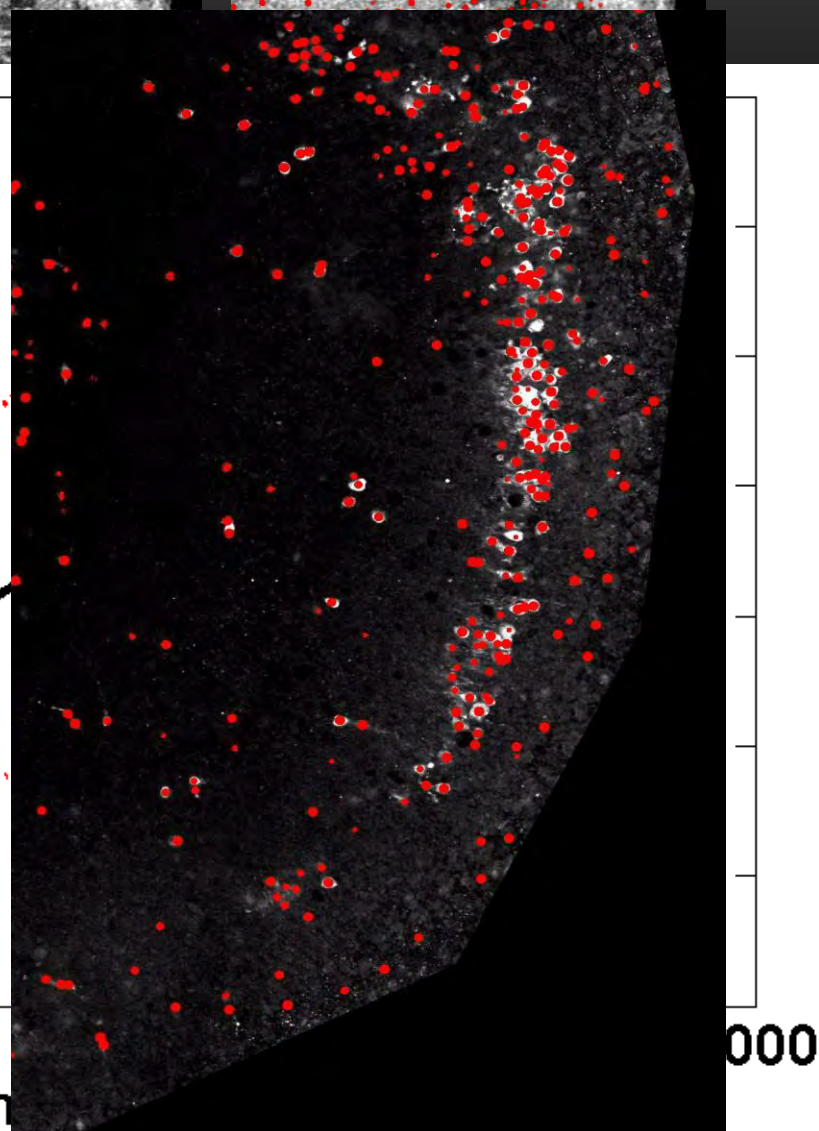
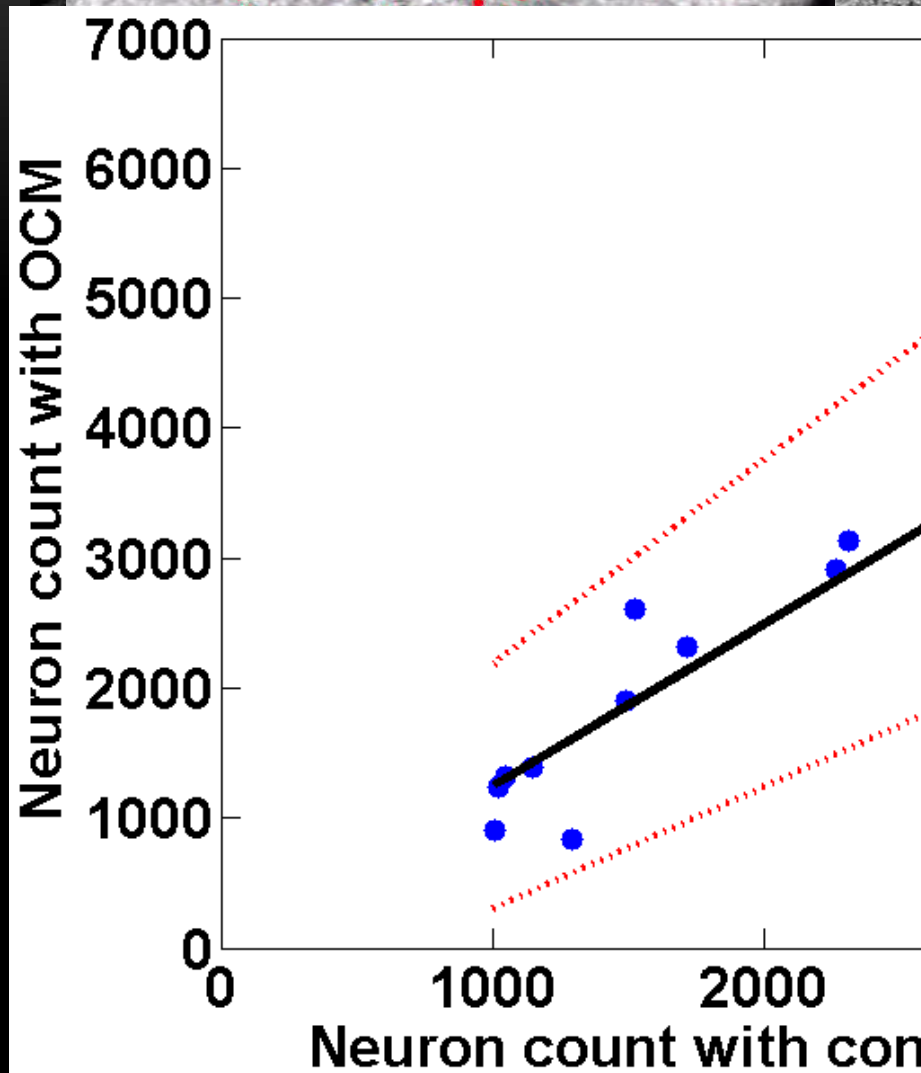


CA3: Cornuammonis III
CA1: Cornuammonis I
DG: Dentate gyrus



Chu *et al*, Journal of Molecular Histology, 2007



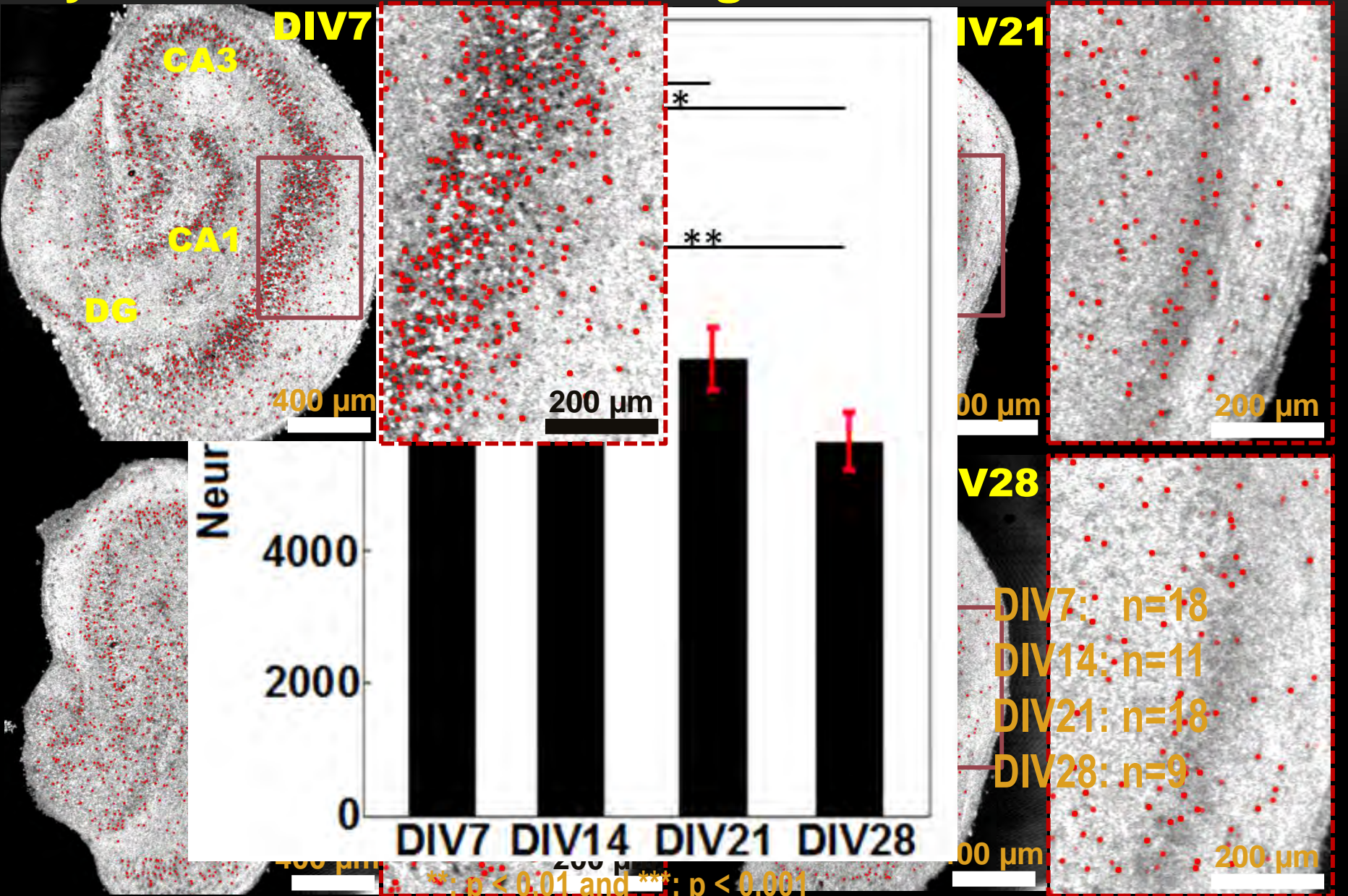


400 μm

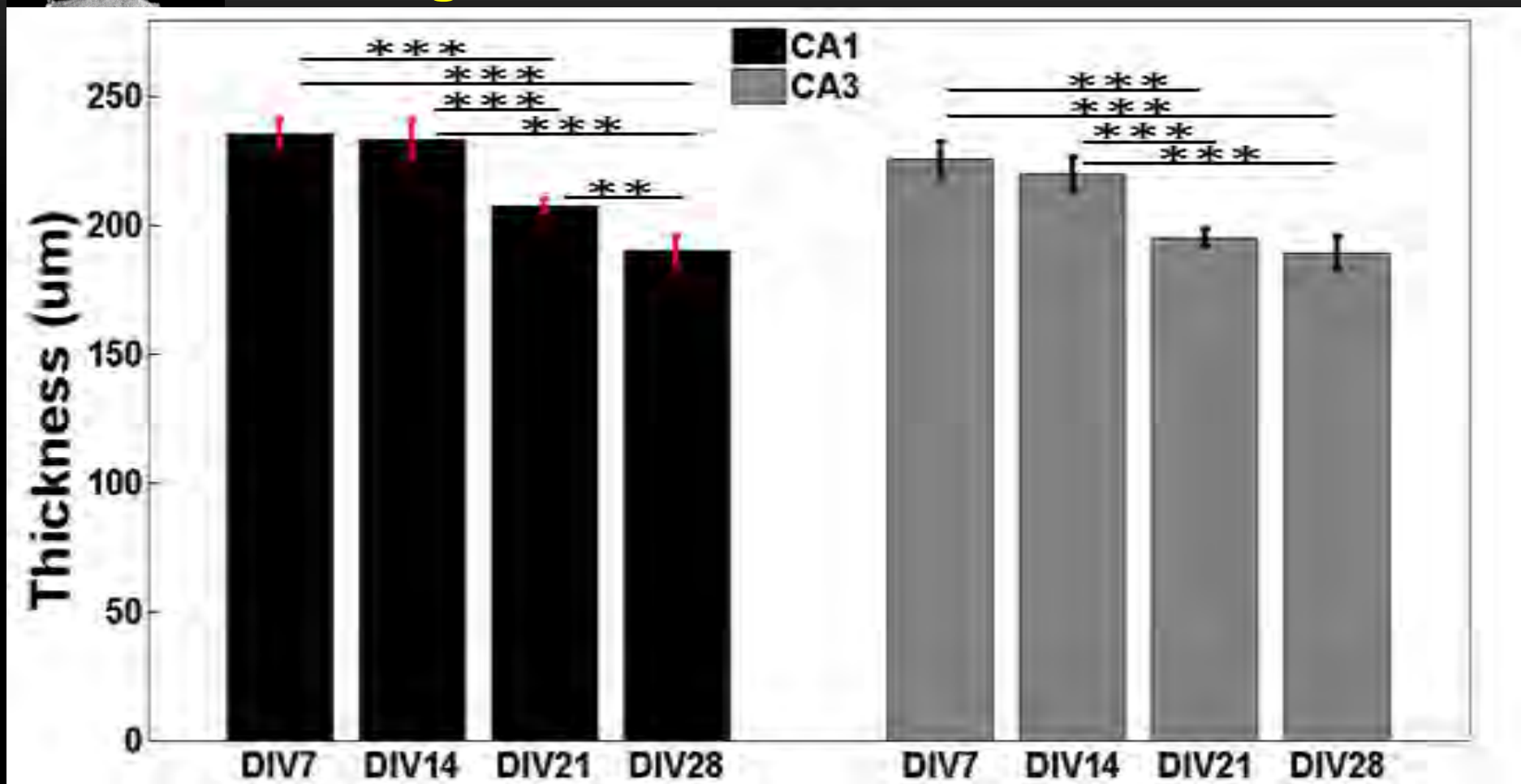
200 μm

200 μm

Evaluation of seizures-induced neuronal injury as days *in vitro* increased using OCM



Slice thickness measurement as days *in vitro* increased using OCM



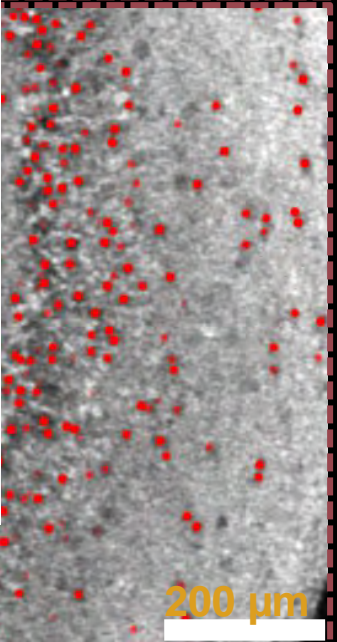
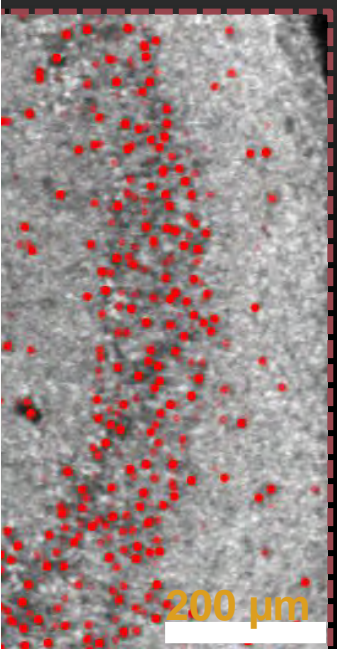
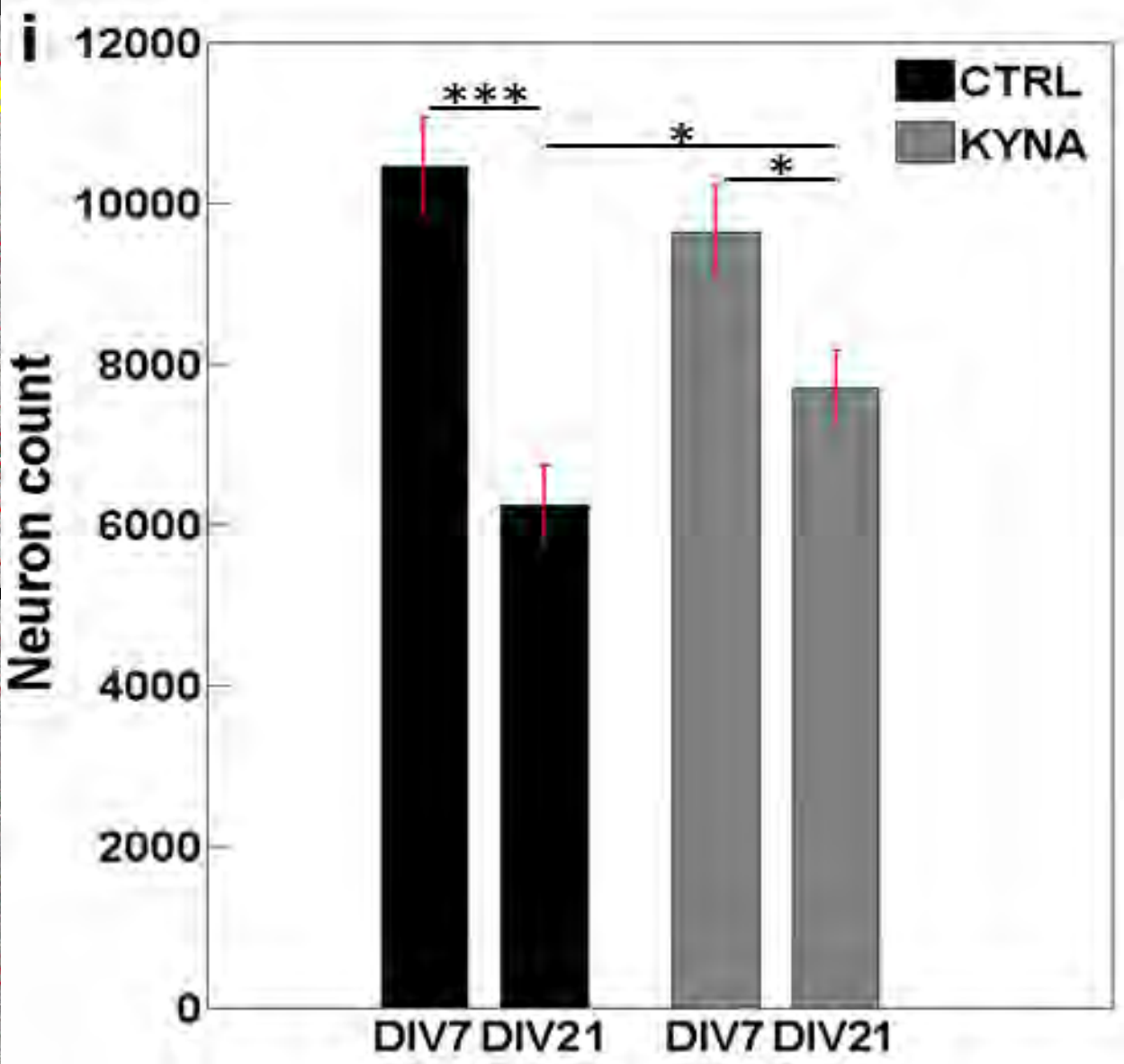
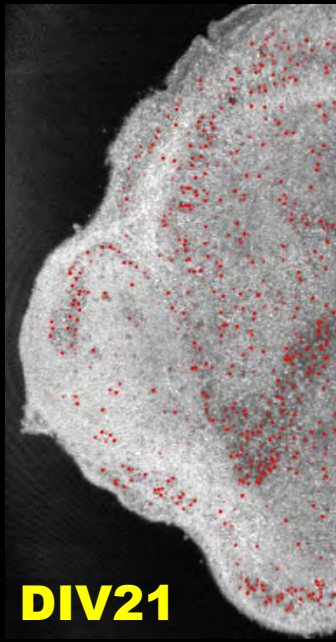
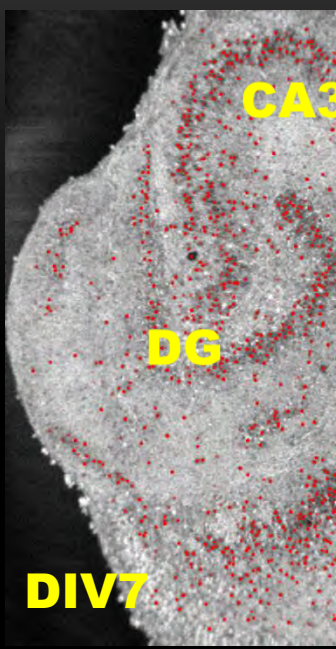
******: $p < 0.01$ and *******: $p < 0.001$

DIV7: n=18; DIV14: n=11; DIV21: n=18; DIV28: n=9

The cross-sectional images are from CA3 region

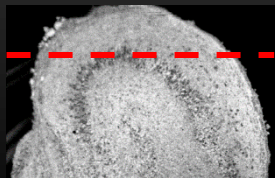
200µm

Neuroprotective Effects of KYNA



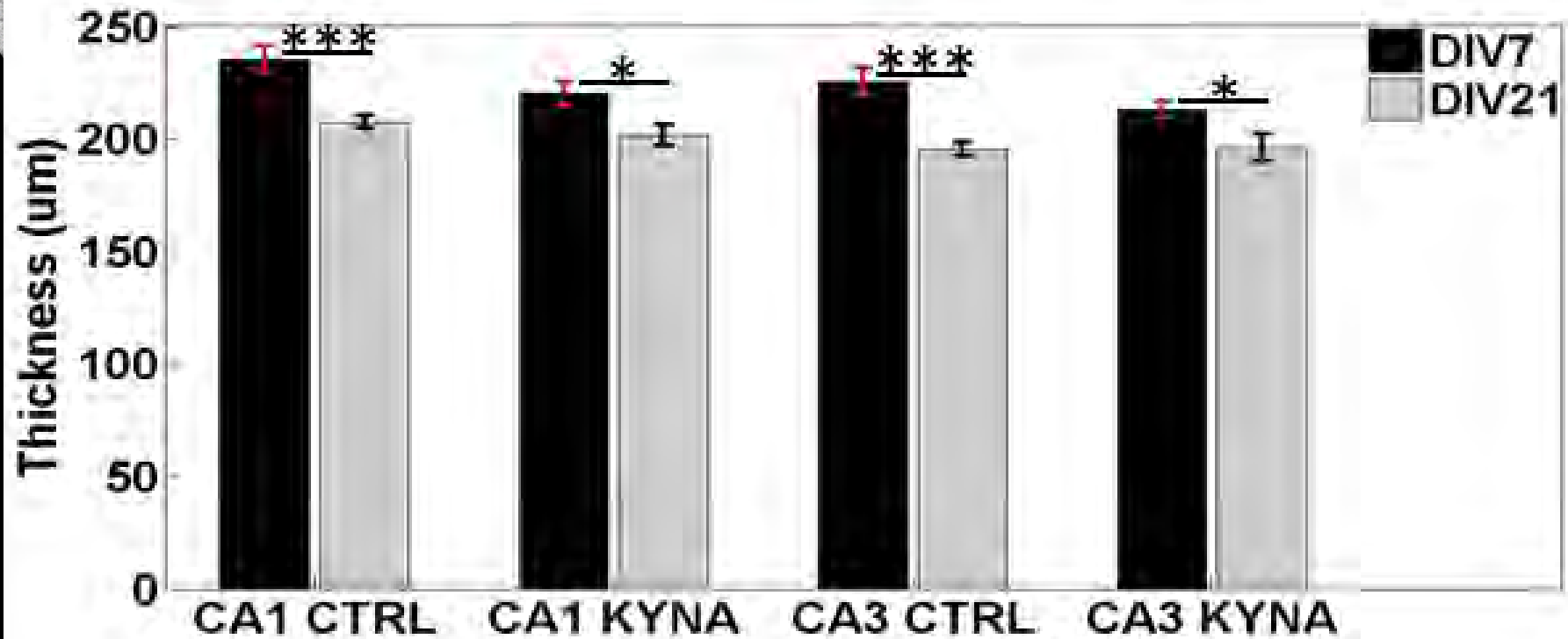
*: $p < 0.05$ and ***: $p < 0.001$
n = 9 in each group

Slice thickness in control and KYNA group



DIV7

CTRL



*: $p < 0.05$ and ***: $p < 0.001$

200 μm

$n = 9$ in each group
The cross-sectional images are from CA3 region

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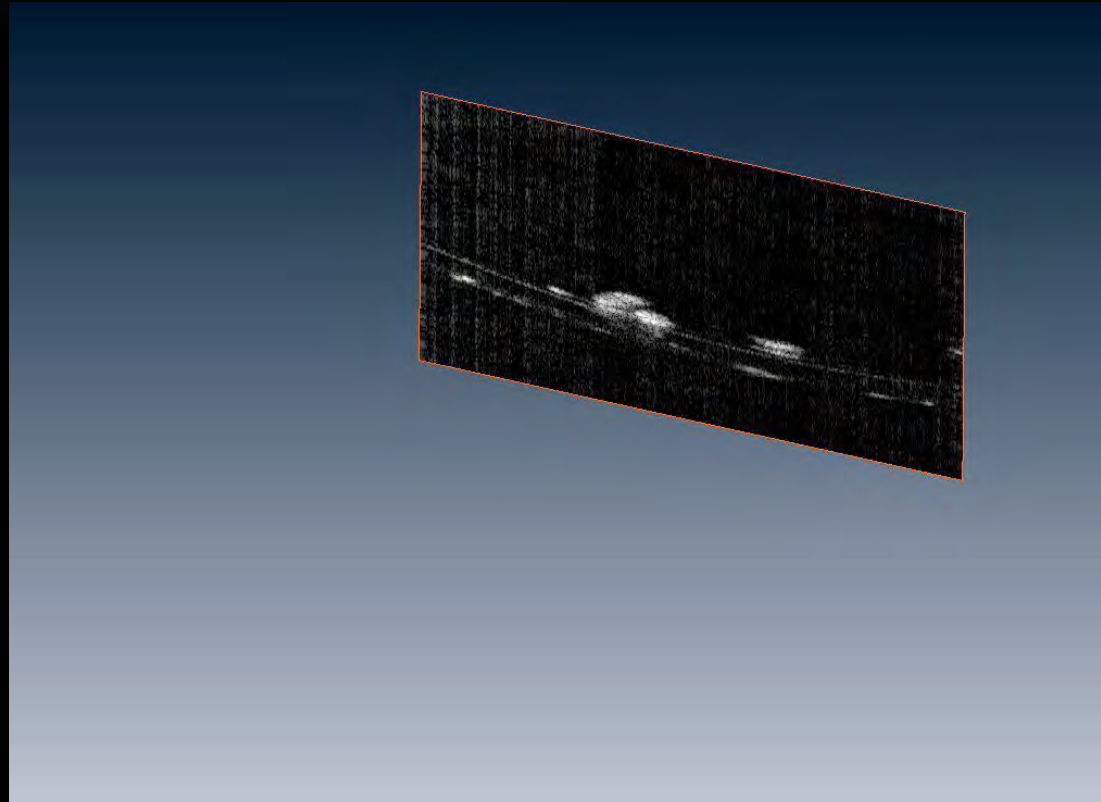
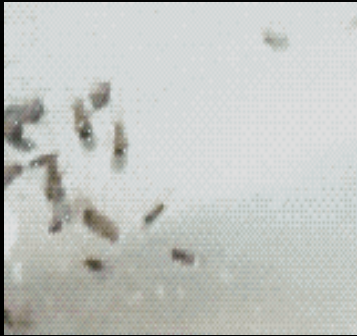
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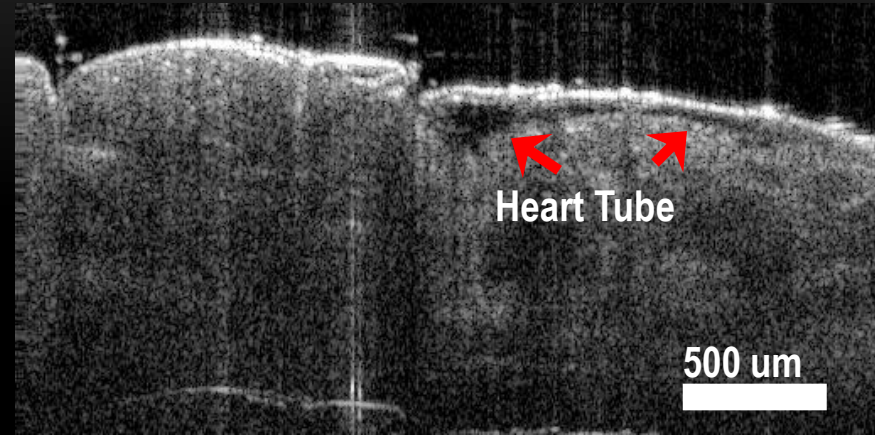
•Summary

OCT IMAGING OF THE DROSOPHILA HEART

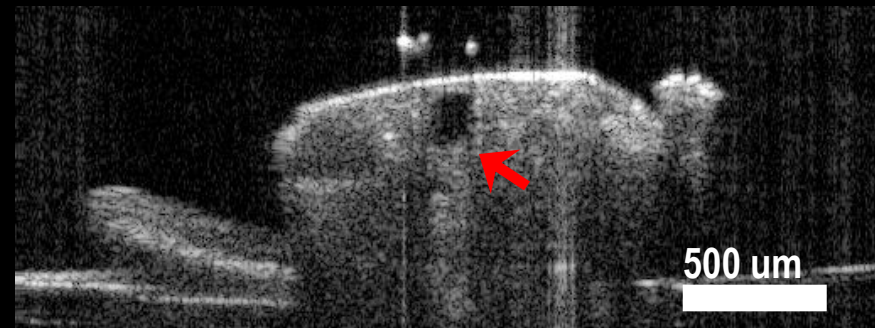


OCT IMAGING OF THE DROSOPHILA HEART

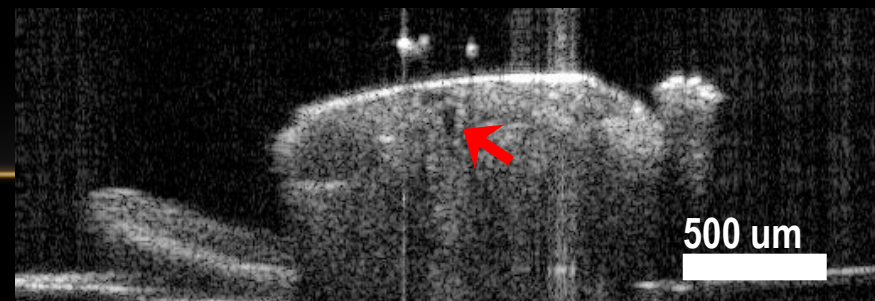
Longitudinal View



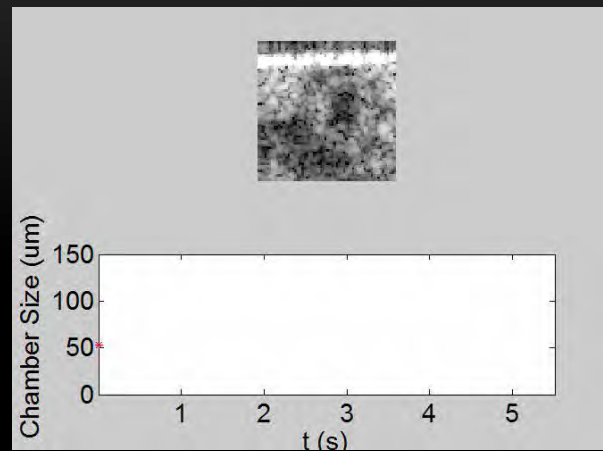
**Cross-sectional View
- Diastolic Phase**



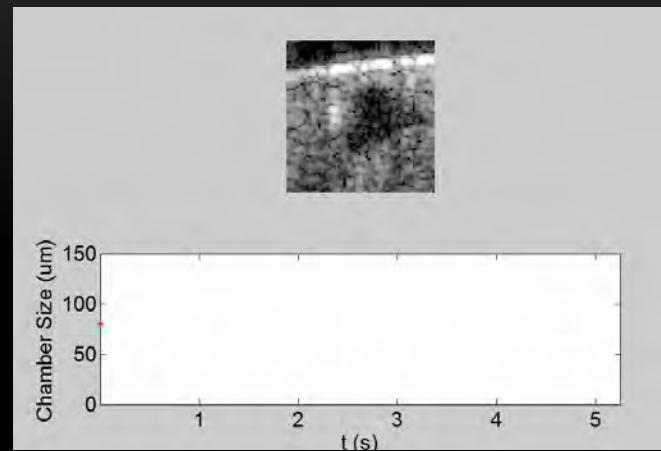
**Cross-sectional View
- Systolic Phase**



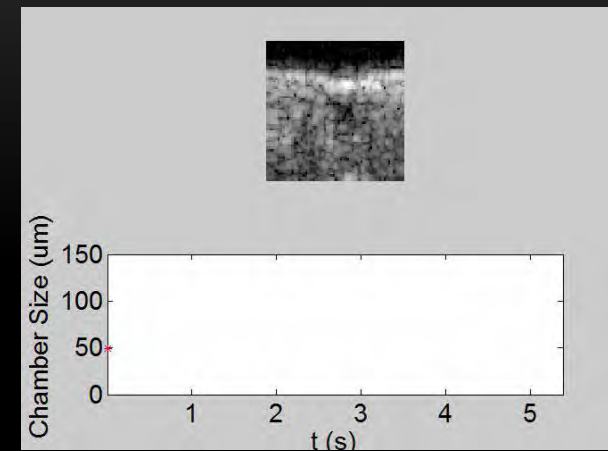
OCT Imaging of the Drosophila Heart



**24B-GAL4/+
(Control)**



**UAS-dPsn; 24B-GAL4
(Over-expression of *dPsn*)**



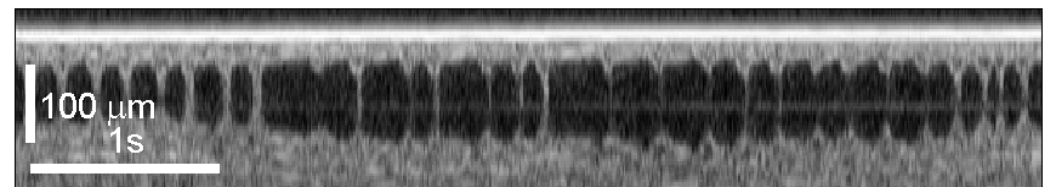
**UAS-dPsnRNAi; 24B-GAL4
(Silencing of *dPsn*)**

M-Mode Imaging

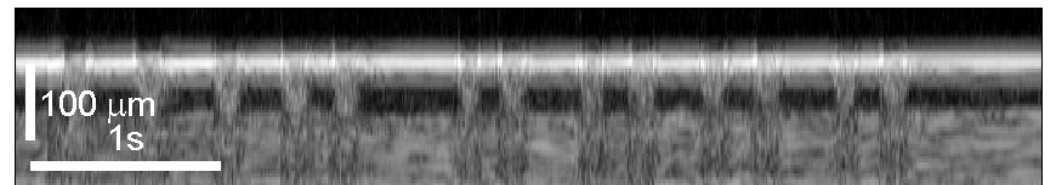
Control



**Over-expression
of *dPsn***



Silencing of *dPsn*



GROUP RESULTS

Parameters	7 Day old				30 Day old			
	24B-GAL4/+ (Control) N=31	UAS-dPsn; 24B-GAL4 N=31	UAS-dPsn ^{RNAi} ; 24B-GAL4 N=24	Total N=86	24B-GAL4/+ (Control) N=30	UAS-dPsn; 24B-GAL4 N=28	UAS-dPsn ^{RNAi} ; 24B-GAL4 N=28	Total N=86
HR (BPM)	262 ± 10	307 ± 11 *** ↑	231 ± 11 * ↓	269 ± 7	254 ± 10	284 ± 11 * ↑	190 ± 8 ****## ↓	243 ± 7 ## ↓
ESD (μm)	20 ± 2	14 ± 2	17 ± 2	17 ± 1	31 ± 4 ## ↑	20 ± 4 * ↓	14 ± 3 **** ↓	22 ± 2 # ↑
EDD (μm)	67 ± 2	56 ± 3 * ↓	52 ± 4 ** ↓	59 ± 2	78 ± 4 # ↑	66 ± 4 * ↓ # ↑	73 ± 4 ##### ↑	73 ± 2 ##### ↑
FS (%)	69 ± 4	76 ± 3	67 ± 4	71 ± 2	62 ± 4	69 ± 5	83 ± 4 *** ↑ ## ↑	71 ± 2

HR: Heart rate; EDD: End-diastolic dimension; ESD: End-systolic dimension; FS: Fractional shortening

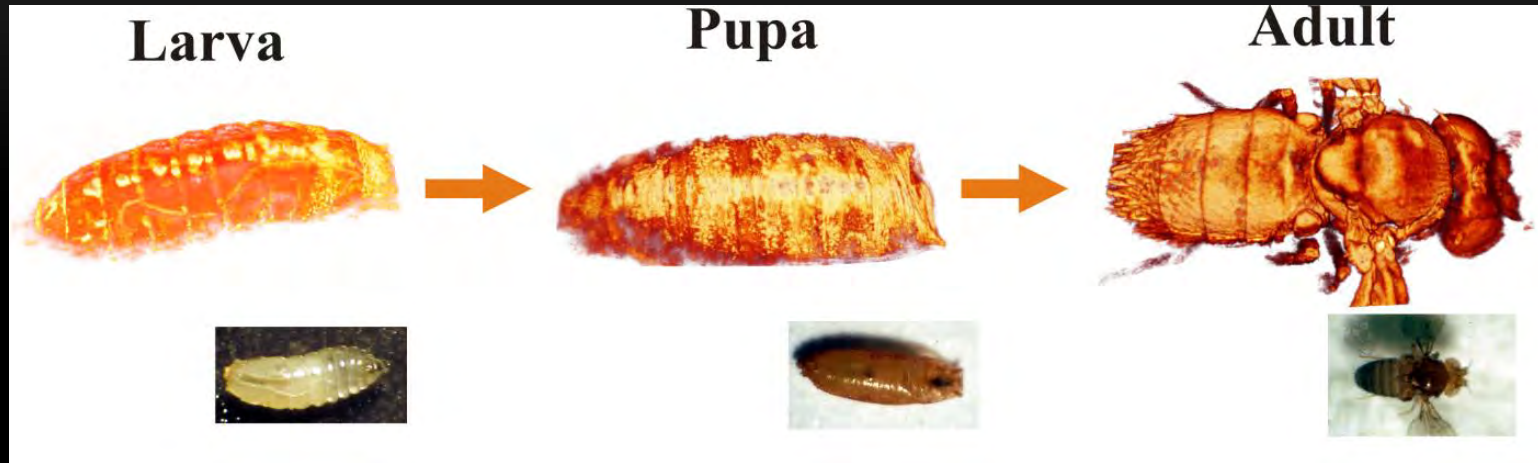
*p<0.05, ** p<0.01, *** p<0.001, **** p<0.0001: vs. Age-matched controls;

#p<0.05, ## p<0.01, ### p<0.001, #### p<0.0001: vs. 7-day old age group

↑ shows significant increase; ↓ shows significant decrease

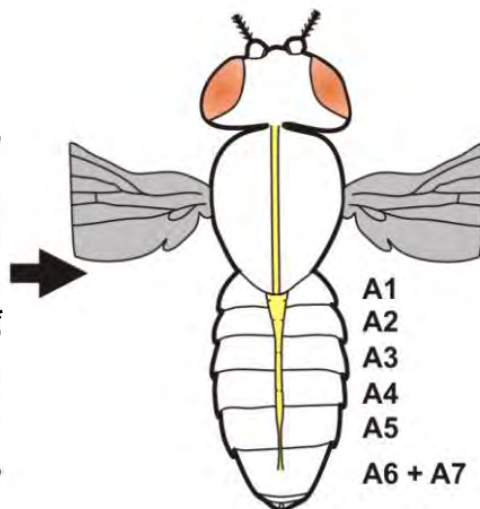
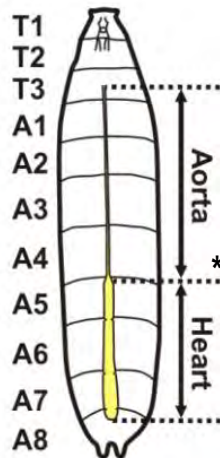
Li, Zhou, *et al.*, *Curr. Alzheimer Res.*, 8(3):313-322, 2011

DROSOPHILA HEART DEVELOPMENT

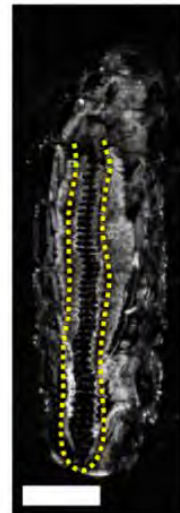


Heart metamorphosis

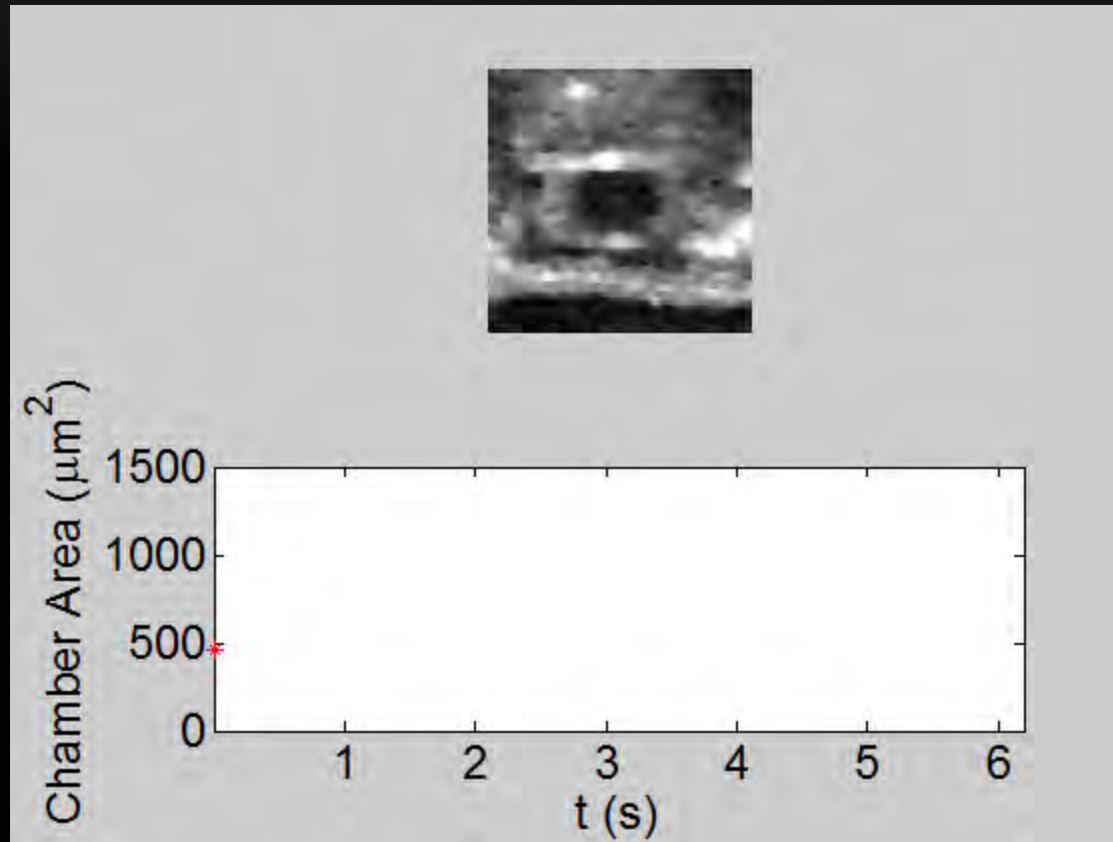
B



C

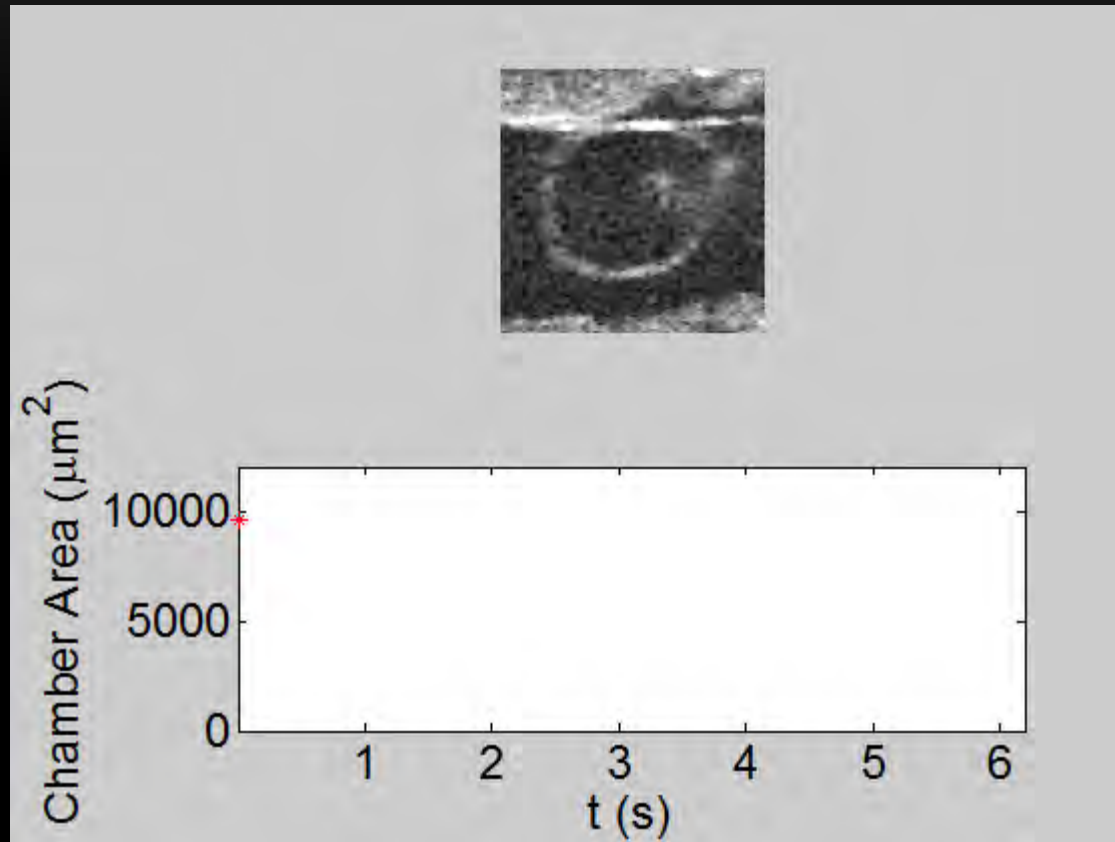


DROSOPHILA HEART DEVELOPMENT



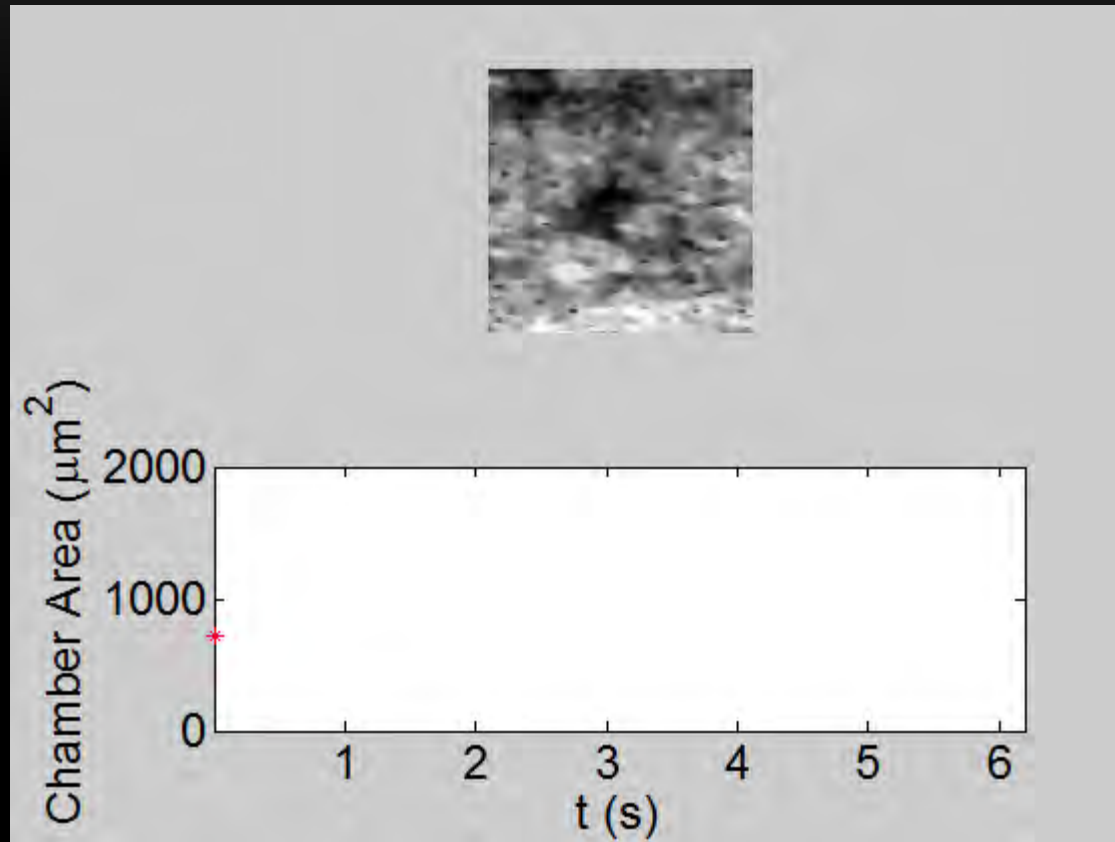
2nd instar larva – L2

DROSOPHILA HEART DEVELOPMENT



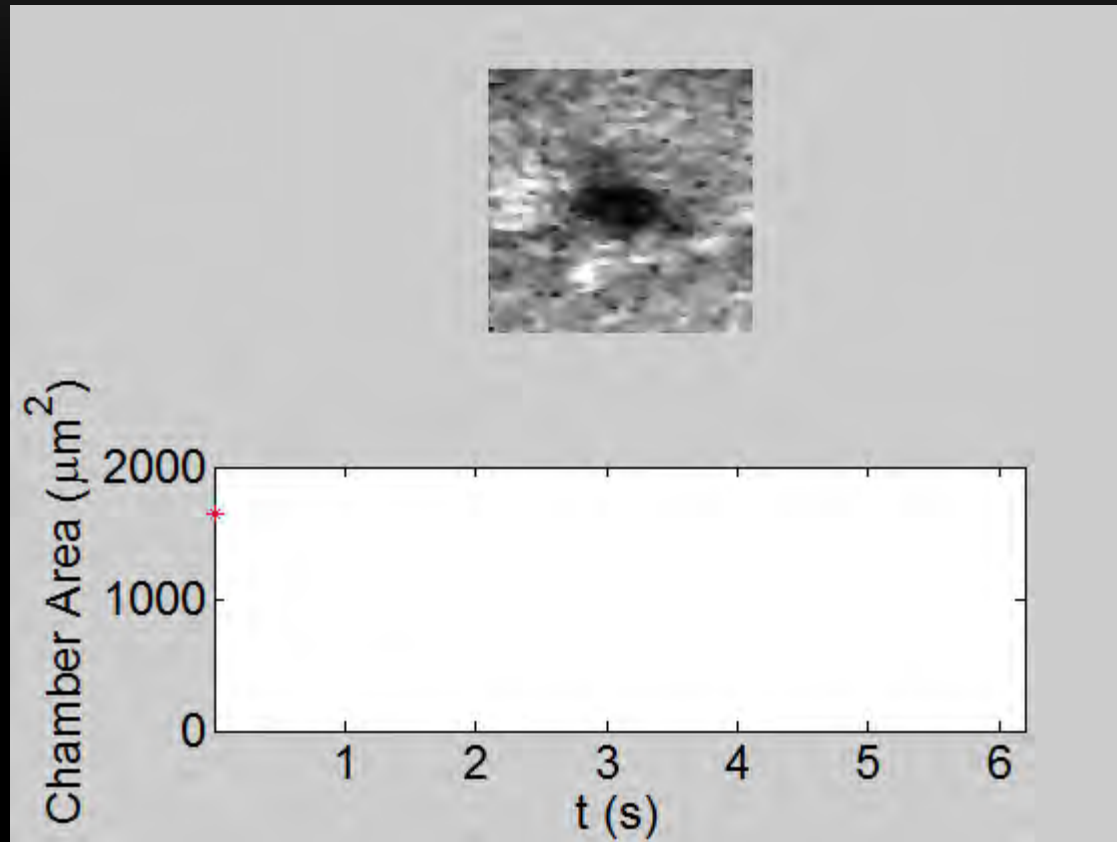
3rd instar larva – L3

DROSOPHILA HEART DEVELOPMENT



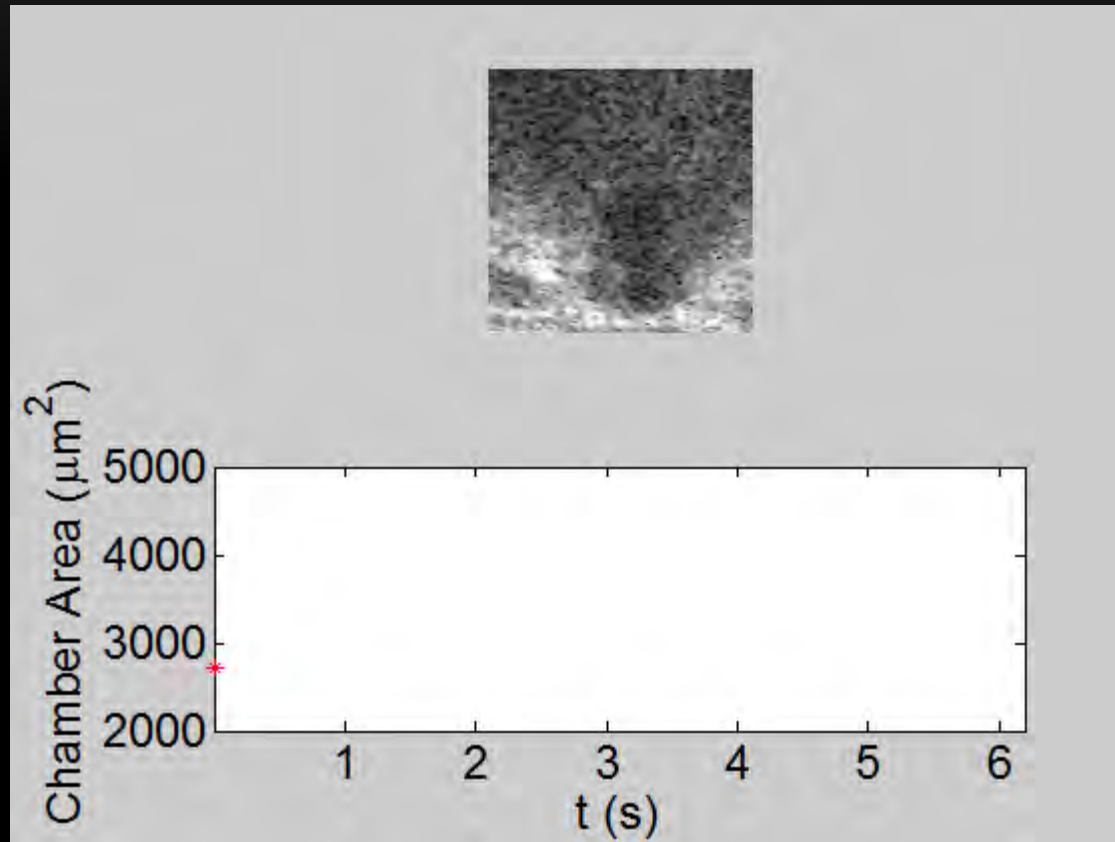
Pupa day 1, 8hr – PD1

DROSOPHILA HEART DEVELOPMENT



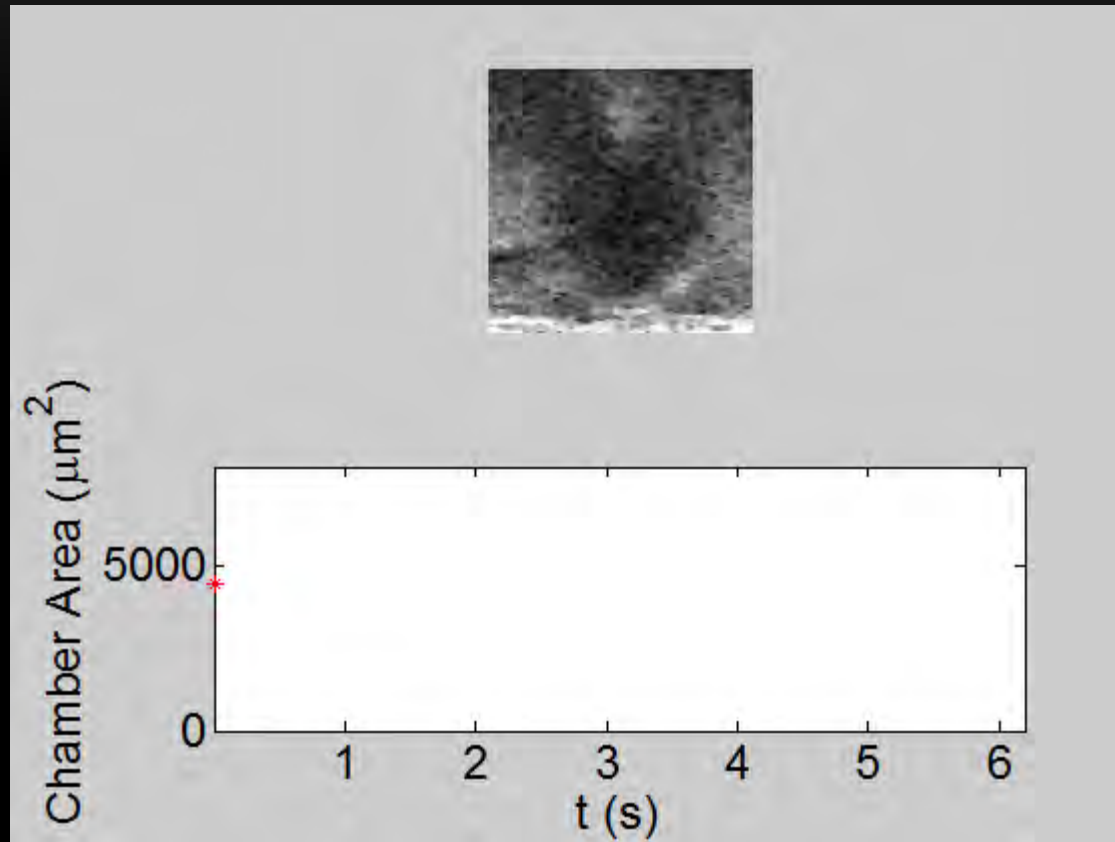
Pupa day 2, 32hr – PD2

DROSOPHILA HEART DEVELOPMENT



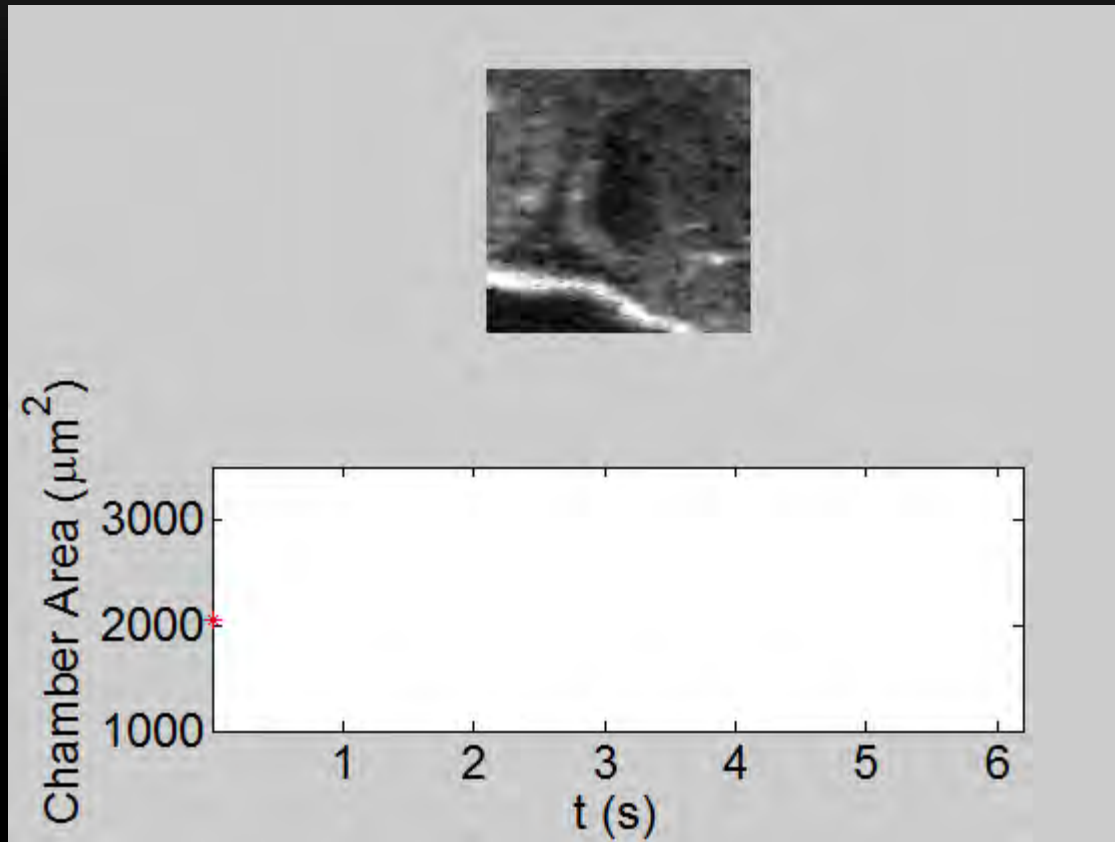
Pupa day 3, 72hr – PD3

DROSOPHILA HEART DEVELOPMENT



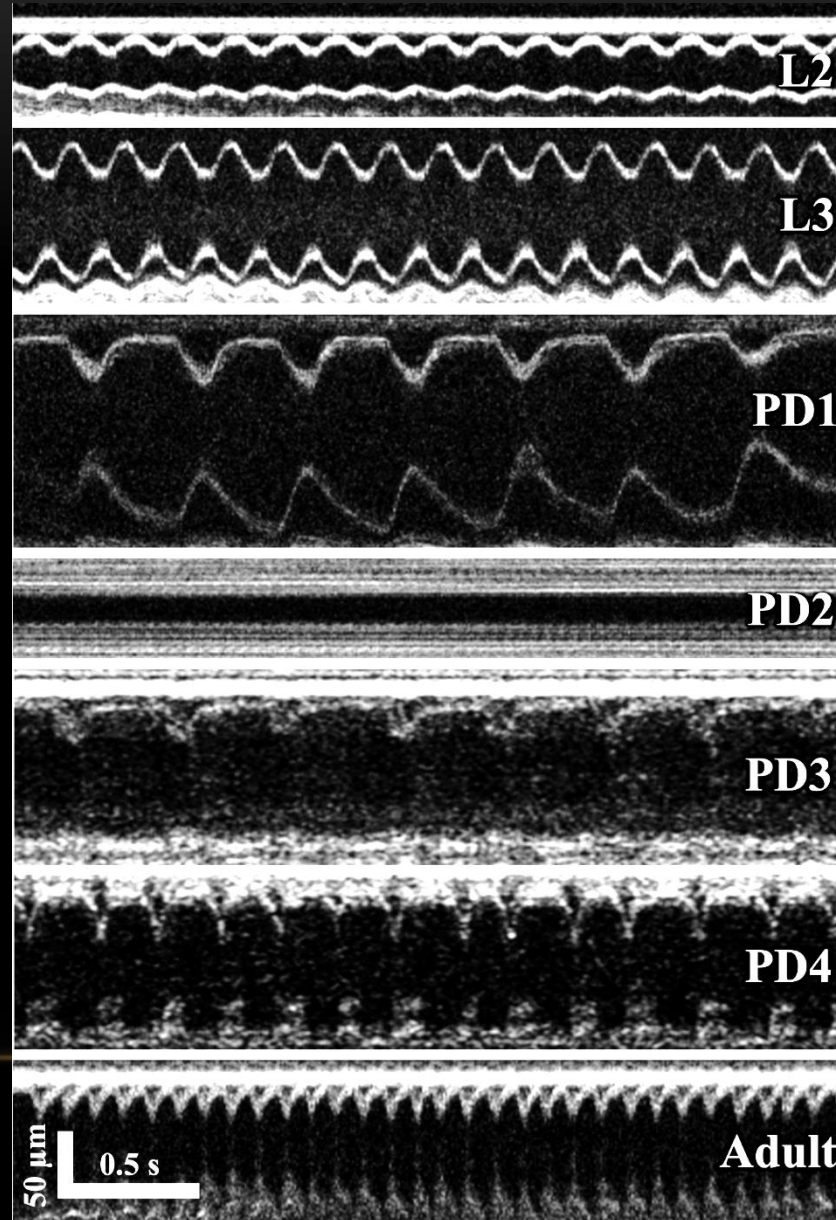
Pupa day 4, 88hr – PD4

DROSOPHILA HEART DEVELOPMENT

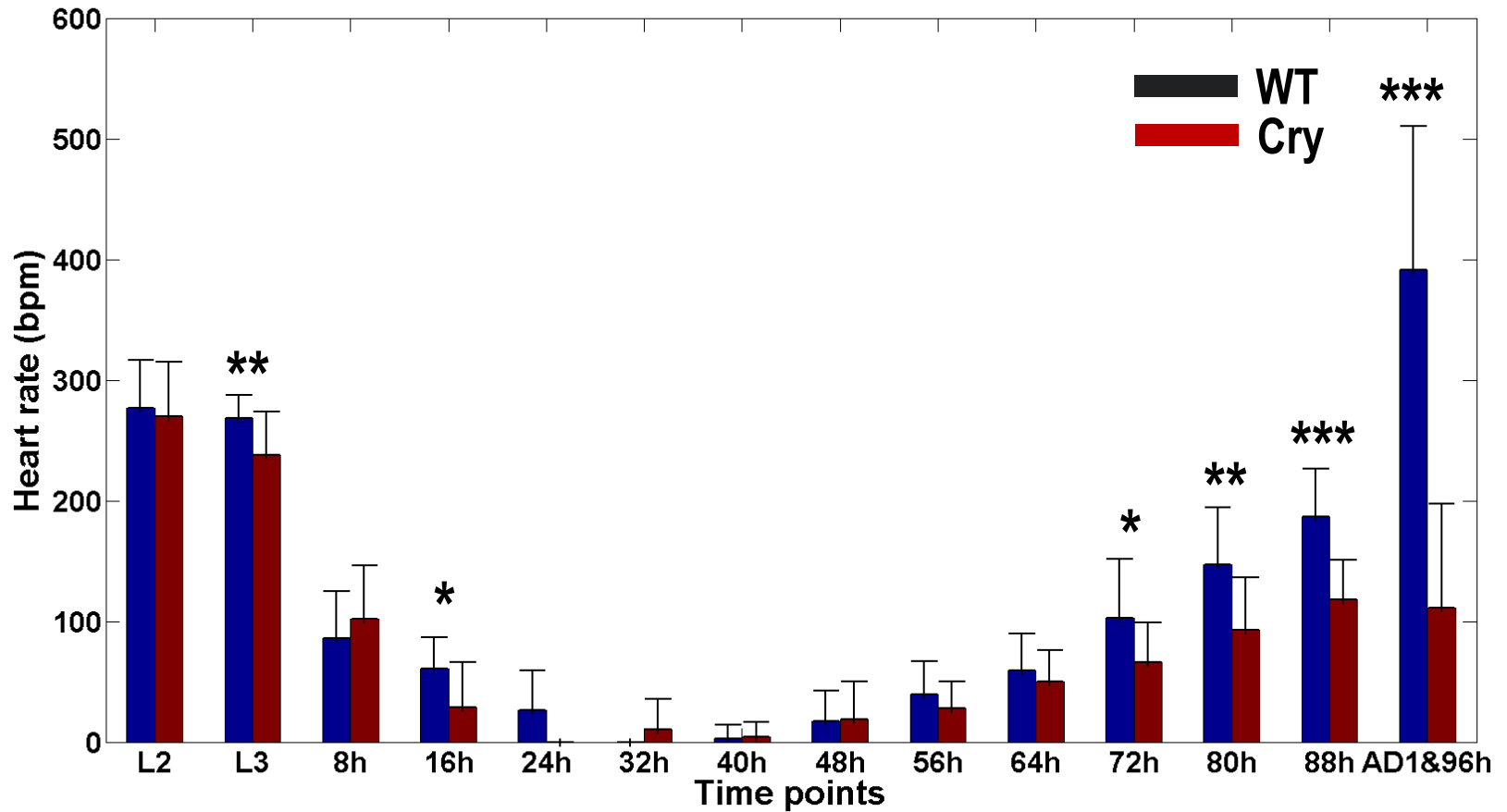


Adult day 1

DROSOPHILA HEART DEVELOPMENT



DROSOPHILA HEART RATE



Outline

•Introduction

- Biomedical Imaging Modalities
- Why Optical Imaging?
- Optical Biopsy with Optical Coherence Tomography (OCT) and Microscopy (OCM)

•Applications in Neuroscience and Developmental Biology

- 3D imaging of brain slices
- Evaluate heart function in fruit flies

•Summary

Summary

- Various imaging modalities can be used for clinical and research applications
 - Optical imaging provides unique advantages (resolution, contrast, etc.)
 - Optical biopsy can be achieved by OCT and OCM
 - None-invasive evaluation of epilepsy models in rat brain slices
 - None-invasive characterization of heart function in fruit flies
-

ECE 368/468, BioE 368/468
Introduction to Biophotonics /
Optical Biomedical Imaging

Spring, 2015

Thank you!